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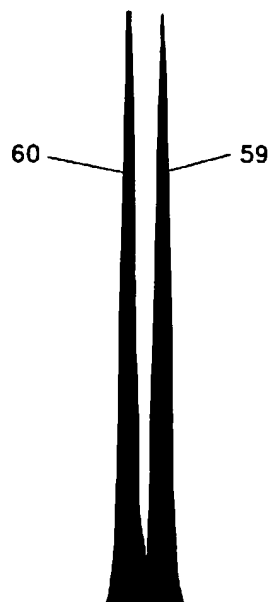
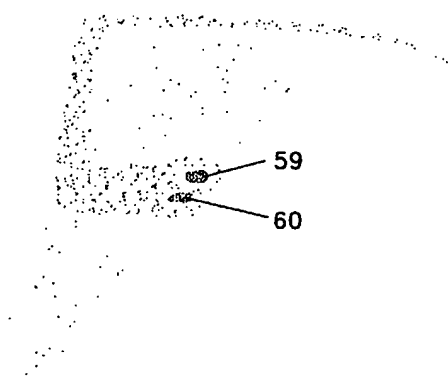
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(54) Title: HIGH PURITY X-CHROMOSOME BEARING AND Y-CHROMOSOME BEARING POPULATIONS OF SPERMATOOA



(57) Abstract: Isolated non-naturally occurring populations of spermatozoa (15) having high purity and technologies to differentiate spermatozoa (28) based on characteristics such as mass, volume, orientation, or emitted light including methods of analysis and apparatus such as beam shaping optics (30) and detectors (32).

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## HIGH PURITY X-CHROMOSOME BEARING AND Y-CHROMOSOME BEARING POPULATIONS OF SPERMATOZOA

### I. TECHNICAL FIELD

Isolated high purity X-chromosome bearing or Y-chromosome bearing  
5 populations of spermatozoa and technologies to isolate spermatozoa, particles, or events  
based upon differentiation characteristics such as mass, volume, DNA content, or the like.

### II. BACKGROUND

Isolated high purity X-chromosome bearing or Y-chromosome bearing  
populations of spermatozoa can be utilized to accomplish in vitro or in vivo artificial  
10 insemination of or fertilization of ova or oocytes of numerous mammals such as bovids,  
equids, ovids, goats, swine, dogs, cats, camels, elephants, oxen, buffalo, or the like. See  
also, United States Patent 5,135,759, hereby incorporated by reference.

However, conventional technologies for separating spermatozoa into X-  
chromosome bearing and Y-chromosome bearing populations can result in spermatozoa  
15 populations having low purity. Regardless of the separation method spermatozoa have  
not been routinely separated into X-chromosome bearing and to Y-chromosome bearing  
sperm samples having high purity, such as 90%, 95%, or greater than 95%.

A number of techniques, directly or indirectly based on differences in size, mass,  
or density have been disclosed with respect to separating X-chromosome bearing from Y-  
20 chromosome bearing spermatozoa. As disclosed by United States Patent No. 4,474,875, a  
buoyant force is applied to all sperm cells simultaneously and X-chromosome bearing and  
Y-chromosome bearing spermatozoa may then be isolated at different locations in the  
separation medium. United States Patent No. 5,514,537 discloses a technique whereby  
spermatozoa traverse a column packed with two sizes of beads. The larger X-chromosome  
25 bearing spermatozoa become isolated in the layer containing the larger beads, while the  
smaller Y-chromosome bearing spermatozoa become isolated in the layer containing the  
smaller beads. United States Patent No. 4,605,558 discloses that spermatozoa may be

made differentially responsive to a density gradient and United States Patent No.4,009,260 exploits the differences in migration-rate, or swimming-speed, between the Y-bearing spermatozoa, and the X-chromosome bearing spermatozoa, through a column of retarding medium.

- 5           A problem common to each of the above-mentioned technologies may be that they each act on all the spermatozoa in a 'bulk-manner', meaning that all the spermatozoa undergo the same treatment at the same time, and the Y-chromosome bearing sperm cells come out faster, earlier, or at a different position than X-chromosome bearing sperm cells. As such, individual sperm cells may not be assessed and there may be no actual
- 10 'measurement' of volume, mass, density, or other sperm cell characteristics. One-by-one assessment of sperm cells can provide advantages in that the actual separation process can be monitored, and objective quantitative data can be generated even during the separation process, and separation parameters altered as desired. Furthermore, these technologies may not be coupled with flow cell sorting devices.
- 15           Flow cytometer techniques for the separation of spermatozoa have also been disclosed. Using these techniques spermatozoa may be stained with a fluorochrome and made to flow in a narrow stream or band passing by an excitation or irradiation source such as a laser beam. As stained particles or cells pass through the excitation or irradiation source, the fluorochrome emits fluorescent light. The fluorescent light may be
- 20 collected by an optical lens assembly, focused on a detector, such as a photomultiplier tube which generates and multiplies an electronic signal, which may then be analyzed by an analyzer. The data can then be displayed as multiple or single parameter chromatograms or histograms. The number of cells and fluorescence per cell may be used as coordinates. See United States Patent No. 5,135,759, hereby incorporated by
- 25 reference. However, with respect to this type of technology a variety of problems remain unresolved and isolating highly purified populations of X-chromosome bearing or Y-chromosome bearing sperm cells be difficult.

A significant problem with conventional flow cytometer technologies can be the

orientation of objects, particles, or cells in the sheath fluid stream. This can be particularly problematic when the object or cell is irregular in shape with respect to more than one axis, such as spermatozoa for example. One aspect of this problem may be establishing the initial orientation of the object within the sheath fluid stream. A second  
5 aspect of this problem may be maintaining the orientation of the object with respect to the detector (photomultiplier tube or otherwise) during the period that emitted light from the object is measured.

Another significant problem with conventional flow cytometer technologies can be the failure to encapsulate the objects or cells in a droplet of liquid. Especially, when  
10 droplets are formed around irregularly shaped objects the droplet may not be of sufficient size to completely surround all the features of the objects or cells. For example, during flow cytometry operation as above-described droplets can be formed at very high speed, even as many as 10,000 to 90,000 droplets per second and in some applications as many as 80,000 droplets per second. When spermatozoa are encapsulated into droplets,  
15 especially at these high rates of speed, a portion of the tail or neck may not be encapsulated in the droplet. That portion of the tail or neck not encapsulated in the droplet may then be responsive with the nozzle or may be responsive to the environment surrounding the droplet in a manner that interferes with subsequent droplet formation or with proper deflection of the droplet. As a result some of the spermatozoa may not be  
20 analyzed at all reducing the efficiency of the procedure, or may not be resolved sufficiently to be assigned to a population, or may be deflected in errant trajectories, or a combination of all may occur.

Another significant problem with conventional flow cytometer technologies, as  
25 well as other technologies, can be a coincidence of measurable events. One aspect of this problem can be that the incident light flux from a first event continues to produce signals after the incident light flux from a second event starts to generate a signal. As such, the two events remain at least partially unresolved from one another. Another aspect of this problem can be that two or more events are simultaneously initiated and the incident light  
30 flux comprises the contribution of all the events. As such, the multiplicity of events may

not be resolved at all and the objects corresponding to the multiplicity of events can be incorrectly assigned to a population or not assigned to a population at all, or both. Specifically, with respect to flow cytometry, individual particles, objects, cells, or spermatozoa in suspension flow through a beam of light with which they interact  
5 providing a measurable response, such as fluorescent emission. In conventional flow cytometry, Hoechst stained spermatozoa traverse a laser beam resulting in a fluorescent light emission. The fluorescent light emission from the excited fluorochrome bound to the DNA can be bright enough to produce an electron flow in conventional photomultiplier tubes for a period of time after the actual emission event has ended.  
10 Moreover, in a conventional flow cytometer, the laser beam can produce a pattern having a height of 30 $\mu$ m while the width can be approximately 80  $\mu$ m. The nucleus of a bovine spermatozoa which contains fluorochrome bound DNA can be about 9  $\mu$ m in length making the height of the laser beam some three (3) times greater than the nucleus. This difference can allow for the laser excitation of the bound fluorochrome in more than one  
15 spermatozoa within the laser beam pattern at one time. Each of these conventional flow cytometry problems decreases the ability to resolve individual events from one another.

Another significant problem with conventional flow cytometer technologies, and other technologies, can be that irregularly shaped objects, such as spermatozoa, generate differing signals (shape, duration, or amount) depending on their orientation within the  
20 excitation/detection path. As such, individuals within a homogenous population can generate a broad spectrum of emission characteristics that may overlap with the emission characteristics of individuals from another homogenous population obviating or reducing the ability to resolve the individuals of the two populations.

Another significant problem with conventional flow cytometer technologies, and  
25 other technologies, can be that objects are not uniformly exposed to the excitation source. Conventional beam shaping optics may not provide uniform exposure to laser light when the objects are close to the periphery of the beam.

Another significant problem with conventional flow cytometer technologies can

be that objects, such as spermatozoa, can be exposed to the excitation source for unnecessarily long periods of time. Irradiation of cells, such as spermatozoa, with laser light may result in damage to the cells or to the DNA contained within them.

Another significant problem with conventional flow cytometer technologies can be that there may be a disruption of the laminar flow within the nozzle by the injection tube. Disruption of the laminar flow can change the orientation of irregularly shaped objects within the flow and lower the speed of sorting and the purity of the sorted populations of X-chromosome bearing sperm or Y-chromosome bearing spermatozoa.

10        There may be additional problems with technologies that utilize stain bound to the nuclear DNA of sperm cells. First, because the DNA in the nucleus is highly condensed and flat in shape, stoichiometric staining of the DNA may be difficult or impossible. Second, stained nuclei may have a high index of refraction. Third, stain bound to the DNA to form a DNA-stain complex may reduce fertilization rates or the viability of the  
15 subsequent embryos. Fourth, the DNA-stain complex is typically irradiated with ultra-violet light to cause the stain to fluoresce. This irradiation may affect the viability of the spermatozoa. Due to these various problems, it may be preferable to use a method that requires less or no stain, or less or no ultra-violet radiation, or less or none of both.

With respect to generating high purity samples of X-chromosome bearing sperm  
20 cell or Y-chromosome bearing sperm cell populations (whether live, fixed, viable, non-viable, intact, tailless, or as nuclei), or generally, with respect to detecting small differences in photo-generated signal between serial events having relatively high incident light flux, or with respect to orienting irregularly shaped objects in a fluid stream, or eliminating coincident events within an optical path, or removing undesirably oriented  
25 objects from analysis, the instant invention addresses every one of the above-mentioned problems in a practical fashion.

### III. DISCLOSURE OF THE INVENTION

A broad object of the invention can be to provide isolated high purity X-chromosome

bearing and Y-chromosome bearing populations of spermatozoa. Isolated non-naturally occurring populations of spermatozoa that have high purity have numerous applications including sex selection of offspring from mammals, various in vitro protocols for the fertilization of ova, various in vivo protocols such as artificial insemination, business  
5 methods involving the production of prize animals or meat animals, or preservation of rare or endangered animals, to recite but a few of the applications for high purity populations of spermatozoa.

Another broad object of the invention involves both devices and methods for the production of high purity X-chromosome bearing and Y-chromosome bearing sperm samples.

10 Particular embodiments of the invention are described, which may be used in numerous applications as above-mentioned, that can be used to achieve the specific objects of differentiating between bright photoemissive events having small measurable differences in total light flux, orienting irregularly shaped objects in a fluid stream, the minimization of coincident events within an optical path, the removal of signal contributed by undesired  
15 unoriented objects within an optical path (including the removal of the object itself), and the encapsulation of irregularly shaped objects within a droplet. As such, the specific objects of the invention can be quite varied.

Another broad object of the invention can be to provide X-chromosome bearing or Y-chromosome bearing spermatozoa samples (live, fixed, viable, non-viable, intact, tailless,  
20 or sperm nuclei) having a graded level of high purity in the range of 80%, 85%, 90%, 95%, or even greater than 95%.

Another significant object of particular embodiments of the invention can be to sort spermatozoa into X-chromosome bearing and Y-chromosome bearing populations having high purity even at high separation rates. The high speed separation can produce live sperm  
25 of each sex at rates of about 500, 1000, 2000, 3000, 4000, 5000, 6000, 7000, 8,000, 9,000 or even 10,000 per second, or higher.



Another significant object of particular embodiments of the invention can be to substantially eliminate or remove spermatozoa (live, fixed, viable, non-viable, intact, tailless, or sperm nuclei) having undesired orientation in the excitation/detection portion of the flow path of a flow cytometer.

- 5        Another significant object of particular embodiments of the invention can be to provide artificial insemination samples of X-chromosome bearing or Y-chromosome bearing spermatozoa having a high level of purity.

Another significant object of particular embodiments of the invention can be to provide in vitro insemination samples of X-chromosome bearing or Y-chromosome bearing  
10 spermatozoa having a high level of purity.

Another significant object of a particular embodiment of the invention can be to preselect the sex of offspring of females inseminated with high purity artificial insemination samples, the sex of offspring of ova fertilized with high purity artificial insemination samples, with selection success rates of 80%, 85%, 90%, 95%, or greater than 95%.

- 15        Another significant object of particular embodiments of the invention can be to differentiate between photoemissive events having small differences in total emitted light flux.

Another significant object of particular embodiments of the invention can be to substantially eliminate or reduce the amount of background noise generated by a  
20 photomultiplier tube, even in the absence of light, during the period after exposure to high incident light flux.

Another significant object of particular embodiments of the invention can be to substantially eliminate saturation of the photocathode of photomultiplier tube(s) used in conjunction with flow cytometry, or otherwise.

Another significant object of particular embodiments of the invention can be to reduce the number electrons migrating from the photocathode of a photomultiplier tube to the first dynode.

Another significant object of particular embodiments of the invention can be to  
5 reduce the total flow of electrons to the N electrode of a photomultiplier tube.

Another significant object of particular embodiments of the invention can be to allow increased light flux to the photocathode of the photomultiplier tube without proportionately increasing the amount of background signal generated by the photomultiplier tube.

Another significant object of particular embodiments of the invention can be to  
10 increase the signal to background signal ratio from measured photoemissive events.

Another significant object of particular embodiments of the invention can be to allow increased amplification of the signal generated from the photomultiplier tube during high incident light flux events or serial high incident light flux events without saturating the photocathode of the photomultiplier tube.

15 Another significant object of particular embodiments of the invention can be to increase the apparent resolution of chromatograms or histograms resulting from sorting fluorochrome stained sperm, or other cells, or other objects, having small differences in emitted light flux upon excitation of the bound fluorochrome(s).

Another significant object of particular embodiments of the invention can be to  
20 improve the calibration of sorting flow cytometer instruments when used for sorting spermatozoa.

Another significant object of particular embodiments of the invention can be to increase the sperm sorting rate of flow cytometer systems.

Another significant object of particular embodiments of the invention can be to increase the purity of the sperm samples sorted by flow cytometry.

Another significant object of particular embodiments of the invention can be to provide techniques for the sorting of X-chromosome bearing sperm from Y-chromosome  
5 bearing sperm where there is a small difference in the amount of Y chromosome DNA to the amount of X chromosome DNA relative to the total amount of nuclear DNA.

Another significant object of particular embodiments of the invention can be to provide techniques which improve the apparent resolution of histograms generated during the process of sorting X-chromosome bearing sperm from Y-chromosome bearing sperm  
10 with a flow cytometer.

Another significant object of particular embodiments of the invention can be to provide beam shaping optics which minimizes coincidence of objects within the excitation/detection path.

Another significant object of particular embodiments of the invention can be to  
15 provide beam shaping optics that minimizes the total lumens an object is exposed to traversing the excitation beam. One aspect of this object can be to decrease the total lumens an object is exposed to. A second aspect of this object can be to increase the power of the light source without increasing the total lumens the object is exposed to.

Another significant object of particular embodiments of the invention can be to  
20 provide beam shaping optics that allow for uniform exposure of objects that pass through the optical path.

Another significant object of particular embodiments of the invention can be to provide a nozzle that orients irregularly shaped objects in a fluid stream. One aspect this  
25 object can be to orient elongated objects in the same direction. A second aspect of this object can be to orient dorso-laterally flattened objects in the same direction.

Another significant object of particular embodiments of the invention can be to fully encapsulate irregularly shaped objects within a drop of fluid.

Another significant object of particular embodiments of the invention can be to  
5 differentiate undesirably oriented objects from desirably oriented objects in a fluid stream.

Another object of an embodiment of the invention can be to provide differential interference contrast technology, whereby the object-plane consists of a fluid stream carrying the objects of interest, and whereby the image-plane can be used to measure the signal from the passing objects.

10 Another object of an embodiment of the invention can be to provide optics that form two laterally separated images from each object in such a way that one can be used to measure the actual volume, and one to determine the orientation. This way, objects that were not orientated properly to allow a accurate measurement of its volume can be discarded. This can be accomplished by modifications so that the light pulses, resulting from these two  
15 images can be detected independently using two pinholes in the image plane. Optics are tuned in such a way that a first image can give rise to a light pulse proportional to the volume of the object, and that a second image can give rise to a light pulse dependent on the orientation the object had when it was measured.

Another object of an embodiment of the invention can provide a manner of  
20 compensating for the fact that the objects are contained inside a fluid stream. The fluid stream can be a cylinder of water, for example, which acts as a cylindrical lens, thus distorting the image of the object. Optically, this corresponds to cylinder of higher refractive index (water) than its surroundings (air). The compensation disclosed in this invention can consist of, for example, a cylinder having a refractive index lower than its surroundings,  
25 although other compensating elements of various shapes and refractive index may also be designed as the need requires. By making sure the light passes through this compensation element, the optical effect of the fluid stream can be compensated by the exactly opposite behavior of the compensation element.

Naturally further objects of the invention are disclosed throughout other areas of the specification and claims.

#### IV. BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows a generalized flow cytometer.

5        Figure 2 shows a second view of a generalized flow cytometer.

Figure 3 shows a comparison of univariate histograms from flow cytometers (#1, #2, and #3) without the amplifier invention (Figure 3A) with univariate histograms for the same flow cytometers using a particular embodiment of the amplification invention (Figure 3B) illustrating the improved resolution between X-chromosome bearing and Y-chromosome  
10        bearing populations of bovine spermatozoa.

Figure 4 shows univariate and bivariate histograms illustrating the conventional resolution between X-chromosome bearing and Y-chromosome bearing populations of bovine spermatozoa.

Figure 5 shows univariate and bivariate histograms illustrating improved resolution  
15        between X-chromosome bearing and Y-chromosome bearing populations of bovine spermatozoa using a particular embodiment of the amplification invention.

Figure 6 shows a second example of univariate and bivariate histograms illustrating the conventional resolution between X-chromosome bearing and Y-chromosome bearing populations of bovine spermatozoa.

20        Figure 7 shows a second example of univariate and bivariate histograms illustrating the improved resolution between X-chromosome bearing and Y-chromosome bearing populations of bovine spermatozoa using a particular embodiment of the amplification invention.

Figure 8 shows univariate and bivariate histograms illustrating the conventional resolution between X-chromosome bearing and Y-chromosome bearing populations of equine spermatozoa.

Figure 9 shows univariate and bivariate histograms illustrating the improved  
5 resolution between X-chromosome bearing and Y-chromosome bearing populations of equine spermatozoa using a particular embodiment of the amplification invention.

Figure 10 shows univariate and bivariate histograms illustrating the improved resolution between X-chromosome bearing and Y-chromosome bearing populations of equine spermatozoa nuclei using a particular embodiment of the amplification invention.

10 Figure 11 shows a particular embodiment of the circuit board modification to make the amplification invention with respect to a MoFlo® flow cytometer.

Figure 12 shows an electrical schematic diagram of a particular embodiment of the amplification invention with respect to a MoFlo® flow cytometer.

Figure 13 shows the laser beam pattern using conventional beam shape optics (Figure  
15 13A) and the laser beam pattern using the reduced height beam shape optics (Figure 13B).

Figure 14 shows a bar graph that compares the purity of separated X-chromosome bearing spermatozoa (Figure 14A) and Y-chromosome bearing spermatozoa (Figure 14B) using conventional technology or using the amplification invention independently or in conjunction with reduced height beam shaping optics.

20 Figure 15 shows a front view of the reduced height beam shaping optics.

Figure 16 shows a top view of the reduced height beam shaping optics.

Figure 17 shows a perspective and two cross sections of the object orienting nozzle

invention.

Figure 18 shows a graded series of cross sections of the object orienting nozzle invention.

Figure 19 shows a front view and an end view of an embodiment of the beveled  
5 injection tube invention.

Figure 20 illustrates the removal of undesired unoriented spermatozoa (RUUS) invention by comparison of signal(s) from the oriented spermatozoa (Figures 20A and 20B) and the signal(s) from the unoriented spermatozoa (Figures 20C and 20D).

Figure 21 shows a perspective of another embodiment of the beveled injection tube  
10 invention having a paddle shaped beveled blade.

Figure 22 shows a conventional optics technology coupled to a flow cytometer.

Figure 23A shows the shape and size of a typical spermatozoon and Figure 23B shows the difference between correctly and non-correctly orientated spermatozoa.

Figure 24 shows an embodiment of the invention having construction allowing the  
15 measurement of two signals, for example volume and orientation.

Figure 25A and B shows an embodiment of the invention having two halves with a pinhole corresponding to each half, Figure 25C shows an image plane of an embodiment of the invention, Figure 25D shows an embodiment of the invention having two independently rotatable polarizers.

20 Figures 26A and 26B illustrates the compensation method for the fluid stream for an embodiment of the invention, Figure 26C shows an embodiment of a compensation element, 26 D shows another embodiment of a compensation element where images of a fluid stream

and from the compensation element fall on top of each other in the image plane.

Figure 27 shows an embodiment of the interference optics invention.

Figure 28 shows an a second view of the interference optics invention.

#### V. MODE(S) FOR CARRYING OUT THE INVENTION

5 The invention involves isolated high purity X-chromosome bearing and Y-chromosome bearing populations of spermatozoa or sperm cells. High purity X-chromosome bearing and Y-chromosome bearing populations of spermatozoa can comprise populations of intact live spermatozoa, and may also comprise populations of  
10 tailless spermatozoa (sperm nuclei), or populations of other viable or non-viable forms of spermatozoa, as may be desired. While particular examples are provided that describe the invention in the context of separating intact live sperm cells each having a sperm cell head, necks, and tail, it should be understood that the technologies described can have various applications with respect to sperm nuclei as well. X-chromosome bearing and Y-chromosome bearing populations of spermatozoa should further be understood to  
15 encompass spermatozoa from any male of a species of mammal including, but not limited to, spermatozoa from humans and spermatozoa from commonly known animals such as bovids, equids, ovids, canids, felids, goats, or swine, as well as less commonly known animals such as elephants, zebra, camels, or kudu. This list of animals is intended to be exemplary of the great variety of animals from which spermatozoa can be routinely sorted  
20 at 90% or greater purity, and is not intended to limit the description of the invention to the spermatozoa from any particular species of mammals.

High purity separated spermatozoa from the various species of mammals can be incorporated into products that can be used with artificial insemination protocols or as part of commercial business methods such as those as described in United States Patent  
25 Application Nos. 60/211,093, 60/224,050, or Patent Cooperation Treaty Application No. US99/17165; or be used with low dose insemination protocols as described in Patent Cooperation Treaty Application No. US98/27909, or used in vitro fertilization of oocytes



from animals, including humans, as described in United States Patent Application No. 60/253,785, each of the above-mentioned references are hereby incorporated by reference.

The use of the term purity or high purity should be understood to be the percent of the isolated spermatozoa population bearing a particular differentiating characteristic or  
5 desired combination of characteristics. For example, where a population of spermatozoa are separated based upon bearing an X-chromosome as opposed to a Y-chromosome, a X-chromosome bearing population having 90% purity comprises a population of spermatozoa of which 90% of the individual spermatozoa bear an X-chromosome while 10% of such population of spermatozoa may bear a Y-chromosome. As such, high purity  
10 with respect to X-chromosome bearing populations or Y-chromosome bearing populations can comprise a purity selected from the group consisting of between 90% to about 100%, between about 91% to about 100%, between about 92% to about 100%, between about 93% to about 100%, between about 94% to about 100%, between about 95% to about 100%, between about 96% to about 100%, between about 97% to about  
15 100%, between about 98% to about 100%, between about 99% to about 100%.

Importantly, while numerous embodiments of the invention describe isolated high purity X-chromosome and Y-chromosome bearing populations of spermatozoa, and while the description further discloses high purity spermatozoa separation devices and methods of how to isolate and how to use isolated high purity populations of spermatozoa, the  
20 basic concepts of the invention should be understood to be applicable to other types of particles or events having particle differentiation characteristics or event differentiation characteristics. It should be understood that the invention can be applicable to a variety of circumstances in which resolving small differences in photogenerated signal may be necessary, such as product defect detection, field flow fractionation, liquid  
25 chromatography, electrophoresis, computer tomography, gamma cameras, time of flight instruments, or the like as would be readily understood by those skilled in those arts.

Moreover, while this disclosure provides descriptions of embodiments of apparatus and methods for flow separation of X-chromosome bearing spermatozoa from

Y-chromosome bearing spermatozoa, the description of these embodiments of the invention is not meant to reduce the scope of the invention to only flow separation of spermatozoa or only to high purity flow cytometer spermatozoa separation systems but rather these examples are intended to exemplify the basic concepts of the invention in a  
5 practical manner so that they may be applied to the wide variety of applications.

Now referring to Figures 1 and 2, a flow cytometer embodiment of the invention is shown which includes a particle or cell source (1) which acts to establish or supply particles or cells stained with at least one fluorochrome for analysis. The particles or cells are deposited within a nozzle (2) in a manner such that the particles or cells are introduced  
10 into a fluid stream or sheath fluid (3). The sheath fluid (3) is usually supplied by some sheath fluid source (4) so that as the particle or cell source (1) supplies the particles or cells into the sheath fluid (4) they are concurrently fed through the nozzle (2).

In this manner it can be easily understood how the sheath fluid (3) forms a sheath fluid environment for the particles or cells. Since the various fluids are provided to the  
15 flow cytometer at some pressure, they flow out of nozzle (2) and exit at the nozzle orifice (5). By providing some type of oscillator (6) which may be very precisely controlled through an oscillator control (7), pressure waves may be established within the nozzle (2) and transmitted to the fluids exiting the nozzle (2) at nozzle orifice (5). Since the oscillator (6) acts upon the sheath fluid (3), the stream (8) exiting the nozzle orifice (5)  
20 eventually and regularly forms drops (9). Because the particles or cells are surrounded by the fluid stream or sheath fluid environment, the drops (9) may entrain within them individually isolated particles or cells, and can be sperm cells with respect to some embodiments of the invention.

Since the drops (9) can entrain particles or cells, the flow cytometer can be used to  
25 separate particles, cells, sperm cells or the like based upon particle or cell characteristics. This is accomplished through a particle or cell sensing system (10). The particle or cell sensing system involves at least some type of detector or sensor (11) which responds to the particles or cells contained within fluid stream (8). The particle or cell sensing system

(10) may cause an action depending upon the relative presence or relative absence of a characteristic, such as fluorochrome bound to the particle or cell or the DNA within the cell that may be excited by an irradiation source such as a laser exciter (12) generating an irradiation beam to which the particle can be responsive. While each type of particle, cell,  
5 or the nuclear DNA of sperm cells may be stained with at least one type of fluorochrome different amounts of fluorochrome bind to each individual particle or cell based on the number of binding sites available to the particular type of fluorochrome used. With respect to spermatozoa, the availability of binding sites for Hoechst 33342 stain is dependant upon the amount of DNA contained within each spermatozoa. Because X-  
10 chromosome bearing spermatozoa contain more DNA than Y-chromosome bearing spermatozoa, the X-chromosome bearing spermatozoa can bind a greater amount of fluorochrome than Y-chromosome bearing spermatozoa. Thus, by measuring the fluorescence emitted by the bound fluorochrome upon excitation, it is possible to differentiate between X-bearing spermatozoa and Y-bearing spermatozoa.

15 In order to achieve separation and isolation based upon particle or cell characteristics, emitted light can be received by sensor (11) and fed to some type of separation discrimination system or analyzer (13) coupled to a droplet charger which differentially charges each droplet (9) based upon the characteristics of the particle or cell contained within that droplet (9). In this manner the separation discrimination system or  
20 analyzer (13) acts to permit the electrostatic deflection plates (14) to deflect drops (9) based on whether or not they contain the appropriate particle or cell.

As a result, the flow cytometer acts to separate the particle or cells (16) by causing them to be directed to one or more collection containers (15). For example, when the analyzer differentiates sperm cells based upon a sperm cell characteristic, the droplets  
25 entraining X-chromosome bearing spermatozoa can be charged positively and thus deflect in one direction, while the droplets entraining Y-chromosome bearing spermatozoa can be charged negatively and thus deflect the other way, and the wasted stream (that is droplets that do not entrain a particle or cell or entrain undesired or unsortable cells) can be left uncharged and thus is collected in an undeflected stream into a suction tube or the like as

discussed in United States Patent Application 09/001,394, hereby incorporated by reference herein. Naturally, numerous deflection trajectories can be established and collected simultaneously.

To routinely separate particles, cells, sperm cells, or spermatozoa (intact, live,  
5 fixed, viable, non-viable, or nuclei) into high purity X-chromosome bearing and Y-chromosome bearing populations, the particle differentiation apparatus or methods used must provide high resolution of the differentiation characteristics that are used as the basis of analysis and separation.

With respect to spermatozoa, differentiating between the light emitted by the  
10 fluorochrome bound to the nuclear DNA of X-chromosome bearing sperm cells and the light emitted by the fluorochrome bound to the nuclear DNA of Y-chromosome bearing sperm cells may be difficult as discussed above.

In many applications, the total emitted light from photoemissive events incident to the detector, which can be a photomultiplier tube, can be high while the difference  
15 between the emitted light of each photoemissive events to be differentiated can be small. The problem can be exacerbated when the photoemissive events happen serially at high rate of speed and the time period between photoemissive events is short, such as with high speed cell sorting using flow cytometers. When separating particles, cells, or sperm cells based upon the difference in bound fluorochrome the cells flow past an excitation  
20 source and a high number of emissive events per second can be established. As a result, the amount of emitted light generated in the stream of particles, cells, or sperm cells, can be enormous. As the speed of the stream is increased, the intercept point with the excitation source becomes very bright. This high level of incident light upon the photocathode of the photomultiplier tube can cause a very low signal to background  
25 signal ratio. The amount of background signal can be further exacerbated when fluorochrome such as Hoechst 33342 can be used to label the nuclear DNA of sperm cells.

Most solutions to the problem have focused on decreasing the total amount of light flux upon the photocathode tube by placing optical filters in front of the photomultiplier tube. This approach does not change the proportion of signal to background signal and subsequent attempts to increase the sensitivity of the  
5 photomultiplier tube generates additional background signal as the photomultiplier tube saturates from the amount of background signal.

Typically, photomultiplier tubes have an operation voltage range of about 400 volts to about 900 volts. The lower limit of linear operation of standard photomultiplier tubes, such as the R928 and R1477 photomultiplier tubes available from Hamamatsu  
10 Corporation, may be about 300 volts. As such, equipment or instruments which employ photomultiplier tubes are configured to operate such photomultiplier tubes at or above 400 volts. Even where reduction of the number of electrons at the anode is desired, as disclosed in United States Patent Nos. 4,501,366 and 5,880,457 the voltage between the photocathode and the first dynode is maintained at a high voltage and reduction of the  
15 electrons at the anode is accomplished by either decreasing the voltage to the remaining dynodes, or the inherent dark noise or shot noise is filtered out electronically.

Unexpectedly, reducing the amount of voltage to the photomultiplier tube below 400 volts to about 280 volts, or about 250 volts, or even to 0 volts can allow small differences in photoemissive light to be differentiated even when the total light emitted  
20 from each photoemissive event is high, or even when there are a high number of bright serial events per second. With respect to the rate of photoemissive events generated from the irradiation of fluorochromes bound to the nuclear DNA of spermatozoa, the invention allows the rate of photoemissive events that can be achieved during separation of spermatozoa into X-chromosome bearing and Y-chromosome bearing populations to  
25 be increased to a separable event rate of at least 5000 separable events per second, at least 6000 separable events per second, at least 7000 separable events per second, at least 8000 separable events per second, at least 9000 separable events per second, at least 10,000 separable events per second, at least 11,000 separable events per second, at least 12,000 separable events per second, at least 13,000 separable events per second, at least 14,000

separable events per second, at least 15,000 separable events per second, at least 16, 000  
separable events per second, at least 17,000 separable events per second, at least 18,000  
separable events per second, at least 19,000 separable events per second, at least 20,000  
separable events per second, at least 25,000 separable events per second, at least 30,000  
5 separable events per second, and at least 35,000 separable events per second, or greater.

As a specific example, existing Cytomation SX MoFlo® sorting flow cytometers  
are configured to operate the photomultiplier tube at 400 volts minimum. The gain can be  
adjusted to operate the photomultiplier tube at higher voltages but not lower voltages. SX  
MoFlo® flow cytometers can be converted by reconfiguring the photomultiplier  
10 controllers. The R16C resistor (2.49 kilohms) on channel three can be replaced by a 2.0K  
resistor to alter the gain of the amplifier that controls the photomultiplier tube. This  
conversion allowed the photomultiplier tube to be operated at about 280 volts. Similar  
conversion of SX MoFlo® flow cytometers with two 3.75 kilohm resistors in parallel, or  
a 1.3 kilohm resistors can allow the photomultiplier tube to be operated at voltages of  
15 about 200 volts, or just above zero volts, respectively. Also with respect to this  
conversion, the neutral density filter in front of the photocathode can also be removed as a  
result of operating the photomultiplier tube outside of the typical operation voltage range.

This conversion unexpectedly increases the signal to noise ratio of the  
photoemissive event as it is translated to an electronic signal by the photomultiplier tube.  
20 The cleaner signal may then be amplified by increasing the gain amplifier to the analog to  
digital converter of the analyzer (13) to the appropriate level and output may be generated  
as univariate or bivariate histograms.

Now referring to Figure 3, a comparison of univariate histograms generated on  
three different SX MoFlo® flow cytometers (#1, #2, #3) prior to the use of the invention  
25 (Figure 1A), and using the invention (Figure 1B) with respect to the separation of intact  
live ejaculated bovine sperm are shown. As can be understood from the univariate  
histograms, the resolution (the apparent differentiation of the X-chromosome bearing  
population from the Y-chromosome bearing population represented by the valley between

peaks) of intact live X-chromosome bearing spermatozoa (17) from live Y-chromosome bearing spermatozoa (18) can be substantially improved by use of the invention.

The mean separation rate or sort rates of intact live spermatozoa prior to use of this embodiment of the invention with the SX MoFlo® flow cytometers was about  $17.9 \times 10^6 / 4.5$  hours of both X-chromosome bearing spermatozoa and Y-chromosome bearing spermatozoa (i.e. about 1,100 separations or sorts per second in each of two streams -- the first stream X-chromosome bearing spermatozoa and the second stream Y-chromosome bearing spermatozoa) at about 87% purity with a range of 84% to 93% purity. The separable event rate was 22,000, 23,000, and 20,000 respectively for the three sorts.

10 The mean sort rates of live spermatozoa after the above-mentioned conversion was about  $40.3 \times 10^6 / 4.5$  hour sort (i.e. about 2,500 sorts per second per stream) at about 90.8% purity with a range of 89% to about 92%. The events per second were 13,000, 15,000, and 19,500 respectively for the three sorts.

As can be understood from the data not only did this embodiment of the invention result in increased purity of the separated spermatozoa populations but also allowed the separation rate or sort rate to be more than doubled while the separable events rate was actually decreased.

Similarly, referring now to Figures 4 and 5, which show bivariate histograms from sorting of intact live bull spermatozoa with the SX MoFlo® flow cytometer #1 prior to using the invention (Figure 4) and after the above-mentioned conversion (Figure 5). Prior to using the invention, the SX MoFlo® flow cytometer was initially operated at 440 volts at the photocathode with the laser adjusted to 135 MW, a gain of 1X and with a neutral density filter of 1.0 (1/10th amplitude) at about 10,000 events per second. Upon using the invention, the SX MoFlo® flow cytometer was operated at about 262 volts at the photocathode, with the laser adjusted to about 100mW, a gain of 4X, without the neutral density filter, at about 10,000 separable events per second. As can be understood from this data there is a large increase in resolution as evidenced by the increased depth of the

valley between the X-chromosome bearing population (19) and the Y-chromosome bearing population (20).

Similarly, referring now to Figures 6 and 7, which show bivariate histograms from sorting of intact live bull spermatozoa with the SX MoFlo® flow cytometer #2 before  
5 using this embodiment of the invention (Figure 6) and upon using this embodiment of the invention (Figure 7) operated at the same parameters as shown in Figures 3 and 4 respectively. Again, there can be a large increase in resolution as evidenced by the depth of the valley between the X-chromosome bearing population (21) and the Y-chromosome bearing population (22).

10 Now referring to Figures 8 and 9, which show bivariate histograms from separation or sorting of intact live equine spermatozoa with the SX MoFlo® flow cytometer before using this embodiment of the invention (Figure 8) and upon using this embodiment of the invention (Figure 9). When using this embodiment of the invention, live equine spermatozoa were separated or sorted with the laser power at 100mW with the  
15 photomultiplier tube voltage below 300 volts. The separation rates or sort rates exceeded 4,800 sorts per second average at 12,000 events per second. The increased resolution of the X-chromosome bearing population (23) and the Y-chromosome bearing population (24) is dramatic. The data shows that about 8 to about 9 channels separation can be achieved with this embodiment of the invention as compared to 5 channels of separation  
20 between the peaks without the use of this embodiment of the invention. The purity of both the sorted X-chromosome bearing population and the sorted Y-chromosome bearing population was about 93%.

Now referring to Figure 10, which shows a univariate histogram and a bivariate dot plot from sorting of Hoechst 33342 stained stallion sperm nuclei (S-05400) separated  
25 using this embodiment of the invention. The nuclei were prepared from freshly ejaculated stallion sperm. The sperm were washed by centrifugation, sonicated and the resultant heads and tails separated using Percoll density gradient centrifugation. the isolated heads were washed, fixed with 2% formalin and then stained with Hoechst 33342. The stained



nuclei were stabilized using sodium azide (0.5%). The sample was run at 5000 events per second to produce the histograms. The stained nuclei were then used to calibrate an SX MoFlo® flow cytometer was converted as above-mentioned to incorporate the photomultiplier tube embodiment of the invention. Compensation was used to level the two populations (X stained nuclei and Y stained nuclei) in the bivariate plot. Note that the two populations of equine sperm nuclei are nearly fully resolved to baseline as shown by the univariate plot.

Now referring to Figure 11, a modification specifically for SX MoFlo® flow cytometer includes the use of two resistors in parallel to provide the correct value of 1.8K. Two 3.57K resistors (25) and (26) are equal to about 1.785K which can be sufficiently close to the value to be effective. With this modification the photomultiplier tube on this particular instrument can then be run at about 200 volts. Naturally, a similar modification can be made to other flow cytometer instruments or other instruments which use a photomultiplier tube to measure the amount of light emitted from particular events. Figure 12, provides a electrical schematic diagram for this particular embodiment of the invention.

Another important embodiment of the invention can be a reduced height irradiation beam pattern optics. As shown by Figure 13A, conventional irradiation beam shaping optics generate a beam pattern (27) that can have a height can be greater than much greater than the height of the sperm cell head(s) (28) passing through it. As a result, more than a single sperm cell head containing fluorochrome bound DNA can enter the irradiation beam pattern at the same time. In that case, the fluorochrome(s) bound to the DNAs contained within the multiple sperm heads can be excited simultaneously and fluoresce within a single emissive event. As such, the prior or subsequent emissive event can include coincident light flux contributed from other sperm head(s) in the beam pattern (27). This results in a reduced difference in mean light flux between light emissive events which distinguish between X-chromosome bearing spermatozoa and Y-chromosome bearing spermatozoa. It can also decrease the difference in mean light flux between events that compare light emissions of X-chromosome bearing spermatozoa or

Y-chromosome bearing spermatozoa. Importantly, coincident excitation of fluorochrome bound to multiple DNAs increases the mean brightness of the events making the measurable difference in light flux between events an even smaller percentage of the total light flux emitted. This makes quantification of the differences between events even more  
5 difficult.

By reducing the height of the beam shape as shown by Figure 13B, the coincidence of multiple sperm heads being within the reduced height beam (29) pattern during the same measured event is reduced. This results in an increased mean difference between light emissive events which distinguish between X-chromosome bearing  
10 spermatozoa and Y-chromosome bearing spermatozoa. It can also reduce the mean total light flux for each measured emissive event. For particular embodiments of the invention used for sorting bovine sperm which have a nucleus of about  $9\mu\text{m}$ , it has been found that the height of the beam can be about  $20\mu\text{m}$ . In this application, it has been found that vertical beam heights of less than  $20\mu\text{m}$  did not provide an additional gain in resolution.

15 Referring to Figure 14, it can be understood that the use of reduced height irradiation beam pattern optics can improve the purity of sorted populations of X-chromosome bearing bovine spermatozoa (Figure 14A) and sorted populations of Y-chromosome bearing bovine sperm (Figure 14B) that have been stained with Hoechst 33342 stain. This is true for both 25% and 40% sort gates of the univariate peak. As can  
20 be further understood from Figure 14, the reduced height beam pattern optics can improve purity of separated spermatozoa independent of any other aspect of the invention, such as modification of photomultiplier circuitry embodiment of the invention (new PMT) as described above, or can be used in conjunction with the modified photomultiplier embodiment of the invention to increase the purity of separated spermatozoa samples  
25 even further.

Another advantage of the reduced height beam pattern optics can be that the transit time of the spermatozoa in the excitation laser beam or irradiation beam can be reduced. A reduced amount of irradiation time within the excitation laser beam may

result in less stress or damage to the spermatozoa.

Again referring to Figure 14B, it can be understood that the reduced height beam pattern can be used in conjunction with a irradiation beam pattern having greater area than conventionally used. For example, conventional beam patterns (27), such as that shown  
5 in Figure 14A, have an elliptical pattern of about 30  $\mu\text{m}$  X 80  $\mu\text{m}$  while the invention when used for sorting bovine sperm generates optimal resolution between X-chromosome bearing and Y-chromosome bearing populations when the beam has a 20  $\mu\text{m}$  X 160  $\mu\text{m}$  beam pattern (29). The 20  $\mu\text{m}$  X 160  $\mu\text{m}$  beam pattern has approximately 1.3 times the area of the 30  $\mu\text{m}$  X 80  $\mu\text{m}$  beam pattern. As such, there can be an inverse proportion in  
10 loss of energy at the incident point. This makes it possible to increase the excitation laser power without concern for increasing the irradiation damage to the spermatozoa. For example, if an instrument has conventional beam shaping optics that produce a 30  $\mu\text{m}$  X 80 $\mu\text{m}$  irradiation beam pattern and the excitation laser is conventionally powered at 150mW, then particular embodiments of the invention with a 20  $\mu\text{m}$  X 160  $\mu\text{m}$  beam  
15 pattern can have an excitation laser powered at 300mW without increasing the total amount of power at the incident point. Alternately, the excitation laser can be run at 150mW to take advantage of the lower per unit area irradiation energy, decreased irradiation damage, longer laser life, and the like.

In comparison to conventional beam shaping optics and conventional  
20 photomultiplier tube amplification devices, the reduced height beam pattern optics invention and the photomultiplier tube amplification invention can increase the purity of X-chromosome bearing and Y-chromosome bearing populations of spermatozoa by about 4%, or more.

The beam shaping optics invention (30) can be installed to a flow cytometer as  
25 shown in Figures 15 and 16. As can be understood, the light emitted (31) by laser excitation of fluorochrome(s) bound to the DNA contained within spermatozoa can be detected by photomultiplier tubes (32) situated at 0 and 90 degrees relative to the flat surface of the sperm head (28) as it flows through the excitation laser beam pattern.

As can be understood, stained spermatozoa must be pumped through the excitation beam or irradiation beam in a precise manner so that each sperm head is oriented with the flat surface of the sperm head directed toward the photomultiplier tube that is the 0 degree detector. Accurate measurement of the DNA content of the spermatozoa can only be measured from the flat surface of the paddle-shaped sperm head(28). Thus, only that proportion of the spermatozoa that enter the excitation beam in the proper orientation can be measured accurately and sorted based upon DNA content.

Now referring to Figures 17, 18, and 19, particular embodiments of the invention can also have an particle or sperm cell orienting nozzle (33) that hydrodynamically forces the flattened sperm head into the proper orientation as they pass in front of the photomultiplier(s). As shown by Figure 17, the orienting nozzle has interior surfaces (34) that form a cone-like shape. The internal cone gradually changes from circular at the inlet end (35) into a highly elliptical shape near the orifice (36) where the stream exits the tip. The orifice (36) can be circular rather than elliptical. Thus, the internal aspect of the orienting nozzle (34) goes from a round entrance to a narrow ellipse to a round exit shortly before the orifice(36). This internal shape is further clarified by the cross sections of the orienting nozzle shown by Figure 18.

As shown by Figure 19 and 21, the injection tube (37)(which may be about 0.061 inches in diameter) can be used with the orientation nozzle (or with a conventional nozzle) (33) which can be beveled near the tip to form a blade (38). The flattened blade (38) can be oriented at an angle 90 degrees from the greatest dimension of the ellipse in the orientation nozzle (33). The internal diameter of the injection needle can be about 0.010 inch in diameter forming a rounded orifice (39) in the center of the flattened needle tube blade (38).

In particular embodiments of the beveled injection tube the beveled blade can be configured in the paddle shape illustrated by Figure 21. The paddle shaped beveled blade can assist in maintaining laminar flow of the sheath fluid within the nozzle (whether conventional nozzle or orienting nozzle). As such, the laminar flow of liquid maintained

by the paddle shaped beveled blade presents less disruption of the objects injected into it. Spermatozoa introduced into the laminar flow of sheath fluid maintained by an embodiment of the injector tube invention having the paddle shaped beveled blade allows for a 20%, 30%, 40%, 50% or even greater increase in spermatozoa sorting rates over  
5 conventional injection tube technology. High speed sorting of spermatozoa at rates of about 4,000 to about 10,000 sorts of each sex per second can be accomplished. High purity (90% or greater) of the X-chromosome bearing and Y-chromosome bearing populations can be established at even these high sort rates. The injector tube invention with the beveled paddle shaped tip can be used independently of or in combination with  
10 the other inventions described herein or other technology such as that described in United States Patent Application No. 09/454,488 or International Patent Application No. PCT/US00/42350, each hereby incorporated by reference.

As shown by Figure 21, certain embodiments of the beveled blade injector tube invention or beveled blade paddle shape invention can further include laminar flow  
15 enhancement grooves (40). The laminar flow enhancement grooves (40) assist in maintaining a laminar flow to the orifice of the injector tube. Again, the enhanced laminar flow allows for more spermatozoa to maintain the correct orientation in the laminar sheath fluid flow resulting in higher numbers of sortable event rates which in turn leads to higher sort rates for each sex or spermatozoa.

20 In another embodiment of the invention, the orienting nozzle orifice (39) or other conventional can be sized to form droplets which encapsulate intact live sperm as they exit the orifice (39). Encapsulation of the sperm cells does not occur in conventional sperm cell entrainment technology. Rather a portion of the sperm cell tail resides outside of the droplet. For example, bovine sperm cells have a length of about 13.5 microseconds  
25 when the fluid stream has a pressure of about 50 pounds per square inch (i.e. the length of time for the entire length of the sperm cell to pass through the irradiation beam at about 50 pounds per square inch fluid stream pressure). Conventional droplet formation techniques for entraining bovine sperm cells establish a 14 microsecond droplet (i.e. the time it takes to form a single droplet waveform in a fluid stream) from a nozzle having an

orifice with a diameter of about 70 micrometers which can be made responsive to an oscillator operated at about 35 kilohertz. As such, a portion of the sperm cell tail readily protrudes from the droplet. To prevent the sperm cell tail from protruding from the droplet, one embodiment of the droplet encapsulation invention provides an orifice of  
5 about 100 micrometers that can produce a droplet of about 28 microseconds at about 50 pounds per square inch at about 30 kilohertz. By entirely encapsulating the intact live sperm cell, including the tail portion, the sperm cell interacts with the nozzle less upon charging of the droplet and the deflection of the droplet can be more accurate. This leads to less cross contamination of X-chromosome bearing sperm with Y-chromosome bearing  
10 sperm and also allows deflected spermatozoa to be more uniformly collected. Sperm that are uniformly deflected can be directed to collection surfaces that are cushioned by various liquids. Cushioning the separated spermatozoa can be important in reducing stress as described in United States Patent Application No. 09/001,394, hereby incorporated by reference. With respect to spermatozoa from other species of mammals,  
15 the invention can be varied to produce droplet sizes to encapsulate the varying lengths of sperm cells. Depending on the length of the spermatozoa and the pressure of the fluid stream the droplet encapsulation invention can still achieve droplet formation rates of at least 10,000 droplets per second, at least 20,000 droplets per second, at least 30,000 droplets per second, at least 40,000 droplets per second, at least 50,000 droplets per  
20 second, at least 60,000 droplets per second, at least 70,000 droplets per second, at least 80,000 droplets per second, at least 90,000 droplets per second, at least 100,000 droplets per second and so on up to about 200,000 droplets per second in some embodiments of the droplet encapsulation invention.

Even with the orienting nozzle invention there will be a certain number of  
25 spermatozoa, or particles, which are not properly oriented in the beam pattern. As described above, if the orientation of a sperm head is not proper then the DNA content cannot be measured accurately based upon the emitted light. Particular embodiments of the present invention provide for the removal of undesired unoriented spermatozoa (RUUS) or particles within a fluid stream.

Referring now to Figures 16 and 20A, it can be understood that accurate measurement of the DNA content of a spermatozoa depends upon the flat surface of the paddle-shaped sperm head (28) being oriented properly with the detector. Thus, only that proportion of the spermatozoa that enter the excitation beam in the proper orientation as shown by Figures 16 and 20A can be measured accurately and sorted in to X-chromosome bearing and Y-chromosome bearing populations based upon DNA content. As shown by Figures 20A and 20B, spermatozoa which transit through the excitation beam in proper orientation generate an oriented emission signal plot (40) that can be shaped differently than the unoriented emission signal plot (41) that is generated by unoriented spermatozoa shown by Figure 20D. Naturally, the shape of the unoriented emission signal plot (41) generated by unoriented spermatozoa will vary depending on the degree of improper orientation in the excitation beam. These improper orientations can include the orientation shown in Figure 20C but can also include all manner of orientations that rotate the sperm head any portion of a rotation that orients the surface of the paddle-shaped head out of alignment with the detector (proper alignment shown by Figure 16), or any portion of a rotation that orients the axis of the sperm head (42) out of alignment with the direction of flow. Naturally, proper orientation may be defined differently from species to species. For some species, in which the sperm head is not paddle-shaped, the proper orientation within the excitation beam, or relative to the detectors or otherwise, may be defined by other anatomical characteristics or signal characteristics. Nonetheless, an optimized signal for the properly oriented spermatozoa of various species within the excitation window can be generated as the standard emission signal plots for subsequent comparison with serial emission events.

By comparing the shape (or the integrated area or both) of each emission signal plot with the standard emission signal plot (or standard integrated area or both) established for an oriented spermatozoa of a species of mammal, unoriented sperm can be identified, the signal subtracted from univariate or bivariate histograms, and the unoriented sperm can be affirmatively removed, if desired, so that unoriented sperm are not collected into either the X-chromosome bearing population or the Y-chromosome bearing population.

Importantly, as the invention(s) improve(s) resolution between the two spermatozoa populations being separated which increase the rate at which the populations can be separated from one another, and improves the purity of the populations that are separated. As such, it is now possible to sort spermatozoa at remarkably high speeds.

5 Sortable or separable event rates can be as high as about 35,000 per second (not including coincident events -- multiple spermatozoa within the excitation/detection window at the same time). Sortable or separable event rates correlate with high separation or sort rates which can be about 5000 to about 11,000 intact live sperm of each sex per second with a purity of 90%, 92%, 93%, 95%, or greater. The above-described

10 inventions also allow for even higher purity X-chromosome bearing and Y-chromosome bearing populations to be obtained of about 97% to about 98% or even higher by reducing the sort or separation rates to around 2000 live sperm of each sex per second.

As can be understood, the above inventions described are particularly important in achieving the highest possible sortable or separable event rates and highest possible

15 resulting separation rates which can be at least 1,000 separations per second, at least 2,000 separations per second, at least 3,000 separations per second, at least 4,000 separations per second, at least 5,000 separations per second, at least 6,000 separations per second, at least 7,000 separations per second, at least 8,000 separations per second, at least 9,000 separations per second, or at least 10,000 separations per second of each sex

20 per second, or greater.

The invention allows for high speed sorting, as set forth above, of spermatozoa even when they are difficult to stain, or have other anatomical or chemical features, that make differentiation between the X-bearing chromosome and Y-bearing chromosome populations more difficult. Even in these difficult cases, high purity X-chromosome

25 bearing and Y-chromosome bearing populations of bovine spermatozoa can be isolated at high purity of 92% to 93% by achieving sortable event rates of about 15,000 - 20,000 sortable events per second or higher as described above, and sort or separation rates of intact live spermatozoa of each sex (X-chromosome bearing and Y-chromosome bearing) of 2000 intact live sperm of each sex per second.



Now referring to Figures 23 and 24, an embodiment of the invention utilizes differential interference contrast technology to measure the volume of a particle or capsule. A basic embodiment of the invention can comprise particles that have a difference volume, such as sperm cell heads (28) that have a difference in volume  
5 between X-chromosome bearing and Y-chromosome bearing sperm cells. An electromagnetic radiation source (43) generates electromagnetic radiation or a beam of electromagnetic radiation (44) having initial waveform characteristics differentially responsive to the difference in volume between the particles or sperm cell heads (28). The electromagnetic radiation which can be laser light, but could also be numerous types  
10 of electromagnetic radiation including, but not limited to, microwave radiation, ultraviolet radiation, or the like. Upon traversing the particle or capsule or sperm head volume containing phase shifting material the electromagnetic radiation can be focused through an objective lens (45) onto a detector (46) responsive to the waveform characteristics of the electromagnetic radiation. The detector can be coupled to an analyzer (47). The  
15 analyzer can differentiate between particles based on the change in the waveform characteristics prior to traversing the volume of the particle and after traversing the volume of the particle and can analyze the signal based on integrated areas or signal shape or both. In certain embodiments the invention analyzing waveform characteristics can comprise superimposing initial waveform characteristics with altered waveform  
20 characteristics upon traversing the volume of the particle, capsule, or sperm cell head.. Superimposing the initial waveform characteristics and the phase shifted waveform characteristic can differentially modulate the intensity of the beam of electromagnetic radiation in manner that correlates to the amount of phase shift media the electromagnetic radiation traverses. The invention may also include additional filters (48), such as color  
25 filters.

Now referring to Figure 24, an embodiment of the optics invention involves using differential interference contrast optics that increase the actual distance over which the light is split up compared to conventional DIC microscopy which corresponds to the resolution limit of the microscope. In this embodiment of the invention, the induced split  
30 is larger than the size of the objects, thus giving rise to two individual images, separated

laterally, originating from one object. The second modification involves using plates of birefringent material, such as Savart plates, at a location away from the objective lens. This embodiment of the invention is easier to construct since the birefringent materials do not have to be located inside the objective housing. In conventional DIC microscopes the  
5 birefringent material is used in the form of so-called Wollaston prisms, that have to be located inside the objective housing, making it necessary to use expensive objective lenses that have been manufactured specifically for this purpose.

Components of an embodiment of the invention may be arranged in line with each other and consist of: a source of electromagnetic radiation (43), for example, a Mercury  
10 arc lamp; a spectral adjustment element, for example, a bandpass filter; a polarization adjustment element (49), for example, a sheet polarizer (53) and a waveplate (54) responsive to a rotatable mount; a light condenser (51) allowing the light to be condensed onto the particle or sperm cell, for example, a condenser lens, or set of lenses, or microscope objective; a fluid stream (8) that may contain particles or sperm cells (28),  
15 for example a fluid jet ejected under pressure; a light collector (45) to collect the light from the particle or cell, for example a 50x high working distance microscope objective and a tube lens; a beam splitter (50) to split up the beam into two, or more, components, for example, a piece of birefringent material in the form of a Savart plate, mounted in such a way that its orientation and location can be controlled accurately; image light  
20 selector (55) to select only the light corresponding to the particle or sperm cell, for instance a set of pinholes, one pinhole (53) for each of the images formed.

In one embodiment of the invention, the components may be arranged in such a way that the light source (43) or its image are located at the back focal plane of the light condenser (45) often referred to as Köhler type illumination. The image of the object  
25 plane may best coincide with the object light selector (55) or pinhole(s) (53), in order to capture the light from individual particles or sperm cells. As shown in Figures 27 and 28, components can be mounted on a sturdy optical table, or bench, using mountings, posts, and holders. Components can be mounted in such a way that focusing of the object plane can be done accurately. This can be done by equipping the fluid stream with a stream

position controller, such as micrometers, in order to turn the stream in and out of focus. In addition it may be necessary to equip the light condenser (51) with a light condenser position controller (61) allowing it to be focused onto the object plane. It may be necessary to take special care about the mounting of the birefringent elements or beam  
5 splitter (50), a three axis rotation element may be preferable.

Now referring to Figure 25, embodiments of the present invention may also include the use of both generated images, in order to determine the orientation of a asymmetrical particles the fluid stream, including, but not limited to, spermatozoa such as bull sperm cells. An orientation assessment embodiments of the invention can include an  
10 optical system that allows for control of the polarization state of the light entering the system for both generated images independently. The interference optics invention may further provide polarization adjustment element (56) that controls the polarization state of light entering the system. For the orientation detection invention the polarization adjustment element (56) may be selected in such a way that it consists of two parts, that  
15 are imaged onto image light selector (55) that in one embodiment of the invention contains the pinholes (53). This can be accomplished by locating the polarization adjustment element (56) in the conjugate plane of the image plane (55), or by using other optics, to accomplish the same thing. A simple example of this component may be a 'half-shade' piece, for instance consisting of two hemi-circular parts of polarizing material,  
20 such as sheet-polarizer, the orientation angle of which may be chosen independently. Each pinhole in the image plane can fall in one of the halves of said hemisphere. The polarization angles can be chosen in such a way that the signal of one pinhole (53) corresponds to the volume, and is relatively independent from the orientation angle of the passing object, and the other pinhole (53) has a signal that depends, to a great degree,  
25 upon this orientation angle. The two signals may be processed by analyzer (47) in a manner similar to a conventional multi-channel flow cytometry, as but one example. With respect to this example, bivariate dotplots can be made, and also allow the user to select windows on this plot.

An improvement of the 'half-shade' piece described above may be the

construction shown by Figure 25D. The same said two hemispherical parts are projected onto the image plane but the way they are generated is different. A mirror (57) breaks up the light (44) into the hemi-circular parts, and recombines them back to back. Each of the halves traverses a separate means to control its polarization state. An advantage of this  
5 embodiment is that the polarization angles can be controlled continuously and independently, thus facilitating the adjustment of the set-up. Materials used in this embodiment can be supplied by standard optical supply firms, and can be mounted in the set-up using similar mounting materials as used for the interferometric optics.

Now referring to Figure 26, In order to correct for artifacts introduced by having  
10 light pass through a non-flat region of transparent material, such as a substantially cylindrical fluid stream but including other geometries as well, embodiments of the present invention disclose the incorporation of a component similar in shape to the non-flat region, but opposite in terms of relative refractive indices. In the specific case of a flow cytometer this shape approximates a cylinder. To correct for artifacts introduced by  
15 the fact that the objects to be assessed are located within a cylindrical stream of water, is the incorporation of an optical component (58) which can be in the shape of a transparent cylinder, located inside transparent material (59) of a higher refractive index. It may be preferred that the image of the stream and of the compensation element fall on top of each other in the image plane. This can be done by locating the compensation element between  
20 the objective lens and the image plane, and by incorporating auxiliary lenses.

An embodiment of the optical component (58) can be located within a thin slice of transparent material of higher refractive index (59), for instance glass, or perspex, with a cylindrical hole drilled across it. Perspex has the advantage that it can be easier to drill a round channel into it. The cylindrical hole may be filled by a transparent material, the  
25 refractive index of which is lower than that of the surrounding material. The difference in refractive index between the substance and the surrounding material can be the same as but opposite to the difference in refractive index between the water in the stream and the surrounding air for certain applications. It may not be necessary to have the cylinder the same size as the stream of water, as long as magnification by the lenses used, makes the

resulting images in the image plane the same size. In some applications, it may be desired or necessary to adjust the refractive index difference to compensate for this magnification. Manufacturing of such element out of perspex can be quite simple, and can be done by most mechanical workshops that have experience with machining perspex or the selected  
5 material. It may be made in such dimensions that it fits in a standard optical mounting hardware, to facilitate incorporation into the optics.

Exactly matching the refractive indices may be difficult. An embodiment of the invention that facilitates adjustment can be to make the substance inside the perspex, or other selected material, a transparent refractive index fluid (58), as but one example, an  
10 organic oil, or mixture of oils that have a refractive index close to the desired one. Due to the fact that the refractive index of most fluids changes with temperature, much more so than solids, or glasses, it may be possible to fine-tune the difference in refractive index by temperature. This may be done by incorporating a temperature controller (60).

Optical component (58) of transparent fluids or refractive index fluids can be  
15 supplied by chemical supply firms. These firms often have data regarding the refractive index of their fluids readily available. Some firms even offer fluids that are specially made to serve as refractive index fluids, and have a guaranteed and stable refractive index. Temperature controllers and thermostats are supplied by many firms. A practical way to apply heat to the refractive index fluid can be to use a hollow mounting made of heat  
20 conducting material, a metal as but one example, containing the refractive index fluid. Using a conventional immersion thermostat cycler, found in many laboratories, water can be pumped through the mounting, thus keeping the element at a fixed and controllable temperature.

The discussion included in this PCT application is intended to serve as a basic  
25 description. The reader should be aware that the specific discussion may not explicitly describe all embodiments possible; many alternatives are implicit. It also may not fully explain the generic nature of the invention and may not explicitly show how each feature or element can actually be representative of a broader function or of a great variety of

alternative or equivalent elements. Again, these are implicitly included in this disclosure. Where the invention is described in functionally-oriented terminology, each aspect of the function is accomplished by a device, subroutine, or program. Apparatus claims may not only be included for the devices described, but also method or process claims may be  
5 included to address the functions the invention and each element performs. Neither the description nor the terminology is intended to limit the scope of the claims which now be included.

Further, each of the various elements of the invention and claims may also be achieved in a variety of manners. This disclosure should be understood to encompass  
10 each such variation, be it a variation of an embodiment of any apparatus embodiment, a method or process embodiment, or even merely a variation of any element of these. Particularly, it should be understood that as the disclosure relates to elements of the invention, the words for each element may be expressed by equivalent apparatus terms or method terms -- even if only the function or result is the same. Such equivalent, broader,  
15 or even more generic terms should be considered to be encompassed in the description of each element or action. Such terms can be substituted where desired to make explicit the implicitly broad coverage to which this invention is entitled. As but one example, it should be understood that all actions may be expressed as a means for taking that action or as an element which causes that action. Similarly, each physical element disclosed  
20 should be understood to encompass a disclosure of the action which that physical element facilitates. Regarding this last aspect, as but one example, the disclosure of a "droplet separator" should be understood to encompass disclosure of the act of "separating droplets" -- whether explicitly discussed or not -- and, conversely, were there only disclosure of the act of "converting liquid-gas", such a disclosure should be understood to  
25 encompass disclosure of a "droplet separator" and even a means for "separating droplets". Such changes and alternative terms are to be understood to be explicitly included in the description.

Additionally, the various combinations and permutations of all elements or applications can be created and presented. All can be done to optimize the design or

performance in a specific application.

Any acts of law, statutes, regulations, or rules mentioned in this application for patent: or patents, publications, or other references mentioned in this application for patent are hereby incorporated by reference. Specifically, United States Patent

5 Application Nos. 60/267571, 60/239,752, and 60/203,089 are each hereby incorporated by reference herein including any figures or attachments, and each of references in the following table of references are hereby incorporated by reference.

	DOCUMENT NO.	DATE	NAME	CLASS	SUBCLASS	FILING DATE
	32,350	02/10/87	Bhattacharya	204	180.1	11/22/74
10	3,687,806	08/29/72	Van den Bovenkamp	195	1.3	11/04/69
	3,829,216	08/13/74	Persidsky	356	36	10/02/72
	3,894,529	07/15/75	Shrimpton	128	1 R	04/10/69
	4,009,260	02/22/77	Ericcson	424	561	10/11/74
	4,067,965	01/10/78	Bhattacharya	424	105	12/17/75
15	4,083,957	04/11/78	Lang	424	78	02/04/76
	4,085,205	04/18/78	Hancock	424	105	01/24/77
	4,092,229	05/30/78	Bhattacharya	204	180 R	10/20/76
	4,155,831	05/22/79	Bhattacharya	207	299 R	02/23/78
	4,191,749	03/04/80	Bryant	424	105	10/11/77
20	4,225,405	09/30/80	Lawson	204	180 R	08/16/78
	4,276,139	06/30/81	Lawson	204	180 R	10/09/79
	4,339,434	07/13/82	Ericcson	424	561	08/17/81
	4,362,246	12/07/82	Adair	209	3.3	07/14/80
	4,448,767	05/15/84	Bryant	424	85	02/15/80
25	4,474,875	10/02/84	Shrimpton	435	002	08/18/80
	4,501,366	02/26/85	Thompson	209	556	12/14/82
	4,511,661	04/16/85	Goldberg	436	503	12/30/83
	4,605,558	08/12/86	Shrimpton	424	561	04/20/84
	4,660,971	04/28/87	Sage et al.	356	39	05/03/84
30	4,680,258	07/14/87	Hammerling et al	435	7	08/09/83
	4,698,142	10/06/87	Muroi et al	204	182.3	07/31/85
	4,749,458	06/07/88	Muroi et al	204	182.3	03/02/87
	4,988,619	01/29/91	Pinkel	435	30	11/30/87

	4,999,283	03/12/91	Zavos et al	435	2	08/18/89
	5,021,244	06/04/91	Spaulding	424	561	05/12/89
	5,135,759	08/04/92	Johnson	424	561	04/26/91
	5,346,990	09/13/94	Spaulding	530	350	03/12/91
5	5,371,585	12/06/94	Morgan et al.	356	246	11/10/92
	5,439,362	08/08/95	Spaulding	424	185.1	07/25/94
	5,466,572	11/14/95	Sasaki et al.	435	2	04/25/94
	5,483,469	01/09/96	Van den Engh et al.	364	555	08/02/93
	5,503,994	04/02/96	Shear et al.	436	90	10/08/93
10	5,514,537	05/07/96	Chandler	435	002	11/28/94
	5,589,457	12/31/96	Wiltbank	514	12	07-03-95
	5,602,039	02/11/97	Van den Engh	436	164	10/14/94
	5,602,349	02/11/97	Van den Engh	73	864.85	10/14/94
	5,660,997	08/26/97	Spaulding	435	7.21	06/07/95
15	5,690,895	11/25/97	Matsumoto et al.	422	73	12/06/96
	5,700,692	12/23/97	Sweet	436	50	09/27/94
	5,726,364	03/10/98	Van den Engh	73	864.85	02/10/97
	5,819,948	10/13/98	Van den Engh	209	158	08/21/97
	5,880,457	03/09/99	Tomiyama et al.	250	207	06/16/97
20	5,985,216	11/16/99	Rens, et al.	422	073	07/24/97
	6,071,689	06/06/00	Seidel et al.	435	2	01/29/98
	WO 96/12171	13/10/95				
	WO 98/34094	06/08/98				
	WO 99/05504	07/24/98				
25	WO 99/33956	08/07/99				
	WO 99/38883	05/08/99				
	WO 99/42810	26/08/99				
	WO 00/06193	10/02/00				

	Akhtar, S., et al., "Sex Preselected in Cattle: a Field Trial ", Veterinary Record 136, 1995, p. 495-496.
30	Akhtar, S., et al., "Prevalence of Five Stereotypes of Bluetongue Virus in a Rambouillet Sheep Flock in Pakistan", Veterinary Record 136, 1995, p. 495.
	Amann, R.P. et al, "Prospects For Sexing Mammalian Sperm," Colorado Associated University Press, Animal Reproduction Laboratory College of Veterinary Medicine and Biomedical Sciences, Colorado State University, Fort Collins, CO, 80523, 1982
35	Amoah, E.A. and Gelaye, S. 1996. Biotechnological advances in goat reproduction. J. Anim. Sci. 75(2):578-585.



	Anderson, V.K., Aamdal, J. and Fougner, J.A. 1973. Intrauterine und tiefzervikale Insemination mit Gefriersperma bein Schat. Zuchthygiene. 8:113-118.
	Baker, R.D., Dziuk, P.J. and Norton, H.W. 1968. Effect of volume of semen, number of sperm and drugs on transport of sperm in artificially inseminated gilts. J. Anim. Sci. 27:88-93.
5	Barnes, F.L. and Eyestone, W.H., "Early Cleavage and the Maternal Zygotic Transition in Bovine Embryos", Theriogenology, Vol. 33, No. 1, January 1990, pp. 141-149
	Becker, S.E. and Johnson, A.L. 1992. Effects of gonadotropin releasing hormone infused in a pulsatile or continuous fashion on serum gonadotropin concentrations and ovulation in the mare. J. Anim. Sci. 70:1208-1215.
10	Bedford, S. J. and Hinrichs, K. 1994. The effect of insemination volume on pregnancy rates of pony mares. Theriogenology 42:571-578.
	Berger, G.S. 1987. Intratubal insemination. Fert. Steril. 48:328-330.
	Beyhan, Z., et al., "Sexual Dimorphism in IVM-IVF Bovine Embryos Produced from X and Y Chromosome-bearing Spermatozoa Sorted by High Speed Flow Cytometry", Theriogenology 52, 1999, pp. 35-48.
15	Blanchard, T. and Dickson, V., "Stallion Management", The Veterinary Clinics of North America, Equine Practice, Vol. 8, No. 1, April 1992, pp 207 - 218.
	Bracher, V. and Allen, W.R., "Videoendoscopic Examination of the Mare's Uterus: Findings in Normal Fertile Mares", Equine Veterinary Journal, Vol. 24 (1992), pp. 274-278
	Braselton, W.E. and McShan, W.H. 1970. Purification and properties of follicle stimulating and luteinizing hormones from horse pituitary glands. Arch. Biochem. Biophys. 139:45-48.
20	Brethour, J.R. and Jaeger, J.R., "The Single Calf Heifer System", Kansas Agric. Sta. Rep of Progress 570, 1989.
	Bristol, S.P. 1982. Breeding behavior of a stallion at pasture with 20 mares in synchronized oestrus. J. Reprod. Fert. Suppl. 32:71.
	Buchanan, B.R., et al, "Insemination of Mares with Low Numbers of Either Unsexed or Sexed Spermatozoa", Theriogenology, Vol. 53, pp 1333-1344, (2000)
25	Burwash, L.D., Pickett, B.W., Voss, J.L. and Back, D.G. 1974. Relationship of duration of estrus to pregnancy rate in normally cycling, non-lactating mares. J.A.V.M.A. 165:714-716.
	Caslick, E.A., "The Vulva and the Vulvo-vaginal Orifice and its Relation to Genital Health of the Thoroughbred Mare", Cornell Veterinarian, Vol. 27, 1937, pp. 178-187
30	Catt, et al., "Assessment of Ram and Boar Spermatozoa During Cell-Sorting by Flow Cytometry", Reproduction Dom Animal, Vol. 32, 1997, pp 251-258.
	Catt, S.L., et al., "Birth of a Male Lamb Derived from an In Vitro Matured Oocyte Fertilized by Intracytoplasmic Injection of a Single Presumptive Male Sperm", Veterinary Record 139, 1996, pp. 494-495.
	Chandler, J.E., et al, "Bovine Spermatozoal Head Size Variation and Evaluation of a Separation Technique Based on this Size", Theriogenology 52, p. 1021-1034 (1999)
35	Chandler, J.E., "Videomicroscopic Comparison of Bull Sperm and Leukocyte Chromosome Areas as Related to Gender", J Dairy Sci 73, pp. 2129-2135, (1990)
	Chin, W.W. and Boime, I. 1990. In: Glycoprotein Hormones. Serona Symp. Norwell, MA. pp. 19-20
	Chung, Y.G., Schenk, J.L., Herickhoff, L.A. and Seidel, G.E. Jr. 1998. Artificial insemination of superovulated heifers with 600,000 sexed sperm. J Anim. Sci. Suppl. 1. 836:215. abstr.

	Clement, F., Vincent, P., Mahla, R., Meriaux, J.C. and Palmer, E. 1998. Which insemination fertilizes when several successive inseminations are performed before ovulation. 7 <sup>th</sup> Int. Symp. Eq. Repro. 151. abstr.
	Cran, D.G., et al, "Production of Lambs by Low Dose Intrauterine Insemination with Flow Cytometrically Sorted and Unsorted Semen", <i>Theriogenology</i> , Vol. 47, pp. 267, (Abstract), (1997)
5	Cran, D.G., et al., "Production of Bovine Calves Following Separation of X- and Y- Chromosome Bearing Sperm and In Vitro Fertilisation", <i>Veterinary Record</i> 132, 1993, pp. 40-41.
	Cran, D.G., Johnson, L.A. and Polge, C., 1995, "Sex preselection in cattle: a field trial", <i>Vet. Rec.</i> 136:495-496.
	Cui, K., "Size Differences between human X and Y Spermatozoa and prefertilization diagnosis", <i>Molecular Human Reproduction</i> , Vol. 3, No. 1, pp. 61-67, (1997)
10	Cui, K., "X Larger than Y", <i>Nature</i> 366, p. 177-118, (1993)
	Curran, S. 1998. In: <i>Equine Diagnostic Ultrasonography. Fetal gender determination.</i> Rantanen & McKinnon. 1 <sup>st</sup> Ed. Williams and Wilkins. pp. 165-169.
15	Day, B.N., Abeydeera, L.R., Johnson, L.A., Welch, G.R., Wang, W.H., Cantley, T.C. and Rieke, A. 1998. Birth of piglets preselected for gender following in vitro fertilization of in vitro matured pig oocytes by X and Y bearing spermatozoa sorted by high speed flow cytometry. <i>Theriogenology</i> . 49(1):360. abstr.
	Dean, P.N., Pinkel, D. and Mendelsob. n, M.L. 1978. Hydrodynamic orientation of spermatozoa heads for flow cytometry. <i>Biophys. J.</i> 23:7-13.
	Demick, D.S., Voss, J.L. and Pickett, B.W. 1976. Effect of cooling, storage, glycerization and spermatozoal numbers on equine fertility. <i>J. Anim. Sci.</i> 43:633-637.
20	DenDaas, J.H.G., De Jong, G., Lansbergen, L.M.T.E. and Van Wagtenonk-De Leeuw, A.M. 1998. The relationship between the number of spermatozoa inseminated and the reproductive efficiency of dairy bulls. <i>J Dairy Sci.</i> 81: 1714-1723.
	Dinnyes, A., et al., "Timing of the First Cleavage Post-insemination Affects Cryosurvival of In Vitro-produced Bovine Blastocysts", <i>Molec Reprod develop</i> 53, 1999, pp 318-324.
25	Donaldson, L. E., "Effect of Insemination Regimen on Embryo Production in Superovulated Cows", <i>The Veterinary Record</i> , July 13, 1985, pp. 35-37
	Donoghue, et al, 1996. "Timing of ovulation after gonadotropin induction and its importance to successful intrauterine insemination in the tiger ( <i>Panthera tigris</i> )", <i>J. Reprod. Fert.</i> 107:53-58.
	Douglas, R.H. 1979. "Review of superovulation and embryo transfer in the equine", <i>Theriogenology</i> . 11:33-46.
30	Douglas, R.H., Nuti, L. and Ginther, O.J. 1974. Induction of ovulation and multiple ovulation on seasonally-anovulatory mares with equine pituitary fractions. <i>Theriogenology</i> . 2(6): 133-142.
	Duchamp, G., Bour, B., Combarnous, Y. and Palmer, E. 1987. Alternative solutions to hCG induction of ovulation in the mare. <i>J. Reprod. Fert. Suppl.</i> 35:221-228.
35	Evans, M.J. and Irvine, C.H.G. 1977. Induction of follicular development, maturation and ovulation by gonadotropin releasing hormone administration to acyclic mares. <i>Bio. Reprod.</i> 16:452-462.
	Fitzgerald, B.P., Peterson, K.D. and Silvia, P.J. 1993. Effect of constant administration of a gonadotropin-releasing hormone agonist on reproductive activity in mares: Preliminary evidence on suppression of ovulation during the breeding season. <i>Am. J. Vet. Res.</i> 54:1746-1751.

	Fluharty, F.L., et al., "Effects of Age at Weaning and Diet on Growth of Calves", Ohio Agri. Res. and Dev. Circular, 1996, 156: 29.
	Foulkes, J.A., et al., 1977. "Artificial insemination of cattle using varying numbers of spermatozoa", Vet. Rec. 101:205.
5	Francon, M. and Yamamoto, T., "Un Nouveau et tres simple dispositif interferentiel applicable as microscope", Optica Acta 9, p. 395-408 (1962)
	Fugger, E.F., "Clinical Experience with Flow Cytometric Separation of Human X- and Y- Chromosome Bearing Sperm", Theriogenology, Vol. 52, pp. 1435-1440 (1999)
	Fulwyler, M.J. 1965. "Electronic separation of biological cells by volume", Science. 150:910.
10	Fulwyler, M.J. 1977. "Hydrodynamic orientation of cells", J Histochem. Cytochem. 25:781-783.
	Garner, D.L., et al., 1983. "Quantification of the X and Y chromosome-bearing spermatozoa of domestic animals by flow cytometry", Biol. Reprod. 28:312-321.
	Ginther, O.J. 1971. "Some factors which alter estrus cycle in mares", J. Anim. Sci. 33:1158. abstr.
15	Ginther, O.J. 1983. "Sexual behavior following introduction of a stallion into a group of mares", Theriogenology. 19:877.
	Ginther, O.J. 1992. "In: Reproductive Biology of the Mare", (2 <sup>nd</sup> Ed.) Equiservices, Cross Plains, WI.
	Gledhill, B.L. 1988. Gender preselection: historical, technical and ethical perspective. Semin Reprod. Endocrinol. 6:385-395.
20	Gourley, D.D. and Riese, R.L. 1990. Laparoscopic artificial insemination in sheep. Vet. Clin. N. Amer: Food Anim. Prac. 6(3):615-633.
	Grondahl, C., et al, "In Vitro Production of Equine Embryos", Biology of Reproduction, Monograph Series I, pp. 299-307 (1995)
	Guillou, F. and Combamous, Y. 1983. Purification of equine gonadotropins and comparative study of their acid-dissociation and receptor-binding specificity. Biochem. Biophys. Acta. 755:229-236.
25	Gurnsey, M.P., and Johnson, L.A., "Recent improvements in efficiency of flow cytometric sorting of X and Y-chromosome bearing sperm of domestic animals: a review", 1998, New Zealand Society of Animal Protection, three pages.
	HAMAMATSU, Photomultiplier Tubes", web page, <a href="http://www.optics.org/hamamatsu/pmt.html">http://www.optics.org/hamamatsu/pmt.html</a> , printed out on 4/15/00, four pages total.
30	HAMAMATSU, Technical Information", web page, <a href="http://www.optics.org/hamamatsu/photodiode.html">http://www.optics.org/hamamatsu/photodiode.html</a> , printed out on 4/15/00, six pages total.
	Hamano, K., et al., "Gender Preselection in Cattle with Intracytoplasmically Injected, Flow Cytometrically Sorted Sperm Heads", Biology of Reproduction 60, 1999, pp. 1194-1197.
35	Harrison, L.A., et al., 1991. Comparison of hCG, buserelin and luprostiol for induction of ovulation in cycling mares. Eq. Vet. Sci. 3:163-166.
	Hawk, H.W., et al., "Fertilization Rates in Superovulating Cows After Deposition of Semen on the Infundibulum Near the Uterotubal Junction or After Insemination with High Numbers of Sperm", Theriogenology, May 1988, Vol. 29, No. 5, pp 1131-1142.

	Hofferer, S., Lecompte, F., Magallon, T., Palmer, E. and Combamous, Y. 1993. Induction of ovulation and superovulation in mares using equine LH and FSH separated by hydrophobic interaction chromatography. <i>J. Reprod. Fert.</i> 98:597-602.
5	Holtan, D.W., Douglas, R.H. and Ginther, O.J. 1977. Estrus, ovulation and conception following synchronization with progesterone, prostaglandin F2 ct and human chorionic gonadotropin in pony mares. <i>J. Anim. Sci.</i> 44:431-437.
	Householder, D.D., Pickett, B.W., Voss, J.L. and Olar, T.T. 1981. Effect of extender, number of spermatozoa and hCG on equine fertility. <i>J. Equine Vet. Sci.</i> 1:9-13.
	Howard, J.G., et al., 1991. "Comparative semen cryopreservation in ferrets ( <i>Mustela putorius furo</i> ) and pregnancies after laparoscopic intrauterine insemination with frozen-thawed spermatozoa". <i>J. Reprod. Fert.</i> 92:109-118.
10	Howard, J.G., et al., 1997. "Sensitivity to exogenous gonadotropins for ovulation and laparoscopic artificial insemination in the cheetah and clouded leopard". <i>Biol. Reprod.</i> 56:1059-1068.
	Hunter, R.H.F. 1980. Transport and storage of spermatozoa in the female reproductive tract. <i>Proc 4<sup>th</sup> Int. Congr. Artira. Repro. and A.I.</i> 9:227-233.
15	Hyland, J.H., et al., 1988. "Gonadotropin-releasing hormone (GnRH) delivered by continuous infusion induces fertile estrus in mares during seasonal acyclicity". <i>Proc. Amer. Assoc. Eq. Prac.</i> 181-190.
	Irvine, C.H.G. and Alexander, S.L., In: <i>Equine Reproduction</i> . Edited by McKirmon and Voss. Lea and Febiger. Philadelphia, London. pp. 37. (1993)
	Jafar, et al., "Sex Selection in Mammals: A Review", <i>Theriogenology</i> , Vol. 46, pp 191-200 (1996)
20	Jasko, D.J., Martin, J.M. and Squires, E.L. 1992. Effect of volume and concentration of spermatozoa on embryo recovery in mares. <i>Theriogenology</i> . 37:1233-1239
	Johnson, A.L. 1986. "Pulsatile release of gonadotropin releasing hormone advances ovulation in cycling mares". <i>Biol. Reprod.</i> 35:1123-1130.
	Johnson, A.L. and Becker, S.E. 1988. "Use of gonadotropin-releasing hormone (GnRH) treatment to induce multiple ovulations in the anestrous mare", <i>Eq. Vet. Sci.</i> 8:130-134. (1988)
25	Johnson, L.A., "Advances in Gender Preselection in Swine" <i>Journal of Reproduction and Fertility Supplement</i> , Vol. 52, pp. 255-266 (1997)
	Johnson, L.A., "Sex Preselection in Swine: Altered Sex Ratios in Offspring Following Surgical Insemination of Flow Sorted X- and Y- Bearing Sperm", <i>Reproduction in Domestic Animals</i> , Vol. 26, pp. 309-314 (1991)
30	Johnson, L.A. and Welch, G.R. "Sex Preselection: High-speed flow cytometric sorting of X and Y sperm for maximum efficiency", <i>Theriogenology</i> , Vol. 52, (1999), pp. 1323-1341
	Johnson, L.A. and Schulman, J.D. "The safety of sperm selection by flow cytometry", <i>Hum. Reprod.</i> 9(5):758. (1994)
	Johnson, L.A., and Pinkel, D., "Modification of a Laser-Based flow Cytometer for High-Resolution DNA Analysis of Mammalian Spermatozoa", <i>Cytometry</i> 7, pp 268 - 273. (1986)
35	Johnson, L.A., et al., "Sex Preselection in Rabbits: Live Births from X and Y Sperm Separated by DNA and Cell Sorting", <i>Exceptional Paper-Rapid Publication</i> , XP-002103476, <i>Biology of Reproduction</i> 41, pp. 199-203 (1999)
	Johnson, L.A., et al., "Flow sorting of X and Y chromosome-bearing sperm for DNA using an improved preparation method and staining with Hoechst 33342", <i>Gam. Res.</i> 17:1-9., (1987)

	Johnson, L.A., et al, "Enhanced flow cytometric sorting of mammalian X and Y sperm: high speed sorting and orienting no77.1e for artificial insemination. <i>Theriogenology</i> . 49(1):361. abstr. (1988)
	Johnson, L.A., et al, "Sex Preselection in Swine: Flow Cytometric Sorting of X- and Y- Chromosome Bearing Sperm to Produce Offspring", <i>Boar Semen Preservation IV</i> , pp. 107-114 (2000)
5	Johnson, L.A., et al, "Improved flow sorting resolution of X- and Y- chromosome bearing viable sperm separation using dual staining and dead cell gating. <i>Cytometry</i> 17 (suppl 7):83. (1989)
	Johnson, L.A., "Flow cytometric determination of spermatozoa sex ratio in semen purportedly enriched for X or Y bearing spermatozoa. <i>Theriogenology</i> . 29:265. abstr. (1988)
10	Johnson, L.A. "Gender preselection in domestic animals using flow cytometrically sorted sperm. <i>J Anim. Sci. Suppl</i> 1.70:8-18. (1992)
	Johnson, L.A., "Gender preselection in Mammals: An overview", <i>Deutsch. Tierarztl. Wschr</i> , Vol. 103, pp 288-291 (1996)
	Johnson, L.A. "Isolation of X- and Y-bearing spermatozoa for sex preselection. <i>In: Oxford Reviews of Reproductive Biology</i> . Ed. HH Charlton. Oxford University Press. 303-326 (1994)
15	Johnson, L.A., "Successful Gender Preselection in Farm Animals", <i>Agricultural Biotechnology</i> , 1998, pp. 439-452.
	Kachel, V., et al, "Uniform Lateral Orientation, Cused by Flow Forces, of Flat Particles in Flow-Through Systems", <i>The Journal of Histochemistry and Cytochemistry</i> , 1997, Vol. 25, No. 7, pp 774 -780.
	Kanayama, K., et al.. "Pregnancy by means of tubal insemination and subsequent spontaneous pregnancy in rabbits", <i>J. Int. Med. Res</i> . 20:401-405. (1992)
20	Karabinus, et al., "Effects of Egg Yolk-Citrate and Milk Extenders on Chromatin Structured Viability of Cryopreserved Bull Sperm", <i>Journal of Dairy Science</i> , Vol. 74, No. 11, 1999, pp 3836-3848.
	Kilicarslan, M.R., Horoz, H., Senunver, S.C., Konuk, S.C., Tek, C. and Carioglu, B. 1996. Effect of GnRH and hCG on ovulation and pregnancy in mares. <i>Vet. Rec</i> . 139:119-120.
25	Lapin, D.R. and Ginther, O.J. 1977. Induction of ovulation and multiple ovulations in seasonally anovulatory and ovulatory mares with an equine pituitary extract. <i>J. Anim. Sci</i> . 44:834-842.
	Lawrenz, R. 1985. Preliminary results of non-surgical intrauterine insemination of sheep with thawed frozen semen. <i>J S Afr. Vet. Assoc</i> . 56(2):61-63.
	Levinson, G., et al, "DNA-based X-enriched sperm separation as an adjunct to preimplantation genetic testing for the preparation of X-linked disease", <i>Mol. Human Reprod</i> . 10:979-982. (1995)
30	Lindsey, A., et al., "Hysteroscopic Insemination of Mares with Nonfrozen Low-dose Unsexed or Sex-sorted Spermatozoa", Currently unpublished, pp. 1-15.
	Linge, F. "Faltforsok med djupfrost sperma (field trials with frozen sperm)", <i>Farskotsel</i> . 52:12-13. (1972)
	Lonergan, P., et al., "Effect of Time Interval from Insemination to First Cleavage on the Development of Bovine Embryos In Vitro and In Vivo", <i>Theriogenology</i> , 1999, p. 326
35	Long, C.R., et al, <i>Theriogenology</i> . 49(1):363. abstr. (1998)
	Loy, R.G. and Hughes, J.P. "The effects of human chorionic gonadotropin on ovulation, length of estrus, and fertility in the mare". <i>Cornell Vet</i> . 56:41-50. (1965)
	Lu, K.H., et al., "In Vitro Fertilization with Flow-Cytometrically-Sorted Bovine Sperm", <i>Theriogenology</i> 52, 1999, pp. 1393-1405.

	Macmillan, K.L. and A.M. Day, "Prostaglandin F2a - A Fertility Drug In Dairy Cattle?", Ruakura Animal Research Station, Private Bag, Hamilton, New Zealand, Theriogenology, September 1982, Vol. 18 No. 3, pages 245-253
	Matsuda, Y. and Tobari, I. "Chromosomal analysis in mouse eggs fertilized <i>in vitro</i> with sperm exposed to ultraviolet light (UV) and methyl and ethyl methanesulfonate (MMS and EMS)", Mutat. Res. 198:131-144. (1988)
5	Maxwell, W. and Johnson, L., "Chlortetracycline Analysis of Boar Spermatozoa after Incubation, Flow Cytometric Sorting, Cooling, or Cryopreservation", Molecular Reproduction and Development 46, 1997, pp. 408-418.
	Maxwell, W., et al., "Fertility of Superovulated Ewes after Intrauterine or Oviductal Insemination with Low Numbers of Fresh or Frozen-Thawed Spermatozoa", Reprod. Fertil. Dev. 5:57-63. (1993)
	McCue, P.M., et al., "Oviductal insemination in the mare", 7 <sup>th</sup> Int Symp. Eq. Reprod. 133. abstr. (1997)
10	McCue, P.M., "Superovulation", Vet. Clin. N. Amer. Eq. Prac. 12:1-11. (1996)
	McDonald, L.E., "Hormones of the pituitary gland", In: Veterinary Pharmacology and Therapeutics. 6 <sup>th</sup> ed., Ames, Iowa State Univ. Press. pp. 590. (1988)
	McKeuna, T., et al., "Nonreturn rates of dairy cattle following uterine body or conual insemination", J. Dairy Sci. 73:1179-1783. (1990)
15	McKinnon, A. and Voss, J., "Equine Reproduction" Lea & Febiger, Philadelphia, pp 291, 299 - 302, 345 - 348, 739 - 797. (1993)
	McKinnon, A. et al., "Predictable ovulation in mares treated with an implant of the GnRH analogue deslorelin", Eq. Vet. J. 25:321-323. (1993)
20	McKinnon, A.O. et al., "Repeated use of a GnRH analogue deslorelin (Ovuplant) for hastening ovulation in the transitional mare", Eq. Vet. J. 29:153-155. (1996)
	McNutt, T.L. and Johnson, L.A., "Flow cytometric sorting of sperm: influence on fertilization and embryo/fetal development in the rabbit", Mol. Reprod. Dev. 43:261-267. (1996)
	Meinert, C., et al., "Advancing the time of ovulation in the mare with a short-term implant releasing the GnRH analogue deslorelin", Equine Veterinary Journal, 25, 1993, pp 65 - 68.
25	Merton, J., et al., "Effect of Flow Cytometrically Sorted Frozen/Thawed Semen on Success Rate of In Vitro Bovine Embryo Production", Theriogenology 47, 1997, pp. 295.
	Meyers, P.J., Bowman, T., Blodgett, G., Conboy, H.S., Gimenez, T., Reid, M.P., Taylor, B.C., Thayer, J., Jochle, W. and Trigg, T.E. 1997. Use of the GnRH analogue, deslorelin acetate, in a slow release implant to accelerate ovulation in oestrous mares. Vet. Rec. 140:249-252.
30	Michaels, Charles, "Beef A.I. Facilities that work", Proc. Fifth N.A.A.B Tech. Conf. A.I. Reprod. Columbia, MO. pp. 20-22.
	Michel, T.H., et al., "Efficacy of human chorionic gonadotrophin and gonadatrophin releasing hormone for hastening ovulation in Thoroughbred mares", Eq. Vet. J. 6:438-442. (1986)
35	Miller, S.J., "Artificial Breeding Techniques in Sheep", In Morrow D.A. (ed): Current Therapy in Theriogenology 2 Philadelphia, WB Saunders. (1986)
	Mirskaja, L.M. and Petrapavlovskii, V.V., "The reproduction of normal duration of heat in the mare by the administration of Prolan", Probl. Zivotn. Anim. Breed. Abstr. 5:387. (1937)

	Molinia, F.C., Gibson, R.J., Brown, A.M., Glazier, A.M. and Rodger, J.C. 1998. Successful fertilization after superovulation and laparoscopic intrauterine insemination of the brushtail possum, <i>Trichosurus vulpecula</i> , and tammar wallaby, <i>Macropus eugenii</i> . J.Reprod. Fert. 112:9-17.
5	Morcom, C.B. and Dukelow, W.R. "A research technique for the oviductal insemination of pigs using laparoscopy", Lab. Anim. Sci. 1030-1031. (1980)
	Morris, L.H., et al., "Hysteroscopic insemination of small numbers of spermatozoa at the uterotubal junction of preovulatory mares", Journal of Reproduction and Fertility, Vol. 118, pp. 95-100 (2000)
	Mullet, W. and Gautier, F., "Interactions of heteroaromatic compounds with nucleic acids" Euro. J Biochem. 54:358 (1975)
10	Munne, S., "Flow cytometry separation of X and Y spermatozoa could be detrimental to human embryos", Hum. Reprod. 9(5):758, (1984)
	Nowshari, et al., "Superovulation of Goats with Purified pFSH Supplemented with Defined Amounts of pLH", Theriogenology, Vol. 43, pp 797-802 (1995)
15	Olson, S.E. and Seidel, G.E. Jr., "Reduced Oxygen Tension and EDTA improve Bovine Zygote Development in a Chemically Defined Medium", Journal of Animal Science 78, 2000, pp. 152-157.
	Pace, M.M. and Sullivan, J.J., "Effect of timing of insemination, numbers of spermatozoa and extender components on pregnancy rates in mares inseminated with frozen stallion semen", J Reprod. Fert. Suppl. 23:115-121 (1975)
	Parrish, J.J., et al., "Capacitation of bovine sperm by heparin", Biology of Reproduction, Vol. 38, pp. 1171-1180 (1988)
20	PCT application PCT/US99/17165, filed 28 July 1999, entitled "Equine System for Non-Surgical Artificial Insemination".
	PCT application PCT/US98/27909, filed 31 December 1998, entitled "Commercially Practical Sex-Specific Insemination of Mammals".
25	Peippo, J., et al., "Sex diagnosis of equine preimplantation embryos using the polymerase chain reaction", Theriogenology, Vol. 44 619-627 (1995)
	Perry, E.J. "Historical Background In: The Artificial Insemination of Farm Animals", 4 <sup>th</sup> ed. Edited by E.J. Perry. New Brunswick, Rutgers University Press, pp. 3-12., (1968)
	Petersen, G.A., et al, "Cow and Calf Performance and Economic Considerations of Early Weaning of Fall-Born Beef Calves", J. Anim. Sci., 1987, 64:15, pp 15-22.
30	Pickett, B.W., et al., "Factors influencing the fertility of stallion spermatozoa in an A.I. program", Proc. 8 <sup>th</sup> Internat. Congr. Anim. Reprod. A.I. Krakow, Poland. 4: 1049 - 1052. (1976)
	Pickett, B.W., and Shiner, K.A., "Recent developments in artificial insemination in horses", Livestock Production Science, 40, 1994, pp 31 - 36.
35	Pickett, B.W. and Back, D.G., "Procedures for preparation, collection, evaluation and insemination of stallion semen". C.S.U. Exp. Sta. Artira. Reprod. Lab. Gen. Series Bull. 935. (1973)
	Pickett, B.W., et al., "Effect of seminal extenders on equine fertility", J. Anim. Sci. 40:1136-1143. (1975)
	Pickett, B.W., et al., "Influence of seminal additives and packaging systems on fertility of bovine spermatozoa", J. Anim. Sci. Suppl. II. 47:12. (1978)

	Pickett, G.W., et al., "Management of the mare for maximum reproductive efficiency" Bulletin No. 6 Colorado State University, Ft. Collins CO. (1989)
	Pinkel, D., et al., "High resolution DNA measurements of mammalian spermatozoa". Cytometry. 3:1-9. (1982)
5	Pinkel, D., et al., "Flow Cytometric Determination of the Proportions of X- and Y- Chromosome-Bearing Sperm in Samples of Purportedly Separated Bull Sperm", Journal of Animal Science, Vol. 60, No. 5, 1985, pp 1303 - 1307.
	Polge, E. J., "Historical Perspective of AI: Commercial Methods of Producing Sex Specific Semen, IVF Procedures", Proceedings of the 16 <sup>th</sup> Technical Conference on Artificial Insemination & Reproduction, Cambridge, England, 1996, pp. 7-11.
10	Preza, C. et al., "Determination of Direction-Independent Optical Path-Length Distribution of Cells Using Rotational-Diversity Transmitted-Light Differential Interference Contrast (DIC) Images", Presented at the Multidimensional Microscopy: Image Acquisition and Processing V, p. 1-11 (1998)
	Rath, D., et al., "Production of Piglets Preselected for Sex Following in Vitro Fertilization with X and Y Chromosome-Bearing Spermatozoa Sorted by Flow Cytometry", Theriogenology, 47, 1997, pp 795 - 800.
	Rath, D., et al., "Low Dose Insemination Technique in the Pig", Boar Semen Preservation IV, 2000, pp. 115-118.
15	Reiling, B.A., et al., "Effect of Prenatal Androgenization on Performance, Location, and Carcass and Sensory Traits on Heifers in Single Calf Heifer System", J. Anim. Sci., 1995, 73: pp 986-992.
	Rens, W., et al., "Improved Flow Cytometric Sorting of X- and Y- Chromosome Bearing Sperm: Substantial Increase in Yield of Sexed Semen", Molecular Reproduction and Development, 1999, pp 50-56.
20	Rens, W., et al., "A Novel Nozzle for More Efficient Sperm Orientation to Improve Sorting Efficiency of X and Y Chromosome-Bearing Sperm", Technical Notes, Cytometry 33, 1998, pp 476-481.
	Rieger, D., et al., "The Relationship Between the Time of First Cleavage of Fertilized Cattle Oocytes and Their Development to the Blastocyst Stage", Theriogenology, 1999, pp. 190.
	Ritar, A. and Ball, A. "Fertility of young cashmere goats after laparoscopic insemination". J. Agr. Sci. 117:271-273. (1991)
25	Roberts, J.R. In: Veterinary Obstetrics and Genital Diseases. Ithaca, New York. pp. 740-749. (1971)
	Roser, JF., et al., "Reproductive efficiency in mares with anti-hCG antibodies", Proc 9 <sup>th</sup> Int. Congr. Artira. Repro. and A.I. 4:627. abstr. (1980)
30	Roth, T.L., et al., "Effects of equine chorionic gonadotropin, human chorionic gonadotropin, and laparoscopic artificial insemination on embryo, endocrine, and luteal characteristics in the domestic cat", Bio Reprod. 57:165-171. (1997)
	Rowley, H-S., et al., "Effect of insemination volume on embryo recover}' in mares", J. Equine Vet. Sci. 10:298-300. (1990)
	Salamon, S. "Artificial Insemination of Sheep". Chippendale, New South Wales. Publicity Press. p.83-84. (1976)
35	Salisbury, G.W. and VanDemark, N.L. "Physiology of Reproduction and Artificial Insemination of Cattle", San Francisco: Freeman and Company. (1961)
	SAS, SAS/STAT® User's Guide (Release 6.03), SAS Inst. Inc., Cary, NC., 1988. 3 pages
	Schenk, J.L. and Seidel, Jr., G.E., "Imminent Commercialization of Sexed Bovine", Proceedings, The Range Beef Cow Symposium XVI, 1999, pp 89-96.



	Schenk, J.L., et al., "Cryopreservation of Flow-Sorted Bovine Spermatozoa", <i>Theriogenology</i> 52, 1999, pp. 1375-1391.
	Schmid R.L., et al, "Fertilization with Sexed Equine Spermatozoa Using Intracytoplasmic Sperm Injection and Oviductal Insemination ", 7th International Symposium On Equine Reproduction, pp. 139 (Abstract) (1998)
5	Seidel, G. Jr., "Use of Sexed Bovine Sperm for In Vitro Fertilization and Superovulation", <i>Animal Reproduction and Biotechnology Laboratory, Colorado State University, Proceedings of the 2000 CETA/ACTE Convention, Charlottetown, Prince Edward Island, August 2000, pp. 22-24.</i>
	Seidel, G.E. and Johnson, L.A., "Sexing Mammalian Sperm - Overview", <i>Theriogenology</i> 52: 1267-1272, (1999)
10	Seidel, G.E. Jr, et al., "Insemination of Heifers with Sexed Sperm ", <i>Theriogenology</i> , Vol. 52, pp. 1407-1421 (1999)
	Seidel, G.E. Jr, et al., "Artificial Insemination of Heifers with Cooled, Unfrozen Sexed Semen ", <i>Theriogenology</i> , Vol. 49 pp. 365 (Abstract) (1998)
	Seidel, G.E. Jr., et al, "Insemination of heifers with sexed frozen or sexed liquid semen", <i>Theriogenology</i> . 51. (in press). abstr. (1999)
15	Seidel, G.E., Jr., et al, "Uterine Horn Insemination of Heifers With Very Low Numbers of Nonfrozen and Sexed Spermatozoa", <i>Animal Reproduction and Biotechnology Laboratory Colorado State University, Atlantic Breeders Cooperative, Theriogenology</i> (1997), pp. 1255-1264
	Seidel, G.E., "Status of Sexing Semen for Beef Cattle", <i>Texas A&amp;M University 45th Annual Beef Cattle Short Course and Trade Show Proceedings, August 9-11, 1999; pp. III 24-III 27</i>
20	Seidel, Jr., G. E., "Artificial Insemination With X-and Y-Bearing Bovine Sperm", <i>Animal Reproduction and Biotechnology Laboratory, Colorado State University, (1996)</i>
	Seidel, Jr., G. E., et al, "Insemination of Holstein Heifers With Very Low Numbers Of Unfrozen Spermatozoa", <i>Colorado State University, Atlantic Breeders Cooperative, (1995)</i>
25	Senger, P.L., et al., "Influence of comual insemination on conception rates in dairy cattle". <i>J Anim. Sci.</i> 66:3010-3016. (1988)
	Shelton, J.N. and Moore, N.W. 1967. The response of the ewe tot pregnant mare gonadotropin and to horse anterior pituitary extract. <i>J. Reprod. Fert.</i> 14:175 - 177.
	Shilova, A.V., Platov, E.M. and Lebedev, S.G. 1976. The use of human chorionic gonadotrophin for ovulation date regulation in mares. <i>VIIIth Int. Congr. On Anim. Repro. and A.I.</i> 204-208.
30	Squires, E., "Simultaneous Analysis of Multiple Sperm Attributes by Flow Cytometry", <i>Diagnostic Techniques and Assisted Reproductive Technology, The Veterinary Clinics of North America, Equine Practice</i> , Vol. 12, No. 1, April 1996, pp127 - 130.
	Squires, E.L., et al., "Effect of dose of GnRH analogue on ovulation in mares", <i>Theriogenology</i> . 41:757-769. (1994)
35	Squires, E.L., "Early Embryonic Loss" in <i>Equine Diagnostic Ultrasonography</i> , 1 <sup>st</sup> Ed. pp 157-163 Eds Rantanen & McKinnon. Williams and Wilkins, Baltimore, Maryland (1998)
	Squires, E.L., et al, "Cooled and frozen stallion semen", <i>Bulletin No. 9, Colorado State University, Ft. Collins, CO.</i> (1999)
	Sullivan, J.J., Parker, W.G. and Larson, LL. 1973. Duration of estrus and ovulation time in nonlactating mares given human chorionic gonadotropin during three successive estrous periods. <i>J.A.V.M.A.</i> 162:895-898.

	Sumner, A.T. and Robinson, J.A., "A Difference in Dry mass between the heads of X and Y-bearing human Spermatozoa", J Reprod Fert 48, p. 9-15 (1976)
	Taljaard, T.L., Terblanche, S.J., Bertschinger, H.J. and Van Vuuren, L.J. 1991. The effect of the laparoscopic insemination technique on the oestrus cycle of the ewe. J. S Afr. Vet. Assoc. 62(2):60-61.
5	Taylor, C.S., "Efficiency of Food Utilization in Traditional and Sex-Controlled Systems of Beef Production", AFRC Animal Breeding Research Organization, West Mains Road, Edinburg EH9 3JQ, pp 401-440.
	Tervit, H.R., et al., "Successful Culture In Vitro of Sheep and Cattle Ova", Agricultural Research Council, Unit of Reproduction Physiology and Biochemistry, University of Cambridge, 1972, p. 493-497.
10	US application 09/015,454 filed January 29, 1998, entitled "System for Improving Yield of Sexed Embryos in Mammals".
	US application 09/001,394, filed December 31, 1997, entitled "Sheath Fluids and Collection Systems for Sex-Specific Cytometer Sorting of Sperm".
	US Application, 09/454,488, entitled "Improved Flow Cytometer Nozzle and Flow Cytometer Sample Handling Methods", filed December 3, 1999.
15	US Application, 09/448,643, entitled "Multiple Sexed Embryo Production System for Mammals", filed November
	US Application, 09/511,959 entitled "Methods For Improving Sheath Fluids and Collection Systems For Sex-Specific Cytometer Sorting of Sperm", filed February 23, 2001.
	US Application, 60/094,720, entitled "System for Low Dose Insemination of Equines", filed July 30, 1998.
20	US Application, 60/224,050., entitled "Integrated System for Herd Management With Terminal-Cross Program Using Sexed Semen", filed August 9, 2000.
	US Application, 60/113,143, entitled "Equine Insemination System", filed December 18, 1998.
	US Application, 60/203,089, entitled "Detector System for Resolving Small Differences in Photo-generated Signal", filed May 9, 2000.
	US Application, 60/238,294, entitled "Hysteroscopic Insemination of Mares" filed October 5, 2000.
25	US Application, entitled "System For Separating Frozen-Thawed Sperm Cells Into X-Chromosome And Y-Chromosome Bearing Populations", filed November 28, 2000.
	US Application, entitled "A System for In-vitro Fertilization with Spermatozoa Separated into X-chromosome and Y-chromosome Bearing Populations", filed November 28, 2000.
30	Van Munster E.B., et al, "Measurement-based evaluation of optical pathlength distributions reconstructed from simulated differential interference contrast images", Journal of Microscopy 192, Pt. 2, p. 170-176 (1998)
	Van Munster, E.B., et al, "Difference in Sperm Head Volume as a Theoretical Basis for Sorting X & Y-Bearing Spermatozoa: Potentials and Limitations", Theriogenology 52, pp. 1281-1293, (1999)
	Van Munster, E.B., et al, "Reconstruction of optical pathlength distributions form images obtained by a wide field differential interference contrast microscope", Journal of Microscopy 188, Pt. 2, p. 149-157 (1997)
35	Van Munster, E.B., "Geslachtsbepaling met interferometrie", Derde prijs NtvN-prijsvraag voor pas-gepromoveerden 65/4, p. 95-98 (1999)
	Van Munster E.B., et al, "Difference in Volume of X- and Y-chromosome Bearing Bovine Sperm Heads Matches Difference in DNA Content" Cytometry 35 p.125-128 (1999)

	Vazquez, J., et al., "A.I. in Swine; New Strategy for Deep Insemination with Low Number of Spermatozoa Using a Non-surgical Methodology", 14 <sup>th</sup> International Congress on Animal Reproduction, Vol. 2, Stockholom, July, 2000, p. 289.
5	Vazquez, J., et al., "Development of a Non-surgical Deep Intra Uterine Insemination Technique", IV International Conference on Boar Semen Preservation, Maryland, August, 1999, p 35 and photo of display board.
	Vazquez, J., et al., "Successful Low-Dose Insemination by a Fiberoptic Endoscope Technique in the Sow ", Proceedings Annual Conference of the International Embryo Transfer Society, Netherlands, Theriogenology, Vol. 53, January, 2000, pp. 201.
10	Vazquez, J., et al., "Hyposematic Swelling Test as Predictor of the Membrane Integrity in Boar Spermatozoa", Boar Semen Preservation IV, IVth International Conference on Boar Semen Preservation, Maryland, pp. 263.
	Vazquez, J. et al., "Nonsurgical Uterotubal Insemination in the Mare", Proceedings of the 44th Annual Convention of the American Association of Equine Practitioners, Baltimore, Maryland, December 6-9, 1998, Vol. 44, pp 68-69
	Vidament, M., Dupere, A.M., Julienne, P., Evain, A., Noue, P. and Palmer, E. 1997. Equine frozen semen freezeability and fertility field results. Theriogenology. 48:907.
15	Voss, J.L. et al., "Reproductive management of the broodmare". C.S.U. Exp. Sta. Anim. Reprod. Lab. Gen. Series. Bull. 961. (1976)
	Voss, J.L., et al., "Effect of number and frequency of inseminations on fertility in mares", J. Reprod. Fertil. Suppl. 32:53-57. (1982)
20	Voss, J.L., "Effect of human chorionic gonadotropin on duration of estrous cycle and fertility of normally cycling, non-lactating mares". J.A.V.M.A. 165:704-706. (1974)
	Welch, G., et al., "Flow Cytometric Sperm Sorting and PCR to Confirm Separation of X- and Y- Chromosome Bearing Bovine Sperm", Animal Biotechnology, 6 (2), 131-139, 1995, pp 131 - 139.
	Welch G.R., et al., "Fluidic and optical modifications to a FACS IV for flow sorting of X- and Y- chromosome bearing sperm based on DNA", Cytometry 17 (suppl. 7): 74. (1994)
25	Wilson, C.G., et al., "Effects of repeated hCG injections on reproductive efficiency in mares", Eq. Vet. Sci. 4:301-308. (1990)
	Wilson, M.S., "Non-surgical intrauterine artificial insemination in bitches using frozen semen", J.Reprod. Fert Suppl. 47:307-311. (1993)
30	Windsor, D.P., et al, "Sex Predetermination by Separation of X and Y Chromosome-bearing Sperm: A Review", Reproduction of Fertilization and Developement 5, pp. 155-171, (1993)
	Woods, J. and Ginther, O.J., "Recent studies related to the collection of multiple embryos in mares". Theriogenology. 19:101 - 108. (1983)
	Woods, J., et al., "Effects of time of insemination relative to ovulation on pregnancy rate and embryonic-loss rate in mares". Eq. Vet. J. 22(6):410-415. (1990)
35	XP-002103478, File Biosis, (1988), one page

In addition, as to each term used it should be understood that unless its utilization in this application is inconsistent with such interpretation, common dictionary definitions

should be understood as incorporated for each term and all definitions, alternative terms, and synonyms such as contained in the Random House Webster's Unabridged Dictionary, second edition are hereby incorporated by reference. However, as to each of the above, to the extent that such information or statements incorporated by reference might be

5 considered inconsistent with the patenting of this/these invention(s) such statements are expressly not to be considered as made by the applicant(s).

In addition, unless the context requires otherwise, it should be understood that the term "comprise" or variations such as "comprises" or "comprising", are intended to imply the inclusion of a stated element or step or group of elements or steps but not the

10 exclusion of any other element or step or group of elements or steps. Such terms should be interpreted in their most expansive form so as to afford the applicant the broadest coverage legally permissible in countries such as Australia and the like.

Thus, the applicant(s) should be understood to have support to claim at least: I) each of the liquid to gas conversion devices described herein, ii) the related methods

15 disclosed and described, iii) similar, equivalent, and even implicit variations of each of these devices and methods, iv) those alternative designs which accomplish each of the functions shown as are disclosed and described, v) those alternative designs and methods which accomplish each of the functions shown as are implicit to accomplish that which is disclosed and described, vi) each feature, component, and step shown as separate and

20 independent inventions, vii) the applications enhanced by the various systems or components disclosed, viii) the resulting products produced by such systems or components, ix) methods and apparatuses substantially as described hereinbefore and with reference to any of the accompanying examples, and the x) the various combinations and permutations of each of the elements disclosed

25 In addition, unless the context requires otherwise, it should be understood that the term "comprise" or variations such as "comprises" or "comprising", are intended to imply the inclusion of a stated element or step or group of elements or steps but not the exclusion of any other element or step or group of elements or steps. Such terms should

be interpreted in their most expansive form so as to afford the applicant the broadest coverage legally permissible in countries such as Australia and the like.

The claims set forth in this specification by are hereby incorporated by reference as part of this description of the invention, and the applicant expressly reserves the right

5 to use all of or a portion of such incorporated content of such claims as additional description to support any of or all of the claims or any element or component thereof, and the applicant further expressly reserves the right to move any portion of or all of the incorporated content of such claims or any element or component thereof from the description into the claims or vice-versa as necessary to define the matter for which

10 protection is sought by this application or by any subsequent continuation, division, or continuation-in-part application thereof, or to obtain any benefit of, reduction in fees pursuant to, or to comply with the patent laws, rules, or regulations of any country or treaty, and such content incorporated by reference shall survive during the entire pendency of this application including any subsequent continuation, division, or

15 continuation-in-part application thereof or any reissue or extension thereon.

## VI. CLAIMS

We claim:

1. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells, comprising the steps of:
  - 5 a. collecting sperm cells from a male of a species of mammal;
  - b. determining a sex differentiation characteristic of said sperm cells;
  - c. differentiating between said sperm cells based upon said sex differentiation characteristic;
  - d. separating differentiated sperm cells into a X-chromosome bearing population and a Y-chromosome bearing population; and
  - 10 e. producing separate X-chromosome bearing and Y-chromosome bearing populations of sperm cells each having a purity of greater than 90%.
2. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 1, wherein said step of differentiating  
15 between said sperm cells based upon said sex differentiation characteristic comprises assessing an amount of DNA within a nucleus of said sperm cells.
3. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 1, wherein said step of differentiating between said sperm cells based upon said sex differentiation characteristic  
20 comprises assessing a volume of a capsule containing said amount of DNA.
4. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 3, wherein said step of assessing said volume of said capsule containing said amount of DNA comprises determining a volume of a nucleus within said sperm cells.
- 25 5. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 3, wherein said step of assessing said

volume of a capsule containing said DNA comprises determining the volume of a sperm cell head.

6. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 5, wherein said step of assessing said  
5 volume of said capsule containing said amount of DNA further comprises the steps of :
  - a. generating a beam of electromagnetic radiation having initial wave form characteristics;
  - b. traversing said volume of said capsule containing said amount of DNA  
10 with said beam of electromagnetic radiation;
  - c. altering said initial wave form characteristics by traversing said volume of said capsule; and
  - d. analyzing altered wave form characteristics.
7. A method of isolating X-chromosome bearing sperm cells and Y-chromosome  
15 bearing sperm cells as described in claim 6, wherein said step of altering said initial wave form characteristics by traversing said volume of said capsule comprises shifting phase of said initial wave form of said beam of electromagnetic radiation.
8. A method of isolating X-chromosome bearing sperm cells and Y-chromosome  
20 bearing sperm cells as described in claim 6, further comprising the step of superimposing said beam of electromagnetic radiation having shifted phase on said beam of electromagnetic radiation having said initial wave form characteristics.
9. A method of isolating X-chromosome bearing sperm cells and Y-chromosome  
25 bearing sperm cells as described in claim 8, further comprising the step of comparing electromagnetic radiation intensity of superimposed beams of electromagnetic radiation with intensity of said beam of electromagnetic radiation

having initial wave form characteristics.

10. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 9, further comprising the step of determining said volume of said capsule containing said amount of DNA based  
5 upon a difference in said light intensity between said superimposed beams of electromagnetic radiation and said beam of electromagnetic radiation having initial waveform characteristics.
11. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 2, further comprising the steps of:
- 10 a. introducing said sperm cells into a fluid stream;  
b. analyzing said sperm cells entrained in said fluid stream;  
c. forming droplets a plurality having one of said sperm cells entrained;  
d. charging each of said droplets differentially based upon said sex differentiation characteristic of said sperm cells entrained in said droplets;  
15 e. deflecting each of said droplets; and  
f. differentially collecting each of said droplets based upon said sex differentiation characteristic of said sperm cells entrained in said droplets.
12. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 11, further comprising the step of  
20 maintaining said amount of DNA within said nucleus of said sperm cell unstained.
13. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 11, further comprising the step of staining said amount of DNA within said nucleus of said sperm cell.
14. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 13, further comprising the step of  
25 irradiating stained DNA within said nucleus of said sperm cell.



15. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 14, further comprising the step of detecting fluorescent light emitted from irradiated stained DNA within the nucleus of said sperm cell.
- 5 16. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 15, wherein said step of detecting fluorescent light from irradiated stained DNA within said nucleus of said sperm cell comprises generating a signal with a photomultiplier tube.
- 10 17. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 16, further comprising the step of operating said photomultiplier tube outside a typical operation voltage range.
- 15 18. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 14, wherein said step of irradiating stained DNA within the nucleus of said sperm cell further comprises generating an irradiation beam pattern having a reduced height.
- 20 19. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 18, wherein said reduced height comprises a height about equal to the length of said sperm cells along the longitudinal axis to about three times the length of said sperm cells along the longitudinal axis.
- 20 20. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 19, wherein said sperm cell has a longitudinal axis of about nine micrometers in length, and said height of said irradiation beam pattern is about 20 micrometers.
- 25 21. A method of isolating X-chromosome bearing sperm cells and Y-chromosome

bearing sperm cells as described in claim 18, wherein said step of forming droplets comprises forming droplets having sufficient size to encapsulate said sperm cell.

22. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 21, further comprising the step of ejecting said fluid stream from a nozzle having an orifice, wherein said orifice has a diameter of 70 micrometers.
23. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 22, further comprising the steps of:
- a. orienting said sperm cells with respect to a detector;
  - b. detecting light having characteristics differentially responsive to sperm cell orientation to said detector;
  - c. converting said light having characteristics deferentially responsive to said sperm cell orientation into at least one signal containing sperm cell orientation information; and
  - d. determining orientation of said sperm cell with respect to said detector.
24. a method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 23, further comprising the step of differentially collecting said droplets based upon determined orientation of said sperm cell with respect to said detector.
25. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claims 1, 6, 12, 13, 17, 18, 21, or 23, wherein said purity of said X-chromosome bearing and said Y-chromosome bearing populations of sperm cells is selected from the group consisting of between 90% to about 100%, between about 91% to about 100%, between about 92% to about 100%, between about 93% to about 100%, between about 94% to about 100%, between about 95% to about 100%, between about 96% to about 100%, between

about 97% to about 100%, between about 98% to about 100%, between about 99% to about 100%.

26. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claims 1, 6, 12, 13, 17, 18, 21, or 23, further comprising the step of establishing a separable event rate from the group consisting of at least 5000 separable events per second, at least 6000 separable events per second, at least 7000 separable events per second, at least 8000 separable events per second, at least 9000 separable events per second, at least 10,000 separable events per second, at least 11,000 separable events per second, at least 12,000 separable events per second, at least 13,000 separable events per second, at least 14,000 separable events per second, at least 15,000 separable events per second, at least 16,000 separable events per second, at least 17,000 separable events per second, at least 18,000 separable events per second, at least 19,000 separable events per second, at least 20,000 separable events per second, at least 21,000 separable events per second.
27. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claims 1, 6, 12, 13, 18, 21, or 23, wherein said step of separating said sperm cells comprises a separation rate selected from the group consisting of at least 500 separations per second, at least 1,000 separations per second, at least 2,000 separations per second, at least 3,000 separations per second, at least 4,000 separations per second, at least 5,000 separations per second, at least 6,000 separations per second, at least 7,000 separations per second, at least 8,000 separations per second, at least 9,000 separations per second, at least 10,000 separations per second.
28. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claims 1, 6, 12, 13, 18, 21, or 23, wherein said step of forming droplets each having one of said sperm cells entrained comprises a droplet formation rate selected from the group consisting of at least 10,000

- droplets per second, at least 20,000 droplets per second, at least 30,000 droplets per second, at least 40,000 droplets per second, at least 50,000 droplets per second, at least 60,000 droplets per second, at least 70,000 droplets per second, at least 80,000 droplets per second, at least 90,000 droplets per second, at least 100,000 droplets per second.
- 5
29. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 1, wherein said species of mammal comprises a bovine mammal.
30. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 1, wherein said species of mammal comprises an equine mammal.
- 10
31. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 1, wherein said species of mammal comprises an ovine mammal.
- 15 32. An apparatus to isolate X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells, comprising:
- a. sperm cells having a sperm heads, wherein said sperm heads have a difference in volume between X-chromosome bearing sperms cells and Y-chromosome bearing sperm cells;
- 20 b. a beam of electromagnetic radiation having initial waveform characteristics differentially responsive to said difference in volume between X-chromosome bearing sperms cells and Y-chromosome bearing sperm cells;
- c. a detector responsive to altered waveform characteristics of said beam of electromagnetic radiation;
- 25 d. an analyzer coupled to said detector, wherein said analyzer differentiates between said difference in volume between said X-chromosome bearing

sperms cells and said Y-chromosome bearing sperm cells based upon said altered waveform characteristics.

33. An apparatus to isolate X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 32, further comprising a fluid stream  
5 into which said sperm cells are introduced.
34. An apparatus to isolate X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 33, further comprising droplets breaking off from said fluid stream a plurality having one of said sperm cells entrained.
- 10 35. An apparatus to isolate X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 34, wherein said droplets breaking off from said fluid stream have sufficient size to encapsulate said one of said sperm cells.
- 15 36. An apparatus to isolate X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 35, further comprising a nozzle having an orifice of about 100 micrometers in diameter.
- 20 37. An apparatus to isolate X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 36, further comprising a droplet charger coupled to said analyzer, wherein said droplets receive a charge differentially based upon said difference in volume between X-chromosome bearing sperms cells and Y-chromosome bearing sperm cells.
- 25 38. An apparatus to isolate X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 37, further comprising a droplet separator, wherein said droplet separator separates said droplet based upon charge of said droplet.

39. An apparatus to isolate X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 38, further comprising at least one collection container in which droplets containing said X-chromosome bearing sperm cells are collected as an X-chromosome bearing population.
- 5 40. An apparatus to isolate X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 38, further comprising at least one collection container in which droplets containing Y-chromosome bearing sperm cells are collected as a Y-chromosome bearing population.
41. An apparatus to isolate X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claims 38 or 39, wherein said X-chromosome bearing population and said Y-chromosome bearing population of said sperm cells have a purity selected from the group consisting of between 90% to about 100%, between about 91% to about 100%, between about 92% to about 100%, between about 93% to about 100%, between about 94% to about 100%, between about 95% to about 100%, between about 96% to about 100%, between about 97% to about 100%, between about 98% to about 100%, between about 99% to about 100%.
- 10 42. An apparatus to isolate X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 41, further comprising a separable event rate wherein said separable event rate is selected from the group consisting of at least 5000 separable events per second, at least 6000 separable events per second, at least 7000 separable events per second, at least 8000 separable events per second, at least 9000 separable events per second, at least 10,000 separable events per second, at least 11,000 separable events per second, at least 12,000 separable events per second, at least 13,000 separable events per second, at least 14,000 separable events per second, at least 15,000 separable events per second, at least 16,000 separable events per second, at least 17,000 separable events per second, at least 18,000 separable events per second, at least 19,000 separable events per
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second, at least 20,000 separable events per second, at least 21,000 separable events per second.

43. An apparatus to isolate X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 42, further comprising a separation rate  
5 selected from the group consisting of at least 500 separations per second, at least 1,000 separations per second, at least 2,000 separations per second, at least 3,000 separations per second, at least 4,000 separations per second, at least 5,000 separations per second, at least 6,000 separations per second, at least 7,000 separations per second, at least 8,000 separations per second, at least 9,000  
10 separations per second, at least 10,000 separations per second, 11,000 separations per second.
44. An apparatus to isolate X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 43, further comprising a droplet formation rate selected from the group consisting of at least 10,000 droplets per  
15 second, at least 20,000 droplets per second, at least 30,000 droplets per second, at least 40,000 droplets per second, at least 50,000 droplets per second, at least 60,000 droplets per second, at least 70,000 droplets per second, at least 80,000 droplets per second, at least 90,000 droplets per second, at least 100,000 droplets per second.
- 20 45. An apparatus to isolate X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 44, wherein said detector comprises at least one photomultiplier tube, and wherein said at least one photomultiplier tube converts said electromagnetic radiation into at least one signal; and wherein said photomultiplier tube has a typical operation voltage range, and wherein said  
25 photomultiplier tube is operated outside said typical operation voltage range.
46. An apparatus to isolate X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 45, wherein said typical operation

voltage range of said photomultiplier tube is from about 400 volts to about 999 volts.

47. An apparatus to isolate X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 46, wherein said photomultiplier tube is operated in a range from about 0 volts to about 300 volts.
48. An apparatus to isolate X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 41, wherein said species of mammal comprises a bovine mammal.
49. An apparatus to isolate X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 41, wherein said species of mammal comprises an equine mammal.
50. An apparatus to isolate X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 41, wherein said species of mammal comprises an ovine mammal.
51. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells, comprising the steps of:
- a. collecting sperm cells from a male of a species of mammal;
  - a. assessing a volume of a capsule containing the nuclear DNA of said sperm cells;
  - c. differentiating between said X-chromosome bearing sperm cells and said Y-chromosome bearing sperm cells based upon said volume of said capsule containing said nuclear DNA of said sperm cells;
52. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 51, wherein said step of assessing the volume of a capsule containing said DNA comprises determining a phase shift in



said electromagnetic radiation.

53. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 52, wherein said step of assessing the volume of a capsule containing said DNA comprises:
- 5 a. generating a beam electromagnetic radiation having initial waveform characteristics;
- b. traversing said volume of said capsule containing said DNA with said beam of electromagnetic radiation having initial waveform characteristics;
- c. altering said initial waveform characteristics by traversing said volume of said capsule; and
- 10 d. analyzing altered wave form characteristics of said beam of electromagnetic radiation.
54. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 53, wherein said step of altering said initial wave form characteristics by traversing said volume of said capsule
- 15 comprises shifting phase of said initial waveform characteristics of said beam of electromagnetic radiation.
55. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 54, further comprising the step of superimposing said initial waveform characteristics and phase shifted waveform characteristics.
- 20
56. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 55, further comprising the step of comparing an intensity of said initial waveform characteristics with superimposed waveform characteristics.
- 25
57. A method of isolating X-chromosome bearing sperm cells and Y-chromosome

bearing sperm cells as described in claim 56, further comprising the step of determining said volume of said capsule containing said DNA based upon the difference in said intensity between said superimposed waveform characteristics and said initial waveform characteristics.

- 5 58. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 57, further comprising the steps of:
- a. introducing said sperm cells into a fluid stream;
  - b. forming droplets a plurality having one of said sperm cells entrained;
  - c. charging each of said droplets differentially based upon the assessed
  - 10 volume of said capsule containing the nuclear DNA of said sperm cells;
  - d. deflecting each of said droplets;
  - e. differentially collecting each of said droplets based upon said volume of said capsule containing said DNA; and
  - f. generating X-chromosome bearing and Y-chromosome bearing
  - 15 populations of said sperm cells.
59. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 58, further comprising the step of maintaining said amount of DNA within the nucleus of said sperm cells unstained.
60. a method of isolating X-chromosome bearing sperm cells and Y-chromosome
- 20 bearing sperm cells as described in claim 59, further comprising the step of forming droplets having sufficient size to encapsulate said sperm cells.
61. a method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 60, further comprises the step of ejecting said fluid stream from a nozzle having an orifice, wherein said orifice has a
- 25 diameter of 100 micrometers.
62. a method of isolating X-chromosome bearing sperm cells and Y-chromosome

bearing sperm cells as described in claim 58, further comprises the steps of:

- a. detecting a signal having sperm cell orientation characteristics;
- b. comparing said signal having sperm cell orientation characteristics to a signal having oriented sperm cell characteristics;
- 5 c. determining said sperm cell has unoriented sperm cell characteristics; and
- d. differentially collecting said sperm cell having unoriented sperm cell characteristics.

63. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claims 58 or 62, wherein purity of said X-chromosome bearing and said Y-chromosome bearing populations of said sperm  
10 cells is selected from the group consisting of between 90% to about 100%, between about 91% to about 100%, between about 92% to about 100%, between about 93% to about 100%, between about 94% to about 100%, between about 95% to about 100%, between about 96% to about 100%, between about 97% to about 100%, between about 98% to about 100%, between about 99% to about 100%.

64. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 63, further comprising the step of establishing a separable event rate wherein said separable event rate is selected  
20 from the group consisting of at least 5000 separable events per second, at least 6000 separable events per second, at least 7000 separable events per second, at least 8000 separable events per second, at least 9000 separable events per second, at least 10,000 separable events per second, at least 11,000 separable events per second, at least 12,000 separable events per second, at least 13,000 separable  
25 events per second, at least 14,000 separable events per second, at least 15,000 separable events per second, at least 16,000 separable events per second, at least 17,000 separable events per second, at least 18,000 separable events per second, at least 19,000 separable events per second, at least 20,000 separable events per second, at least 21,000 separable events per second.

65. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 64, wherein said step of separating said sperm cells comprises a separation rate selected from the group consisting of at least 500 separations per second, at least 1,000 separations per second, at least 2,000 separations per second, at least 3,000 separations per second, at least 4,000 separations per second, at least 5,000 separations per second, at least 6,000 separations per second, at least 7,000 separations per second, at least 8,000 separations per second, at least 9,000 separations per second, at least 10,000 separations per second.
- 10 66. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 65, wherein said step of forming droplets each having one of said sperm cells entrained comprises a droplet formation rate selected from the group consisting of at least 10,000 droplets per second, at least 20,000 droplets per second, at least 30,000 droplets per second, at least 40,000 droplets per second, at least 50,000 droplets per second, at least 60,000 droplets per second, at least 70,000 droplets per second, at least 80,000 droplets per second, at least 90,000 droplets per second, at least 100,000 droplets per second.
- 15 67. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 51, wherein said species of mammal comprises a bovine mammal.
- 20 68. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 51, wherein said species of mammal comprises an equine mammal.
- 25 69. A method of isolating X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells as described in claim 51, wherein said species of mammal comprises an ovine mammal.

70. A method differentiating particles, comprising the steps of:
- a. collecting asymmetric particles;
  - b. establishing said fluid stream with a flow cytometer;
  - c. introducing said asymmetric particles into said fluid stream;
  - 5 d. orienting said asymmetric particles with respect to a detector;
  - e. detecting light having characteristics differentially responsive to asymmetric particle orientation to said detector;
  - g. converting said light having characteristics differentially responsive to asymmetric particle orientation into at least one signal containing asymmetric particle orientation information;
  - 10 h. analyzing asymmetric particle orientation information; and
  - i. determining orientation of said asymmetric particles with respect to said detector.
71. A method differentiating particles as described in claim 70, wherein said fluid  
15 stream comprises a sheath fluid
72. A method differentiating particles as described in claim 70, wherein said detector comprises a photomultiplier tube.
73. A method differentiating particles as described in claim 72, wherein said light comprises florescent light.
- 20 74. A method differentiating particles as described in claim 73, wherein said florescent light emits from a light emission material bound to said asymmetric particles.
75. A method differentiating particles as described in claim 70, wherein said step of analyzing asymmetric particle orientation information comprises the steps of:
- 25 a. plotting electronic signal over a period of time;
  - b. integrating the areas of plots corresponding to said electronic signal; and

- c. comparing integrations of said plots to integrations of oriented asymmetric particles.
76. A method differentiating particles as described in claim 70, further comprising differentially collecting said asymmetric particles based upon determined  
5 orientation of said asymmetric particles with respect to said detector.
77. A method differentiating particles as described in claim 70, wherein said asymmetric particles are sperm cells from a male of a species of mammal.
78. A method differentiating particles as described in claim 70, further comprising the steps of:
- 10 a. assessing the volume of a capsule containing the nuclear DNA of said sperm cells; and
- b. differentiating between said X-chromosome bearing sperm cells and said Y-chromosome bearing sperm cells based upon the volume of said capsule containing the nuclear DNA of said sperm cell.
- 15 79. A method differentiating particles as described in claim 78, wherein said capsule containing said DNA comprises a sperm head.
80. A method differentiating particles as described in claim 78, wherein said step of assessing the volume of a capsule containing said DNA comprises:
- 20 a. generating a beam of electromagnetic radiation having initial waveform characteristics;
- b. traversing said volume of said capsule containing said DNA with said beam of electromagnetic radiation having initial waveform characteristics;
- c. altering said initial wave form characteristics of said beam of electromagnetic radiation by traversing said volume of said capsule; and
- 25 d. analyzing altered wave form characteristics.

81. A method differentiating particles as described in claim 80, wherein said step of altering said wave form characteristics by traversing said volume of said capsule comprises shifting phase of said initial waveform characteristics.
82. A method differentiating particles as described in claim 81, further comprising the  
5 step of superimposing said initial waveform characteristics and said phase shifted waveform characteristics.
83. A method differentiating particles as described in claim 82, further comprising the step of comparing a intensity of said initial waveform characteristics and superimposed waveform characteristics.
- 10 84. A method differentiating particles as described in claim 83, further comprising the step of determining said volume of said capsule containing said DNA based upon the difference in said intensity.
85. A method differentiating particles as described in claim 80, further comprising the steps of:
- 15 a. forming droplets a plurality having one of said sperm cells entrained;  
b. charging each of said droplets differentially based upon the determined volume of said capsule containing the nuclear DNA of said sperm cells;  
c. deflecting each of said droplets; and  
d. collecting each of said droplets based upon charge of said droplet; and  
20 e. generating X-chromosome bearing and Y-chromosome bearing populations of said sperm cells.
86. A method differentiating particles as described in claim 77, further comprising the steps of:
- 25 a. forming droplets a plurality having one of said sperm cells entrained;  
b. charging each of said droplets differentially based upon an amount of nuclear DNA of said sperm cells;

- c. deflecting each of said droplets;
  - d. collecting each of said droplets based upon charge of said droplet; and
  - e. generating X-chromosome bearing and Y-chromosome bearing populations of said sperm cells.
- 5 87. A method differentiating particles as described in claims 85 or 86, wherein said step of forming droplets a plurality having one of said sperm cells entrained comprises forming droplets having sufficient size to encapsulate said sperm cell, wherein said sperm cell comprises an intact live sperm cell having a tail.
- 10 88. A method differentiating particles as described in claim 87, further comprises the step of ejecting said fluid stream from a nozzle having an orifice, wherein said orifice has a diameter of 100 micrometers.
- 15 89. A method differentiating particles as described in claim 88, wherein said X-chromosome bearing and said Y-chromosome bearing populations of sperm cells have a purity selected from the group consisting of between 90% to about 100%, between about 91% to about 100%, between about 92% to about 100%, between about 93% to about 100%, between about 94% to about 100%, between about 95% to about 100%, between about 96% to about 100%, between about 97% to about 100%, between about 98% to about 100%, between about 99% to about 100%.
- 20 90. A method differentiating particles as described in claim 88, further comprising the step of establishing a separable event rate wherein said separable event rate is selected from the group consisting of at least 5000 separable events per second, at least 6000 separable events per second, at least 7000 separable events per second, at least 8000 separable events per second, at least 9000 separable events per second, at least 10,000 separable events per second, at least 11,000 separable events per second, at least 12,000 separable events per second, at least 13,000 separable events per second, at least 14,000 separable events per second, at least
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15,000 separable events per second, at least 16, 000 separable events per second, at least 17,000 separable events per second, at least 18,000 separable events per second, at least 19,000 separable events per second, at least 20,000 separable events per second, at least 21,000 separable events per second.

- 5 91. A method differentiating particles as described in claim 88, wherein said step of separating said sperm cells comprises a separation rate selected from the group consisting of at least 500 separations per second, at least 1,000 separations per second, at least 2,000 separations per second, at least 3,000 separations per second, at least 4,000 separations per second, at least 5,000 separations per second, at least 6,000 separations per second, at least 7,000 separations per second, at least 8,000 separations per second, at least 9,000 separations per second, at least 10,000 separations per second.
- 10
92. A method differentiating particles as described in claim 88, wherein said step of forming droplets each having one of said sperm cells entrained comprises a droplet formation rate selected from the group consisting of at least 10,000 droplets per second, at least 20,000 droplets per second, at least 30,000 droplets per second, at least 40,000 droplets per second, at least 50,000 droplets per second, at least 60,000 droplets per second, at least 70,000 droplets per second, at least 80,000 droplets per second, at least 90,000 droplets per second, at least 100,000 droplets per second.
- 15
- 20
93. A method differentiating particles as described in claim 77, wherein said species of mammal comprises a bovine mammal.
94. A method differentiating particles as described in claim 77, wherein said species of mammal comprises an equine mammal.
- 25 95. A method differentiating particles as described in claim 77, wherein said species of mammal comprises an ovine mammal.

96. A particle differentiation apparatus, comprising:
- a. at least one asymmetric particle having orientation characteristics within a fluid stream;
  - b. a irradiation source generating an irradiation beam responsive to said asymmetric particle;
  - c. optics to focus said irradiation beam;
  - d. a light emission material coupled to said asymmetric particle, wherein said light emission material emits light in response to said irradiation beam;
  - e. a detector deferentially responsive to said light emitted from said light emission material based upon said particle orientation characteristics; and
  - f. an analyzer coupled to said detector, wherein said analyzer differentiates said asymmetric particles based upon orientation within said fluid stream.
97. A particle differentiation apparatus as described in claim 96, wherein said at least one asymmetrical particle comprise at least one sperm cell.
98. A particle differentiation apparatus as described in claim 96, wherein said optics focus a beam pattern having a height of about equal to the length of said asymmetrical particle along the longitudinal axis to about three times the length of said asymmetrical particle along the longitudinal axis;
99. A particle differentiation apparatus as described in claim 98, wherein said at least one sperm cell has a head having a length along the longitudinal axis of about nine micrometers, and wherein said irradiation pattern has a height of about 20 micrometers.
100. A particle differentiation apparatus as described in claim 96, wherein said at least one asymmetric particle having orientation characteristics further comprises at least one particle differentiation characteristic.
101. A particle differentiation apparatus as described in claim 96, wherein said light

emission material bound to said at least one asymmetric particle comprises a stain bound to nuclear DNA of sperm cells of a male of a species of mammal.

102. A particle differentiation apparatus as described in claim 101, wherein said particle differentiation characteristic comprises a difference in amount of said stain bound to said nuclear DNA of X-chromosome bearing sperm cells and said nuclear DNA of Y-chromosome bearing sperm cells.
103. A particle differentiation apparatus as described in claim 96, wherein said detector comprises at least one photomultiplier tube, and wherein said photomultiplier tube has a typical operation voltage range, and wherein said photomultiplier tube is operated outside said typical operation voltage range.
104. A particle differentiation apparatus as described in claim 103, wherein said typical operation voltage range of said photomultiplier tube is from about 400 volts to about 999 volts.
105. A particle differentiation apparatus as described in claim 104, wherein said photomultiplier tube is operated in a range from about 0 volts to about 300 volts.
106. A particle differentiation apparatus as described in claim 96, further comprising droplets breaking off from said fluid stream a plurality having at least one of said asymmetric particles entrained.
107. A particle differentiation apparatus as described in claim 106, wherein said droplets breaking off from said fluid stream have sufficient size to encapsulate one of said sperm cells, wherein said sperm cells comprise intact live sperm cells having a tail.
108. A particle differentiation apparatus as described in claim 107, further comprising a nozzle having an orifice of about 100 micrometers in diameter.

109. A particle differentiation apparatus as described in claim 106, further comprising a droplet charger coupled to said analyzer, wherein said droplets receive a charge differentially based upon said difference in amount of said stain bound the nuclear DNA of X-chromosome sperm cells and the nuclear DNA of Y-chromosome bearing sperm cells.
110. A particle differentiation apparatus as described in claim 109, further comprising a droplet charger coupled to said analyzer, wherein said droplets receive a charge differentially based upon said difference in volume of said particles, and wherein said difference in volume of said particles comprises a difference between X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells.
111. A particle differentiation apparatus as described in claim 110, further comprising a droplet separator, wherein said droplet separator separates said droplet based upon charge of said droplet.
112. A particle differentiation apparatus as described in claim 111, further comprising at least one collection container in which droplets containing said X-chromosome bearing sperm cells are collected as an X-chromosome bearing population.
113. A particle differentiation apparatus as described in claim 111, further comprising at least one collection container in which droplets containing Y-chromosome bearing sperm cells are collected as a Y-chromosome bearing population.
114. A particle differentiation apparatus as described in claim 112 or 113, wherein said X-chromosome bearing population and said Y-chromosome bearing population of said sperm cells are selected from the group consisting of between 90% to about 100%, between about 91% to about 100%, between about 92% to about 100%, between about 93% to about 100%, between about 94% to about 100%, between about 95% to about 100%, between about 96% to about 100%, between about 97% to about 100%, between about 98% to about 100%, between about 99% to

about 100%.

115. A particle differentiation apparatus as described in claim 114, further comprising the step of establishing a separable event rate wherein said separable event rate is selected from the group consisting of at least 5000 separable events per second, at least 6000 separable events per second, at least 7000 separable events per second, at least 8000 separable events per second, at least 9000 separable events per second, at least 10,000 separable events per second, at least 11,000 separable events per second, at least 12,000 separable events per second, at least 13,000 separable events per second, at least 14,000 separable events per second, at least 15,000 separable events per second, at least 16,000 separable events per second, at least 17,000 separable events per second, at least 18,000 separable events per second, at least 19,000 separable events per second, at least 20,000 separable events per second, at least 21,000 separable events per second.
116. A particle differentiation apparatus as described in claim 115, wherein said step of separating said sperm cells comprises a separation rate selected from the group consisting of at least 500 separations per second, at least 1,000 separations per second, at least 2,000 separations per second, at least 3,000 separations per second, at least 4,000 separations per second, at least 5,000 separations per second, at least 6,000 separations per second, at least 7,000 separations per second, at least 8,000 separations per second, at least 9,000 separations per second, at least 10,000 separations per second, at least 11,000 separations per second.
117. A particle differentiation apparatus as described in claim 116, wherein said step of forming droplets each having one of said sperm cells entrained comprises a droplet formation rate selected from the group consisting of at least 10,000 droplets per second, at least 20,000 droplets per second, at least 30,000 droplets per second, at least 40,000 droplets per second, at least 50,000 droplets per second, at least 60,000 droplets per second, at least 70,000 droplets per second, at least 80,000 droplets per second, at least 90,000 droplets per second, at least

100,000 droplets per second.

118. A particle differentiation apparatus as described in claim 102, wherein said species of mammal comprises a bovine mammal.
119. A particle differentiation apparatus as described in claim 102, wherein said species  
5 of mammal comprises an equine mammal.
120. A particle differentiation apparatus as described in claim 102, wherein said species of mammal comprises an ovine mammal.
121. A particle differentiation apparatus, comprising:
- a. a fluid stream;
  - 10 b. intact live sperm cells introduced into said fluid stream;
  - c. a nozzle having a orifice through which said fluid stream exits;
  - d. an oscillator responsive to said fluid stream; and
  - e. droplets breaking off from said fluid stream, wherein a plurality of said  
droplets entrain said intact live sperm cells, and wherein said droplets  
15 have sufficient size to encapsulate one of said live sperm cells.
122. A particle differentiation apparatus as described in claim 121, wherein said orifice has a diameter of 100 micrometers.
123. A particle differentiation apparatus as described in claim 122, wherein said  
droplets breaking off from said fluid stream break off at a rate selected from the  
20 group consisting of at least 10,000 droplets per second, at least 20,000 droplets per second, at least 30,000 droplets per second, at least 40,000 droplets per second, at least 50,000 droplets per second, at least 60,000 droplets per second, at least 70,000 droplets per second, at least 80,000 droplets per second, at least 90,000 droplets per second, at least 100,000 droplets per second.

124. A particle differentiation apparatus as described in claim 123, further comprising a light emission source generating light.
125. A particle differentiation apparatus as described in claim 124, further comprising a detector responsive to said light.
- 5 126. A particle differentiation apparatus as described in claim 125, wherein said intact live sperm cells have at least one sex differentiation characteristic.
127. A particle differentiation apparatus as described in claim 126, wherein said at least one sex differentiation characteristic comprises a volume difference of sperm cell heads of said intact live sperm cells.
- 10 128. A particle differentiation apparatus as described in claim 127, wherein said light emission source emits a beam of electromagnetic radiation, and wherein said beam of electromagnetic radiation traverses said volume of said sperm cell heads, and wherein said beam of electromagnetic radiation has initial waveform characteristics differentially responsive to said difference in volume of said sperm
- 15 cell heads.
129. A particle differentiation apparatus as described in claim 128, further comprising an analyzer responsive to said detector, wherein said analyzer differentiates between said intact live sperm cells based upon said volume difference of said sperm cell heads.
- 20 130. A particle differentiation apparatus as described in claim 129, wherein said volume difference of said sperm cell heads in volume comprises a difference between X-chromosome bearing live intact sperm cells and Y-chromosome bearing live intact sperm cells.
131. A particle differentiation apparatus as described in claim 126, an irradiation source

generating an irradiation beam responsive to said intact live sperm cells.

132. A particle differentiation apparatus as described in claim 131, further comprising optics, wherein said optics focus an irradiation beam pattern having a height equal  
5 to about the length of said sperm heads along the longitudinal axis to about three times the length of said sperm head of said along the longitudinal axis.
133. A particle differentiation apparatus as described in claim 132, wherein said sperm heads have a length along the longitudinal axis of about nine micrometers, and  
10 wherein said irradiation pattern has a height of about 20 micrometers.
134. A particle differentiation apparatus as described in claim 131, wherein said at least one sex differentiation characteristic comprises a difference in amount of nuclear DNA of said intact live sperm cells, and wherein said light emission source  
15 comprises a light emission material bound to nuclear DNA, and wherein said light emission material emits light differentially based upon said difference in amount of said nuclear DNA, and wherein said detector generates at least one signal in response to said light.
135. A particle differentiation apparatus as described in claim 134, further comprising an analyzer responsive to said detector, wherein said analyzer differentiates  
20 between said at least one intact live sperm cell based upon said difference in amount of nuclear DNA.
136. A particle differentiation apparatus as described in claim 135, wherein said difference in said amount of said nuclear DNA comprises a difference between X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells.
- 25 137. A particle differentiation apparatus as described in claims 136, further comprising a droplet charger coupled to said analyzer, wherein said droplet charger generates a charge on said droplets differentially based upon said difference in amount of



said nuclear DNA.

138. A particle differentiation apparatus as described in claim 129, further comprising a droplet charger coupled to said analyzer, wherein said droplet charger generates a charge on said droplets differentially based upon said volume difference of said sperm cell heads, and wherein said volume difference of said sperm cell heads comprises a difference between X-chromosome bearing sperm cells and Y-chromosome bearing sperm cells.
139. A particle differentiation apparatus as described in claims 137 or 138, further comprising a droplet separator, wherein said droplet separator separates said droplets based upon charge of said droplets.
140. A particle differentiation apparatus as described in claim 139, further comprising at least one collection container in which droplets containing said X-chromosome bearing sperm cells are collected as an X-chromosome bearing population.
141. A particle differentiation apparatus as described in claim 139, further comprising at least one collection container in which droplets containing Y-chromosome bearing sperm cells are collected as a Y-chromosome bearing population.
142. A particle differentiation apparatus as described in claim 125, wherein said detector comprises at least one photomultiplier tube, and wherein said at least one photomultiplier tube converts light from said light emission source into at least one signal, and wherein said photomultiplier tube has a typical operation voltage range, and wherein said photomultiplier tube is operated outside said typical operation voltage range.
143. A particle differentiation apparatus as described in claim 142, wherein said typical operation voltage range of said photomultiplier tube is from about 400 volts to about 999 volts.

144. A particle differentiation apparatus as described in claim 143, wherein said photomultiplier tube is operated in a range from about 0 volts to about 300 volts.
- 145 A particle differentiation apparatus as described in claims 140 or 141, wherein said X-chromosome bearing population and said Y-chromosome bearing  
5 population of said sperm cells have a purity selected from the group consisting of between 90% to about 100%, between about 91% to about 100%, between about 92% to about 100%, between about 93% to about 100%, between about 94% to about 100%, between about 95% to about 100%, between about 96% to about 100%, between about 97% to about 100%, between about 98% to about 100%,  
10 between about 99% to about 100%.
146. A particle differentiation apparatus as described in claim 145, further comprising the step of establishing a separable event rate wherein said separable event rate is selected from the group consisting of at least 5000 separable events per second, at least 6000 separable events per second, at least 7000 separable events per second,  
15 at least 8000 separable events per second, at least 9000 separable events per second, at least 10,000 separable events per second, at least 11,000 separable events per second, at least 12,000 separable events per second, at least 13,000 separable events per second, at least 14,000 separable events per second, at least 15,000 separable events per second, at least 16, 000 separable events per second,  
20 at least 17,000 separable events per second, at least 18,000 separable events per second, at least 19,000 separable events per second, at least 20,000 separable events per second, at least 21,000 separable events per second.
147. A particle differentiation apparatus as described in claim 146, wherein said step of separating said sperm cells comprises a separation rate selected from the group  
25 consisting of at least 500 separations per second, at least 1,000 separations per second, at least 2,000 separations per second, at least 3,000 separations per second, at least 4,000 separations per second, at least 5,000 separations per second, at least 6,000 separations per second, at least 7,000 separations per

second, at least 8,000 separations per second, at least 9,000 separations per second, at least 10,000 separations per second, 11,000 separations per second.

148. A particle differentiation apparatus, comprising:
- a. particles having at least one particle differentiation characteristic;
  - 5 b. a light emission source differentially responsive to said particle differentiation characteristics;
  - c. at least one photomultiplier tube, wherein said at least one photomultiplier tube converts light from said light emission source into at least one signal, and wherein said photomultiplier tube has a typical operation voltage range, and wherein said photomultiplier tube is operated outside said  
10 typical operation voltage range; and
  - d. an analyzer responsive to said at least one signal which differentiates particles based upon said particle differentiation characteristics.
- 15 149. A particle differentiation apparatus as described in claim 148, wherein said typical operation voltage range of said photomultiplier tube is from about 400 volts to about 999 volts.
150. A particle differentiation apparatus as described in claim 149, wherein said photomultiplier tube is operated in a range from about 0 volts to about 300 volts.
- 20 151. A particle differentiation apparatus as described in claim 148, further comprising a irradiation source generating an irradiation beam responsive to said particles.
152. A particle differentiation apparatus as described in claim 151, wherein said light emission source comprises a light emission material bound to said particles which emits said light in response to said irradiation beam.
- 25 153. A particle differentiation apparatus as described in claim 152, wherein said light emission material bound to said particles comprises a stain bound to the nuclear

DNA of sperm cells.

154. A particle differentiation apparatus as described in claim 153, wherein said particle differentiation characteristic comprises a difference in amount of said stain bound the nuclear DNA of X-chromosome sperm cells and the nuclear DNA  
5 of Y-chromosome bearing sperm cells.
155. A particle differentiation apparatus as described in claim 152, wherein said particle comprises an asymmetric particle.
156. A particle differentiation apparatus as described in claim 155, further comprising optics to focus said irradiation beam responsive to said particle, wherein said  
10 optics focus a irradiation pattern having a height of about equal to the length of said asymmetrical particle along the longitudinal axis to about three times the length of said asymmetrical particle along the longitudinal axis.
157. A particle differentiation apparatus as described in claim 156, further comprising a fluid stream into which said particles are introduced.  
15
158. A particle differentiation apparatus as described in claim 157, wherein said at least one particle differentiation characteristic comprises orientation of said asymmetric particle within said fluid stream, and wherein said photomultiplier tube is deferentially responsive to said light emitted from said light emission material  
20 based upon said particle orientation characteristics, and wherein said analyzer coupled to said photomultiplier tube differentiates between said asymmetric particles based upon orientation within said fluid stream.
159. A particle differentiation apparatus as described in claim 158, wherein said asymmetrical particles comprise sperm cells having sperm cell heads.
- 25 160. A particle differentiation apparatus as described in claim 159, wherein said sperm

cell heads of said sperm cell have a length along the longitudinal axis of about nine micrometers, and wherein said irradiation pattern has a height of about 20 micrometers.

161. A particle differentiation apparatus as described in claim 148, wherein said  
5 particles have a volume, and wherein said at least one particle differentiation characteristic comprises a volume difference of said particles.
162. A particle differentiation apparatus as described in claim 161, wherein said light  
emission source emits a beam of electromagnetic radiation, and wherein said beam  
of electromagnetic radiation traverses said volume of said particle, and wherein  
10 said beam of electromagnetic radiation has initial waveform characteristics altered  
by said volume of said particles.
163. A particle differentiation apparatus as described in claim 162, wherein said  
photomultiplier tube is responsive to said waveform characteristics altered by said  
volume of said particles.
- 15 164. A particle differentiation apparatus as described in claim 163, wherein said  
analyzer responsive to said at least one signal which differentiates particles based  
upon said particle differentiation characteristics differentiates between said  
particles based upon said difference in volume.
165. A particle differentiation apparatus as described in claim 164, wherein said  
20 volume difference comprises a difference between X-chromosome bearing and Y-  
chromosome bearing sperm cells.
166. A particle differentiation apparatus as described in claim 160, further comprising  
droplets breaking off from said fluid stream a plurality having one of said sperm  
25 cells entrained.

167. A particle differentiation apparatus as described in claim 165, further comprising droplets breaking off from said fluid stream a plurality having one of said sperm cells entrained.
- 5 168. A particle differentiation apparatus as described in claim 166, wherein said droplets breaking off from said fluid stream have sufficient size to encapsulate said one of said sperm cells.
169. A particle differentiation apparatus as described in claim 168, further comprising a nozzle having an orifice of about 100 micrometers in diameter.
- 10 170. A particle differentiation apparatus as described in claim 166, further comprising a droplet charger coupled to said analyzer, wherein said droplets receive a charge differentially based upon said difference in amount of said stain bound the nuclear DNA of X-chromosome sperm cells and the nuclear DNA of Y-chromosome bearing sperm cells.
- 15 171. A particle differentiation apparatus as described in claim 167, further comprising a droplet charger coupled to said analyzer, wherein said droplets receive a charge differentially based upon said volume difference of said sperm cell heads, and wherein said difference in volume of said sperm cell heads comprise a difference between X-chromosome bearing sperm cells and Y-chromosome bearing sperm  
20 cells
172. A particle differentiation apparatus as described in claim 170, further comprising a droplet separator, wherein said droplet separator separates said droplet based upon charge of said droplet.
- 25 173. A particle differentiation apparatus as described in claim 171, further comprising a droplet separator, wherein said droplet separator separates said droplet based upon charge of said droplet.

174. A particle differentiation apparatus as described in claim 172, further comprising collection containers responsive to said droplet separator, wherein X-chromosome bearing sperm cell and Y-chromosome bearing sperm cell populations are collected.
- 5 175. A particle differentiation apparatus as described in claim 173, further comprising collection containers responsive to said droplet separator, wherein X-chromosome bearing sperm cell and Y-chromosome bearing sperm cell populations are collected.
- 10 176. An apparatus for differentiating particles as described in claims 174 or 175, wherein said X-chromosome bearing sperm cell population and said Y-chromosome bearing population of said sperm cells are selected from the group consisting of between 90% to about 100%, between about 91% to about 100%, between about 92% to about 100%, between about 93% to about 100%, between about 94% to about 100%, between about 95% to about 100%, between about 96% to about 100%, between about 97% to about 100%, between about 98% to about 100%, between about 99% to about 100%.
- 15 177. A particle differentiation apparatus as described in claim 176, further comprising the step of establishing a separable event rate wherein said separable event rate is selected from the group consisting of at least 5000 separable events per second, at least 6000 separable events per second, at least 7000 separable events per second, at least 8000 separable events per second, at least 9000 separable events per second, at least 10,000 separable events per second, at least 11,000 separable events per second, at least 12,000 separable events per second, at least 13,000 separable events per second, at least 14,000 separable events per second, at least 15,000 separable events per second, at least 16,000 separable events per second, at least 17,000 separable events per second, at least 18,000 separable events per second, at least 19,000 separable events per second, at least 20,000 separable events per second, at least 21,000 separable events per second.
- 20 25

178. A particle differentiation apparatus as described in claim 177, wherein said step of separating said sperm cells comprises a separation rate selected from the group consisting of at least 500 separations per second, at least 1,000 separations per second, at least 2,000 separations per second, at least 3,000 separations per second, at least 4,000 separations per second, at least 5,000 separations per second, at least 6,000 separations per second, at least 7,000 separations per second, at least 8,000 separations per second, at least 9,000 separations per second, at least 10,000 separations per second, 11,000 separations per second.
179. A particle differentiation apparatus as described in claim 178, wherein said step of forming droplets each having one of said sperm cells entrained comprises a droplet formation rate selected from the group consisting of at least 10,000 droplets per second, at least 20,000 droplets per second, at least 30,000 droplets per second, at least 40,000 droplets per second, at least 50,000 droplets per second, at least 60,000 droplets per second, at least 70,000 droplets per second, at least 80,000 droplets per second, at least 90,000 droplets per second, at least 100,000 droplets per second.
180. A particle differentiation apparatus as described in claim 179, wherein said sperm cells comprise sperm cells from a bovine mammal.
181. A particle differentiation apparatus as described in claim 179, wherein said sperm cells comprise sperm cells from an equine mammal.
182. A particle differentiation apparatus as described in claim 179, wherein said sperm cells comprise sperm cells from an ovine mammal.
183. A particle differentiation apparatus, comprising:
- a. asymmetric particles;
  - b. an irradiation source generating an irradiation beam responsive to said asymmetric particles;



- c. optics to focus said irradiation beam responsive to said asymmetrical particles, wherein said optics focus a beam pattern having a height of about equal to the length of said asymmetrical particles along the longitudinal axis to about three times the length of said asymmetrical particles along the longitudinal axis;
  - d. a light emission material coupled to said asymmetric particles, wherein said light emission material emits light in response to said irradiation beam;
  - e. a detector responsive to said light.
- 10 184. A particle differentiation apparatus as described in claim 183, wherein said at least one asymmetrical particle comprise at least one sperm cell.
185. A particle differentiation apparatus as described in claim 184, wherein said at least one sperm cell has a head having a length along the longitudinal axis of about nine micrometers, and wherein said irradiation pattern has a height of about 20
- 15 micrometers.
186. A particle differentiation apparatus as described in claim 183, wherein said at least one asymmetric particle has at least one particle differentiation characteristic.
187. A particle differentiation apparatus as described in claim 186, further comprising a fluid stream.
- 20 188. A particle differentiation apparatus as described in claim 187, wherein said at least one particle differentiation characteristic comprises orientation of said asymmetric particle within said fluid stream, and wherein said detector is deferentially responsive to said light emitted from said light emission material based upon said particle orientation characteristics.
- 25 189. A particle differentiation apparatus as described in claim 188, further comprising

an analyzer coupled to said detector.

190. A particle differentiation apparatus as described in claim 189, wherein said analyzer differentiates between said asymmetric particles based upon said orientation of said asymmetric particle within said fluid stream.
- 5 191. A particle differentiation apparatus as described in claim 190, wherein said light emission material bound to said at least one asymmetric particle comprises a stain bound to the nuclear DNA of sperm cells.
- 10 192. A particle differentiation apparatus as described in claim 191, wherein said particle differentiation characteristic comprises a difference in amount of said stain bound the nuclear DNA of X-chromosome sperm cells and the nuclear DNA of Y-chromosome bearing sperm cells.
- 15 193. A particle differentiation apparatus as described in claim 192, wherein said detector comprises at least one photomultiplier tube, and wherein said photomultiplier tube has a typical operation voltage range, and wherein said photomultiplier tube is operated outside said typical operation voltage range.
194. A particle differentiation apparatus as described in claim 193, wherein said typical operation voltage range of said photomultiplier tube is from about 400 volts to about 999 volts.
- 20 195. A particle differentiation apparatus as described in claim 194, wherein said photomultiplier tube is operated in a range from about 0 volts to about 300 volts.
196. A particle differentiation apparatus as described in claim 193, further comprising droplets breaking off from said fluid stream each having one of said particles entrained.

197. A particle differentiation apparatus as described in claim 196, wherein said droplets breaking off from said fluid stream have sufficient size to encapsulate said one of said sperm cells, wherein said sperm cells comprise intact live sperm cells having at least a head and a neck and a tail.
- 5 198. A particle differentiation apparatus as described in claim 197, further comprising a nozzle having an orifice of about 100 micrometers in diameter.
199. A particle differentiation apparatus as described in claim 196, further comprising a droplet charger coupled to said analyzer, wherein said droplets receive a charge differentially based upon said difference in amount of said stain bound the nuclear  
10 DNA of X-chromosome sperm cells and the nuclear DNA of Y-chromosome bearing sperm cells.
200. A particle differentiation apparatus as described in claim 199, further comprising a droplet separator, wherein said droplet separator separates said droplet based upon charge of said droplet.
- 15 201. A particle differentiation apparatus as described in claim 200, further comprising at least one collection container in which droplets containing said X-chromosome bearing sperm cells are collected as an X-chromosome bearing population.
202. A particle differentiation apparatus as described in claim 200, further comprising at least one collection container in which droplets containing Y-chromosome  
20 bearing sperm cells are collected as a Y-chromosome bearing population.
203. A particle differentiation apparatus as described in claims 201 or 202, wherein said X-chromosome bearing population and said Y-chromosome bearing population of said sperm cells are selected from the group consisting of between 90% to about 100%, between about 91% to about 100%, between about 92% to  
25 about 100%, between about 93% to about 100%, between about 94% to about

100%, between about 95% to about 100%, between about 96% to about 100%,  
between about 97% to about 100%, between about 98% to about 100%, between  
about 99% to about 100%.

204. A particle differentiation apparatus as described in claim 203, further comprising  
5 the step of establishing a separable event rate wherein said separable event rate is  
selected from the group consisting of at least 5000 separable events per second, at  
least 6000 separable events per second, at least 7000 separable events per second,  
at least 8000 separable events per second, at least 9000 separable events per  
second, at least 10,000 separable events per second, at least 11,000 separable  
10 events per second, at least 12,000 separable events per second, at least 13,000  
separable events per second, at least 14,000 separable events per second, at least  
15,000 separable events per second, at least 16, 000 separable events per second,  
at least 17,000 separable events per second, at least 18,000 separable events per  
second, at least 19,000 separable events per second, at least 20,000 separable  
15 events per second, at least 21,000 separable events per second.

205. An apparatus for differentiating particles as described in claim 204, wherein said  
step of separating said sperm cells comprises a separation rate selected from the  
group consisting of at least 500 separations per second, at least 1,000 separations  
per second, at least 2,000 separations per second, at least 3,000 separations per  
20 second, at least 4,000 separations per second, at least 5,000 separations per  
second, at least 6,000 separations per second, at least 7,000 separations per  
second, at least 8,000 separations per second, at least 9,000 separations per  
second, at least 10,000 separations per second, 11,000 separations per second.

206. An apparatus for differentiating particles as described in claim 205, wherein said  
25 step of forming droplets each having one of said sperm cells entrained comprises a  
droplet formation rate selected from the group consisting of at least 10,000  
droplets per second, at least 20,000 droplets per second, at least 30,000 droplets  
per second, at least 40,000 droplets per second, at least 50,000 droplets per

second, at least 60,000 droplets per second, at least 70,000 droplets per second, at least 80,000 droplets per second, at least 90,000 droplets per second, at least 100,000 droplets per second.

207. A particle differentiation apparatus as described in claim 206, wherein said sperm  
5 cells comprise a bovine sperm cells.
208. A particle differentiation apparatus as described in claim 206, wherein said sperm cells comprise sperm cells from an equine mammal.
209. A particle differentiation apparatus as described in claim 206, wherein said sperm cells comprise sperm cells from an ovine mammal.

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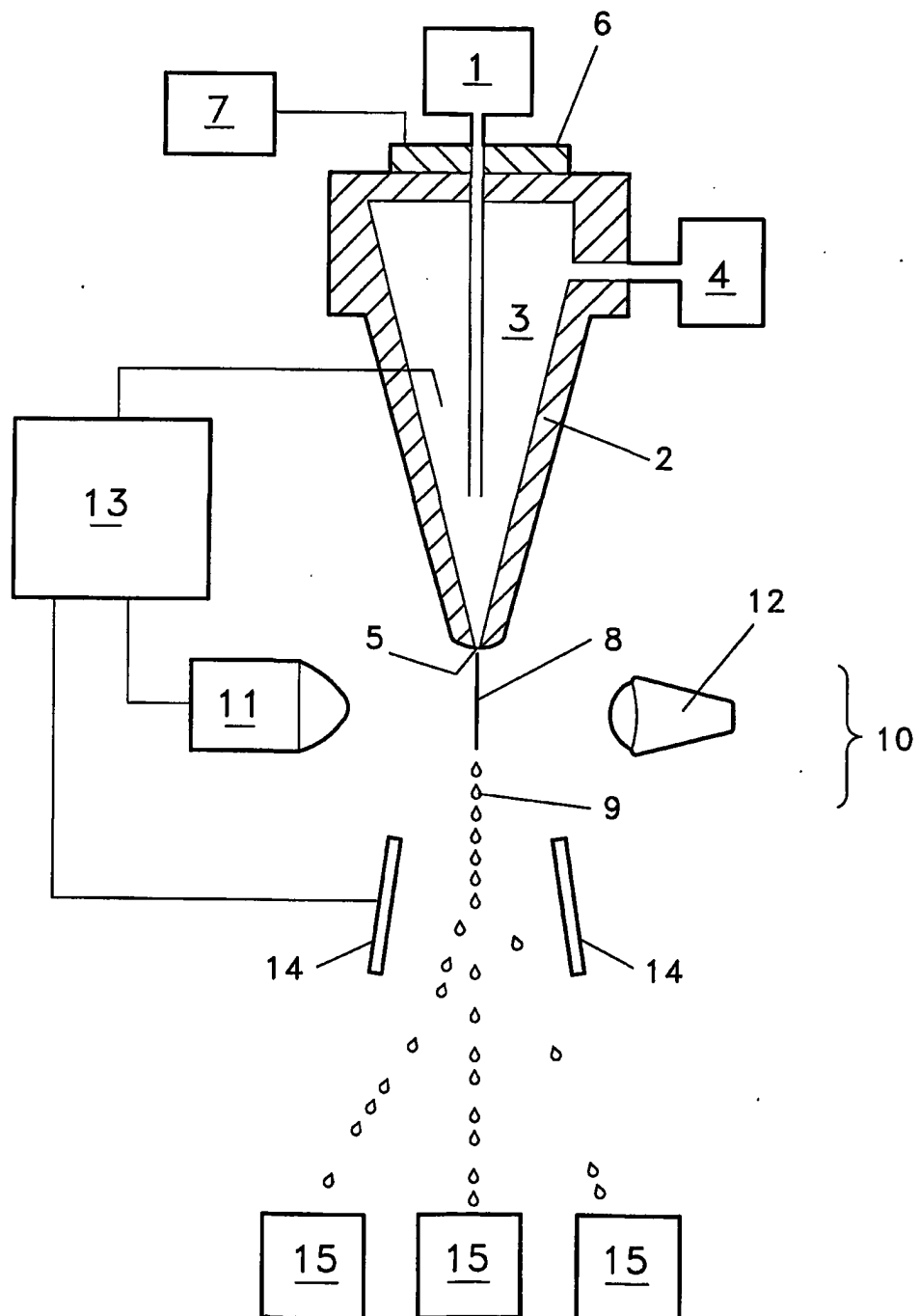


Fig. 1

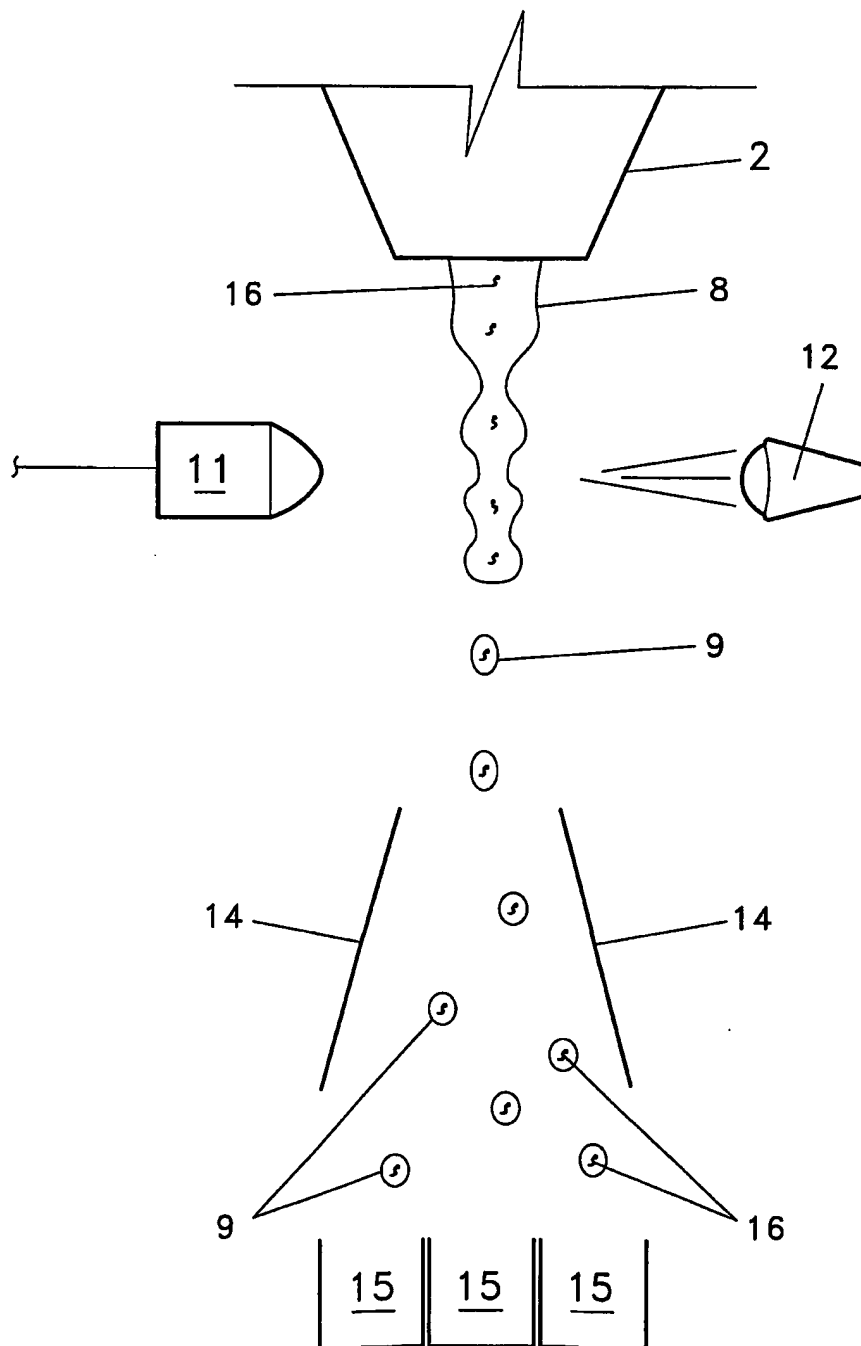


Fig. 2

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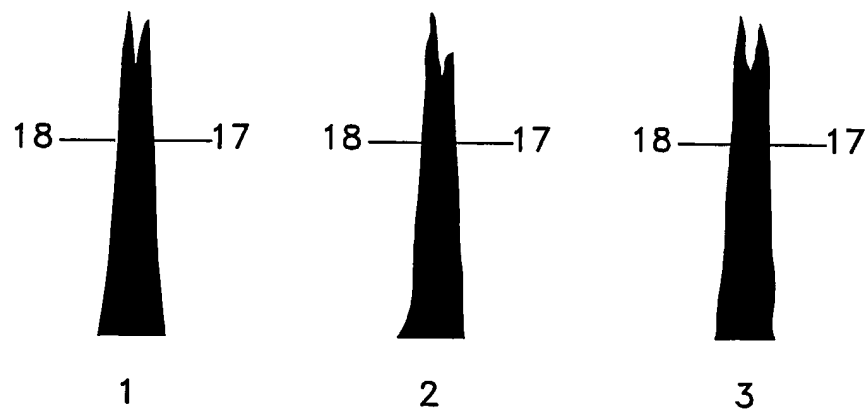


Fig. 3a

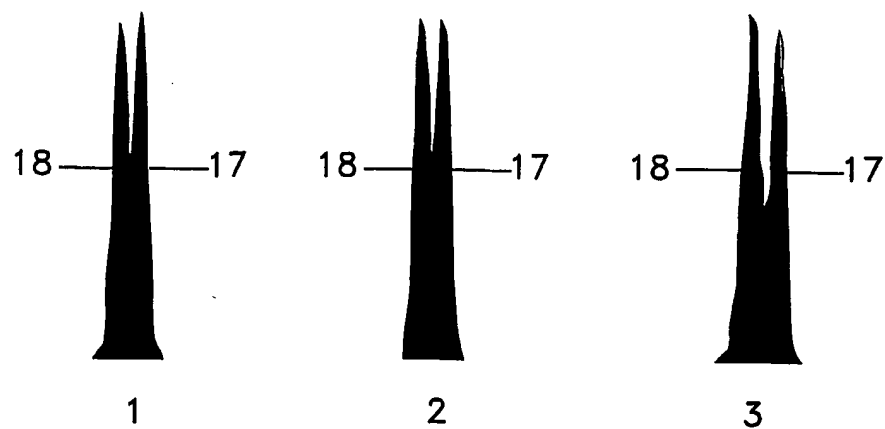


Fig. 3b

Fig. 3



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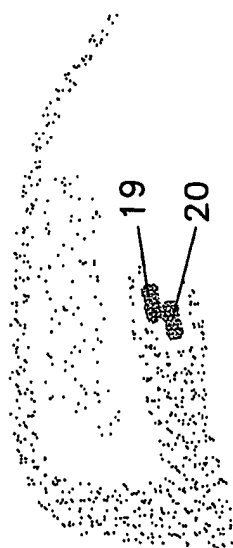


Fig. 4

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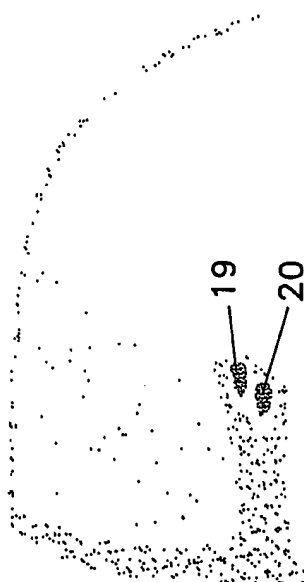
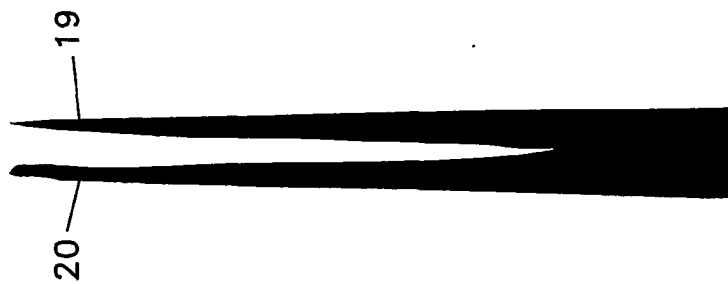


Fig. 5

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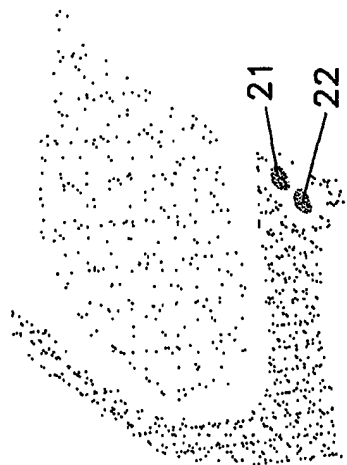
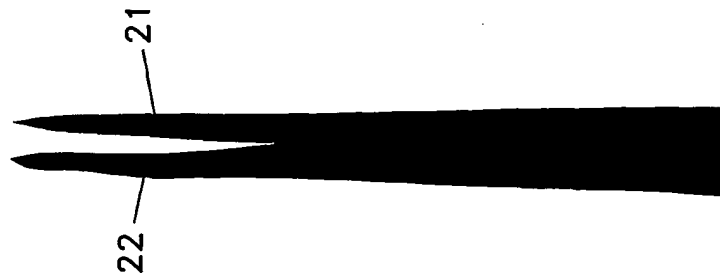


Fig. 6

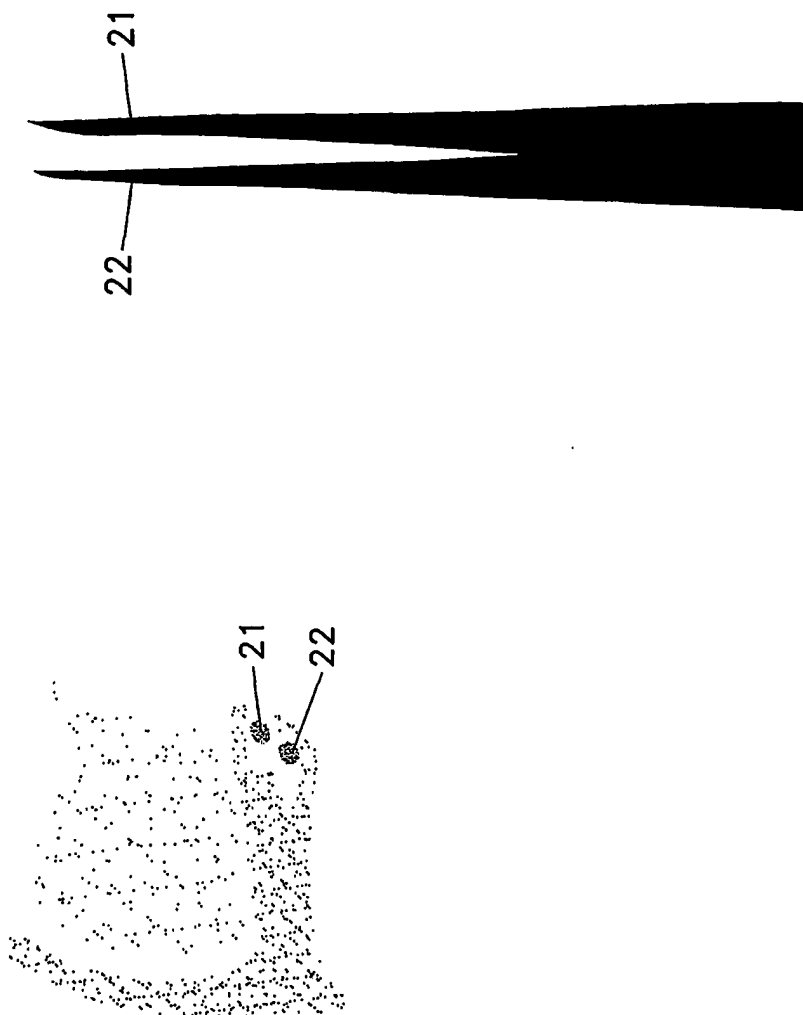


Fig. 7

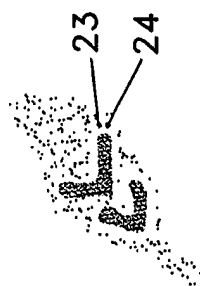
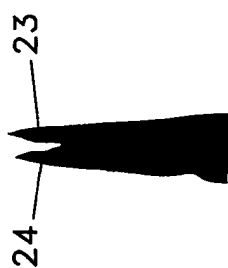


Fig. 8

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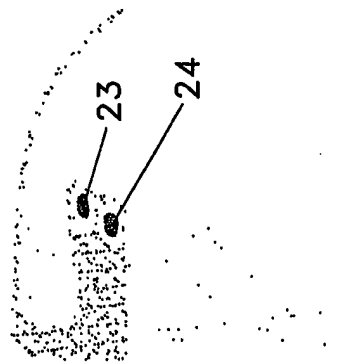


Fig. 9

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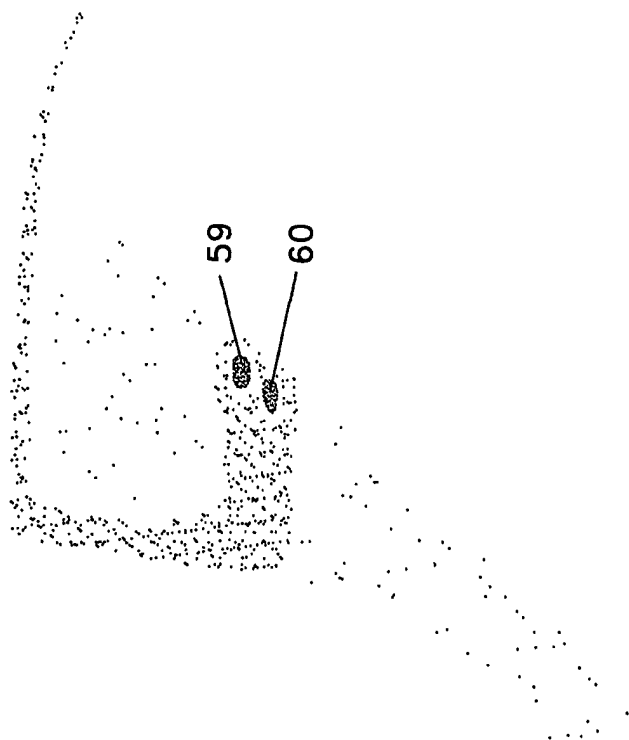
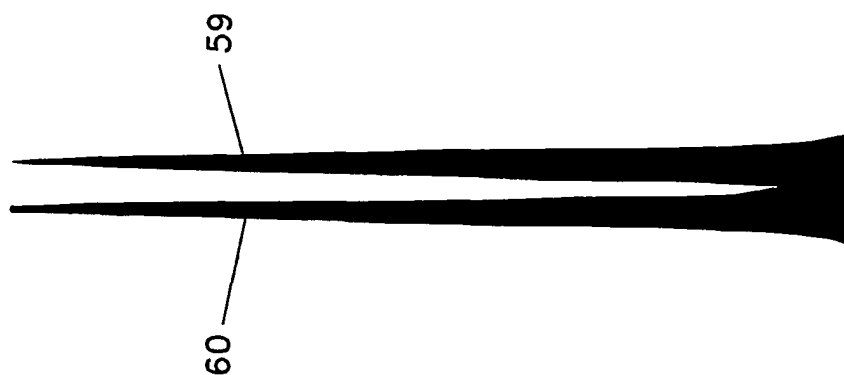


Fig. 10

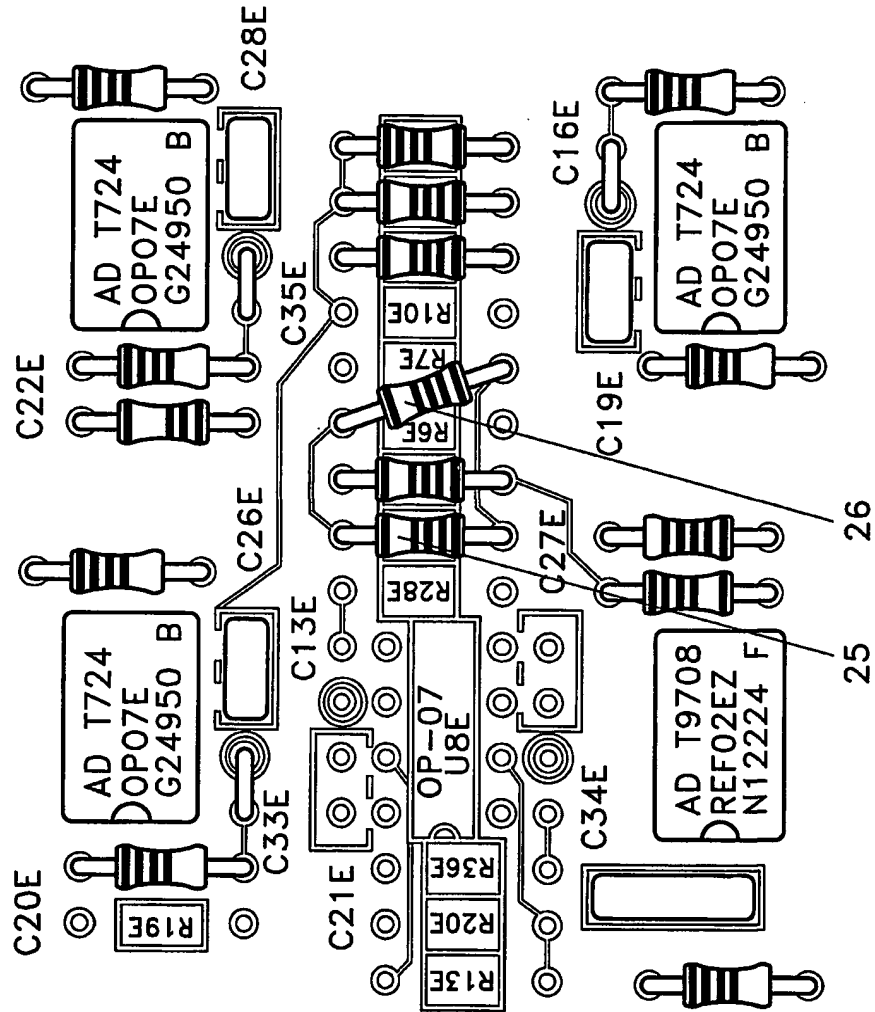


Fig. 11



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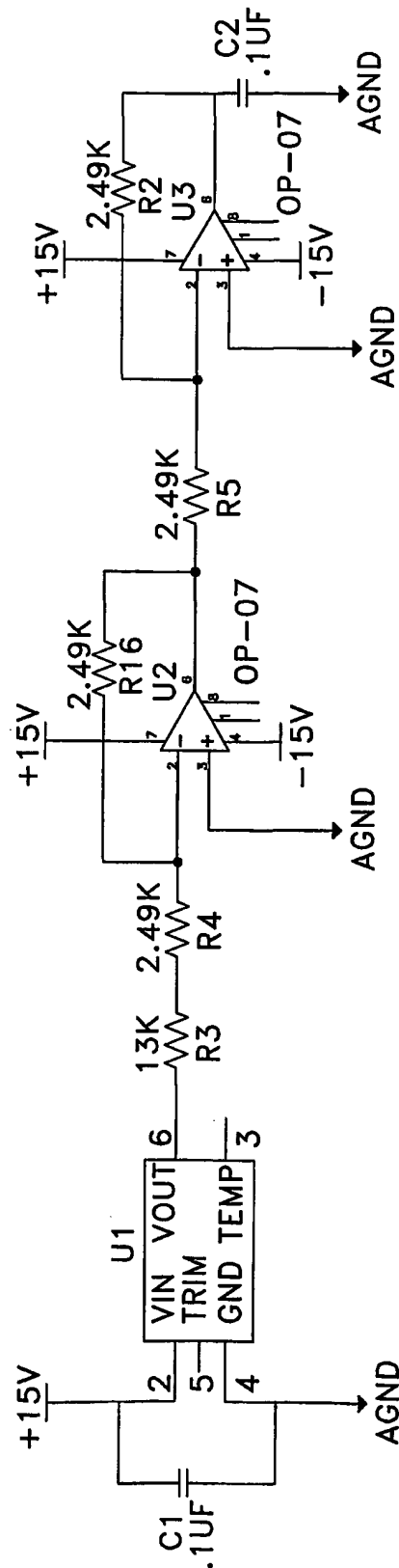


Fig. 12a

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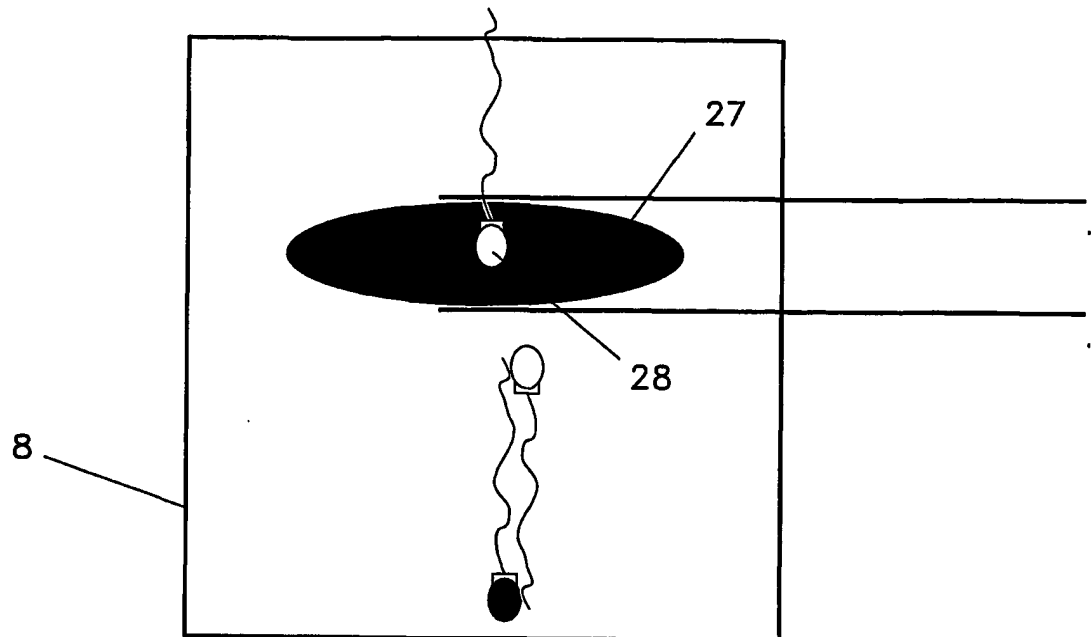


Fig. 13a

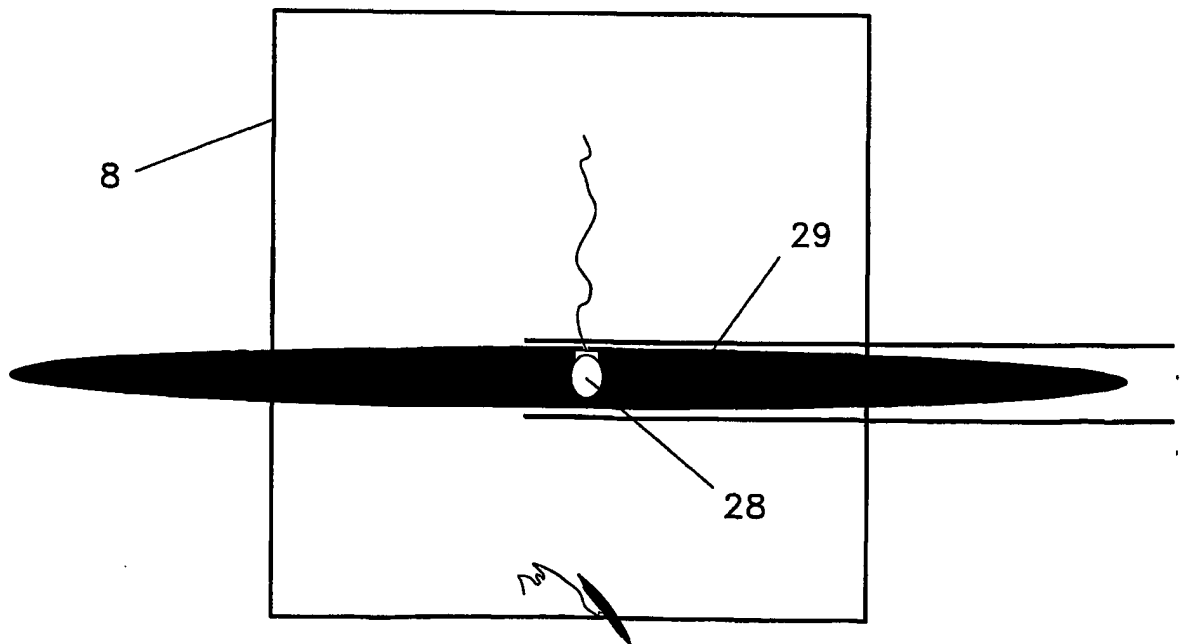


Fig. 13b

Fig. 13

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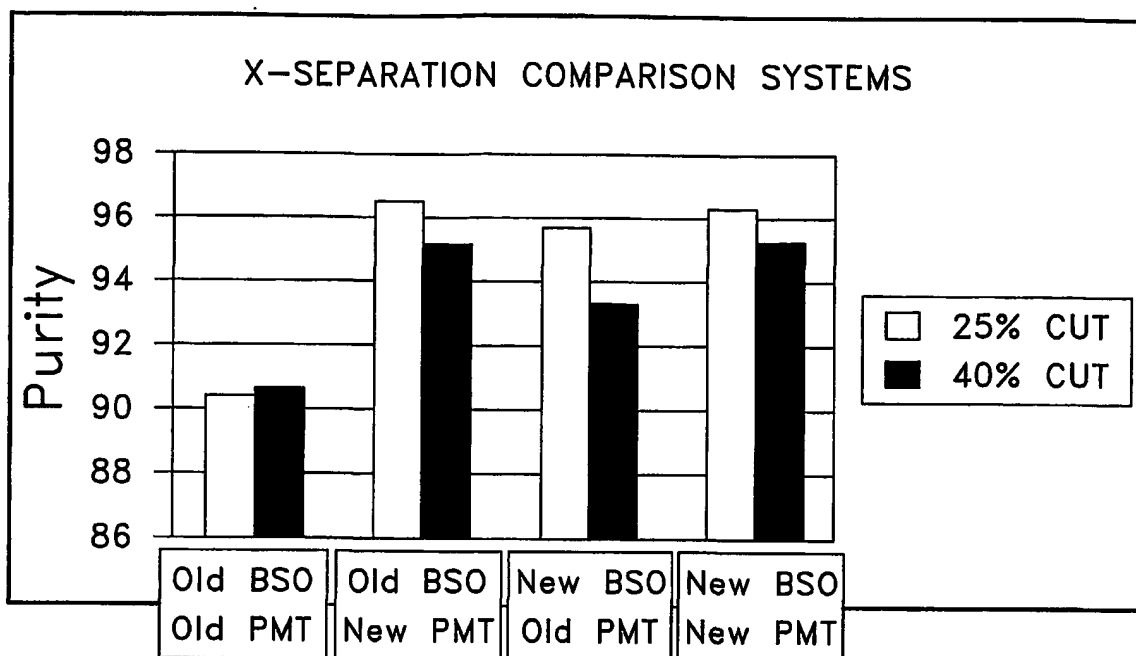


Fig. 14a

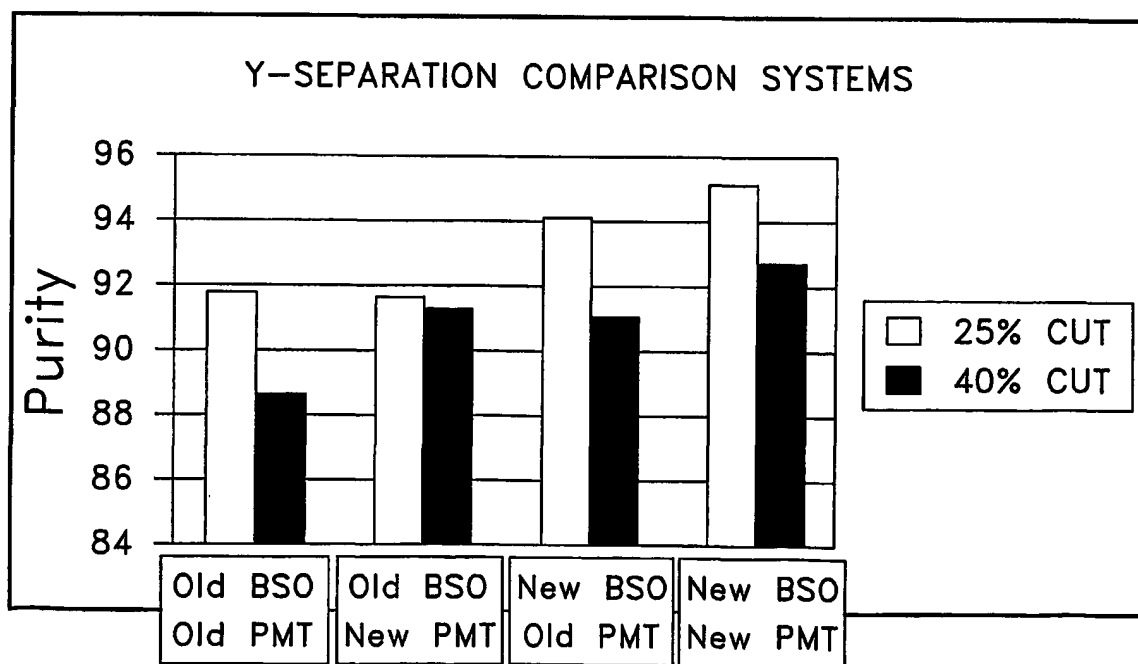


Fig. 14b

Fig. 14

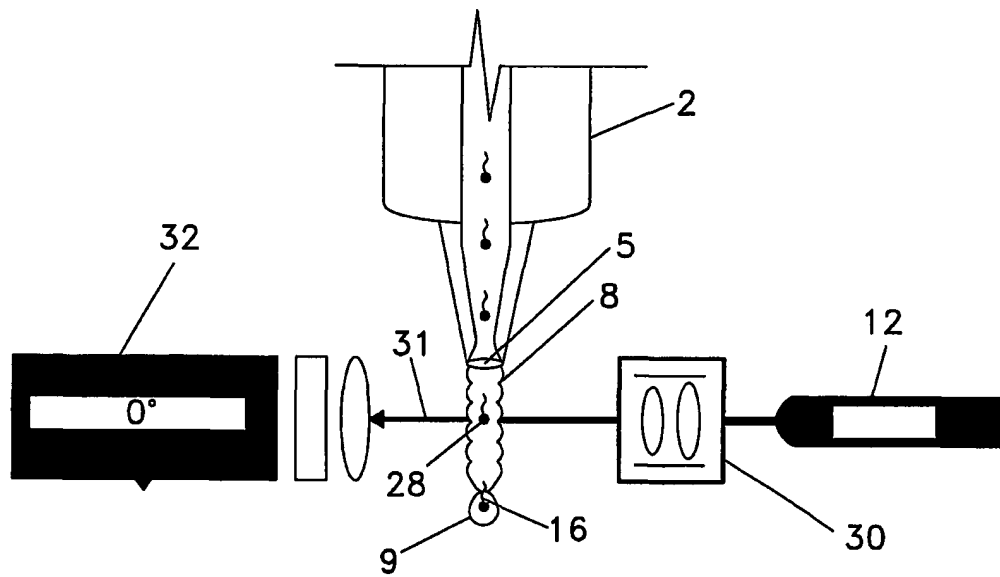


Fig. 15

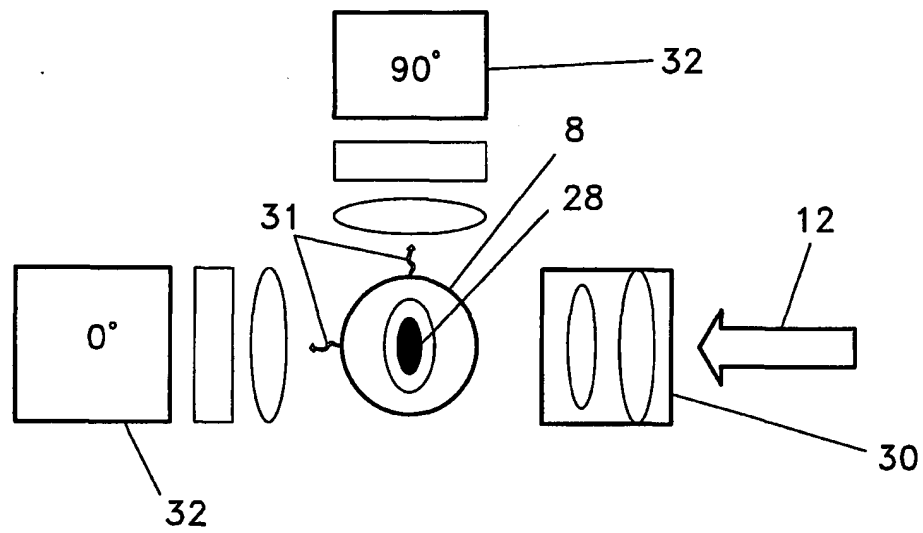


Fig. 16

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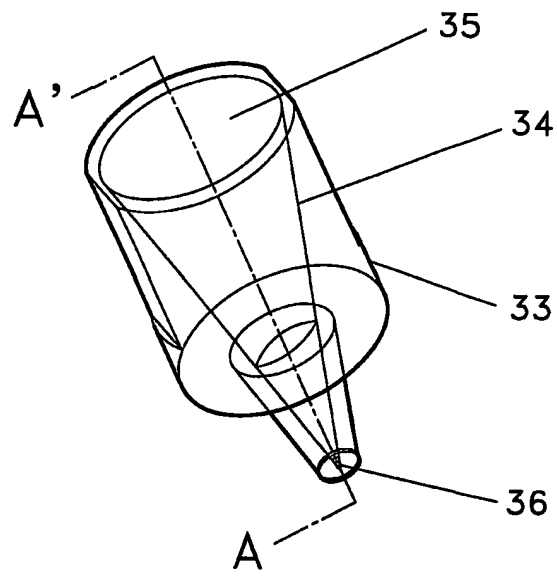
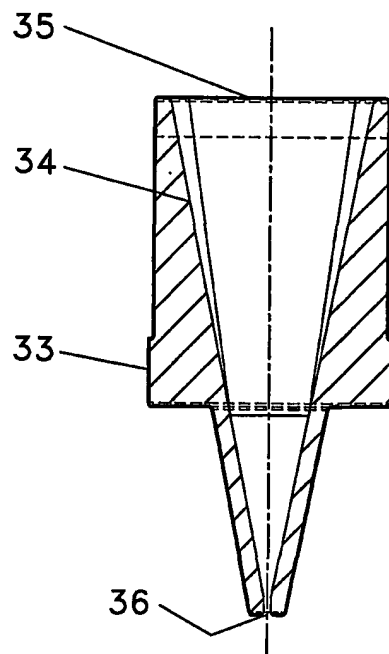


Fig. 17a



Section A-A'

Fig. 17b

Fig. 17

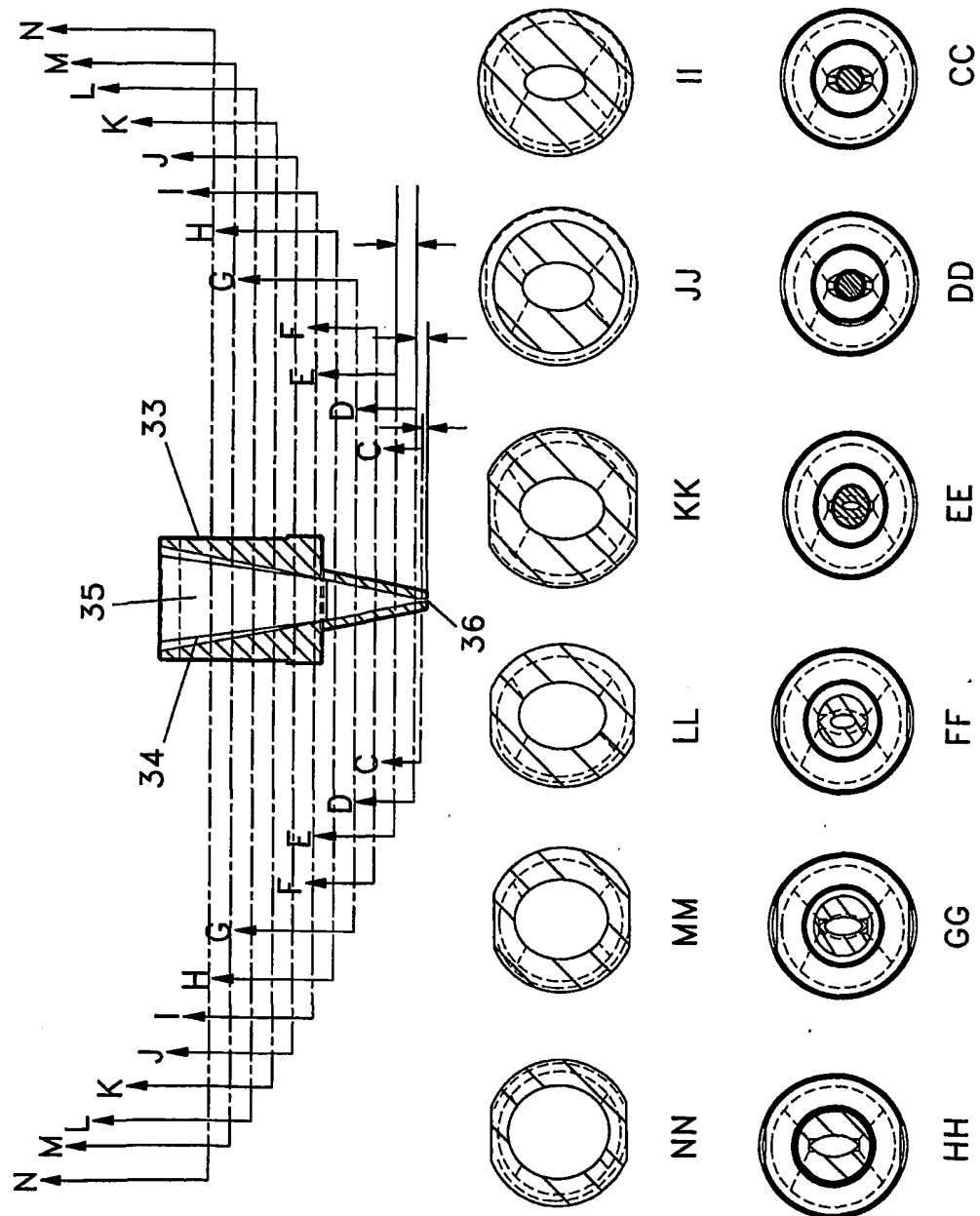


Fig. 18

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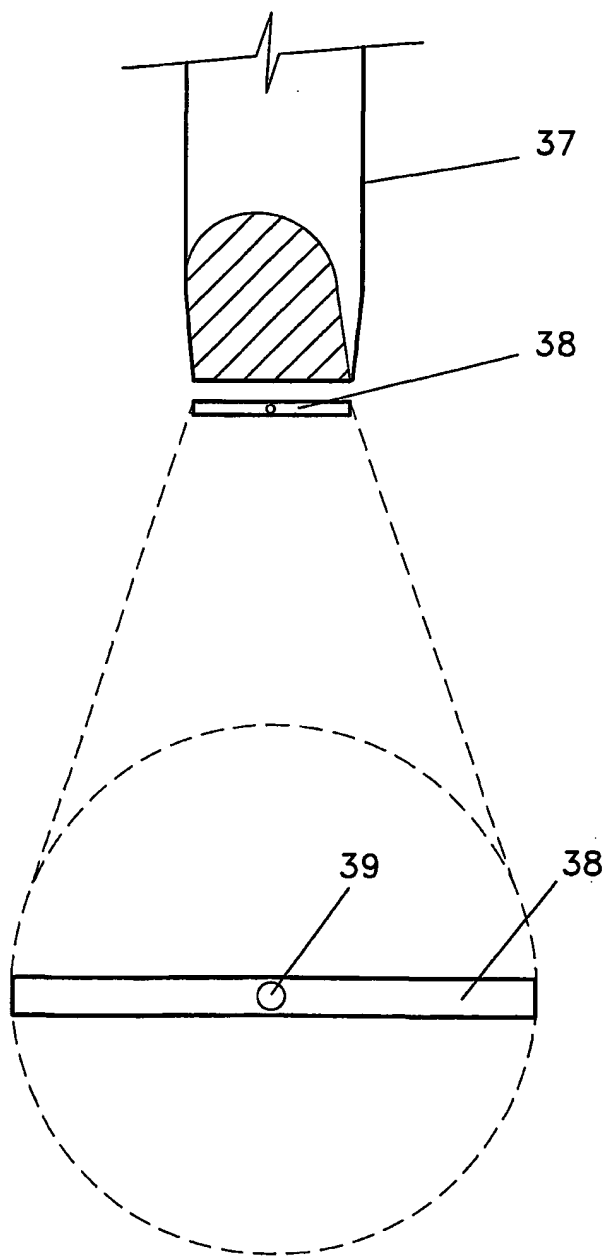


Fig. 19



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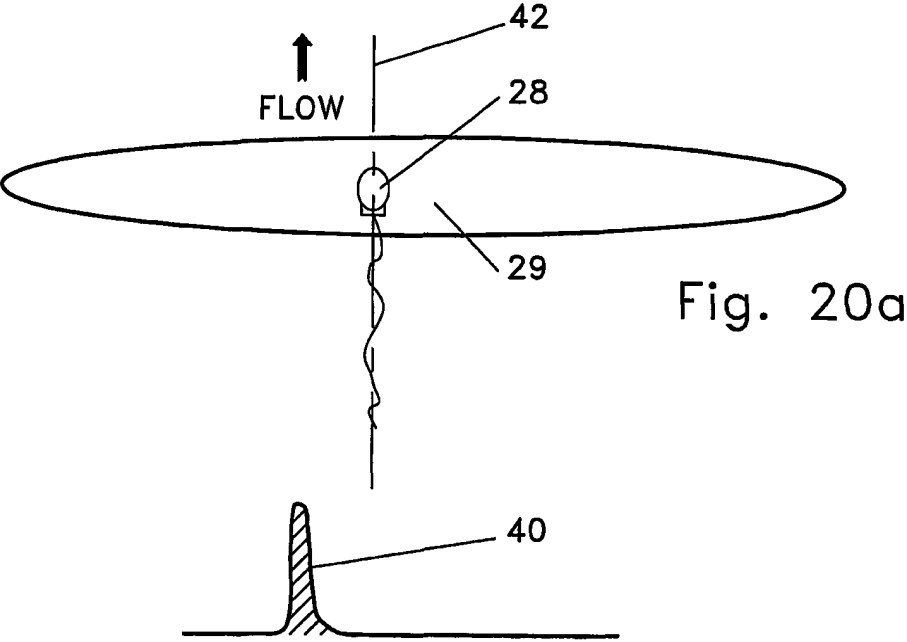


Fig. 20b

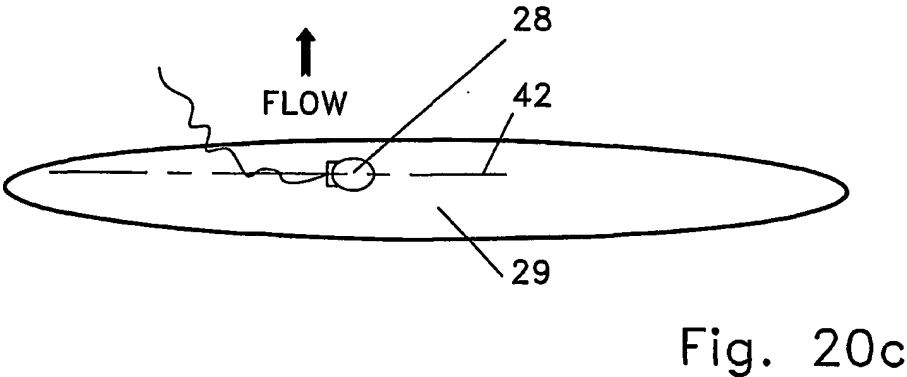


Fig. 20d

Fig. 20

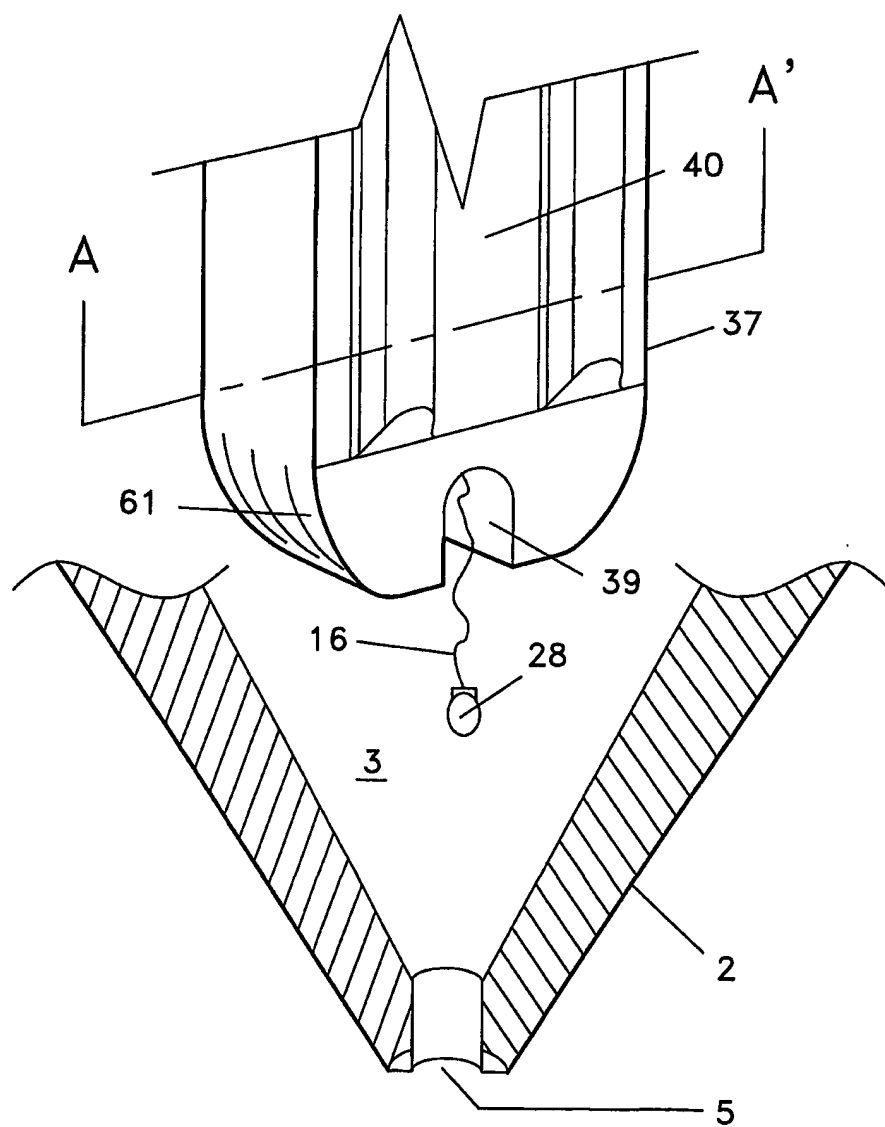
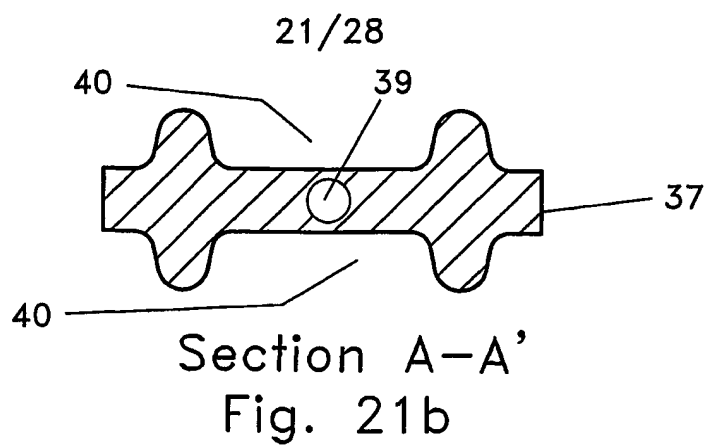


Fig. 21a

Fig. 21

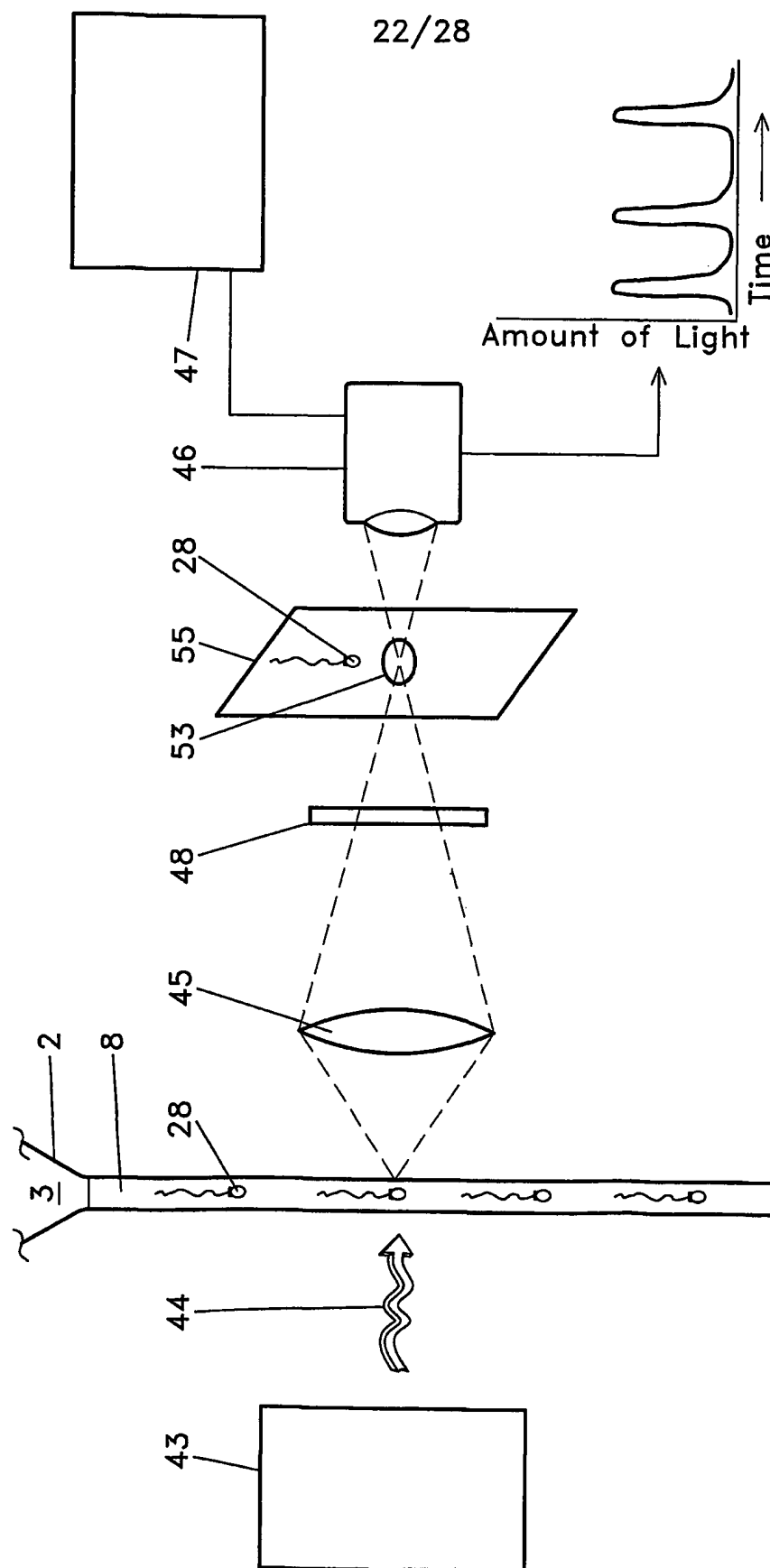


Fig. 22

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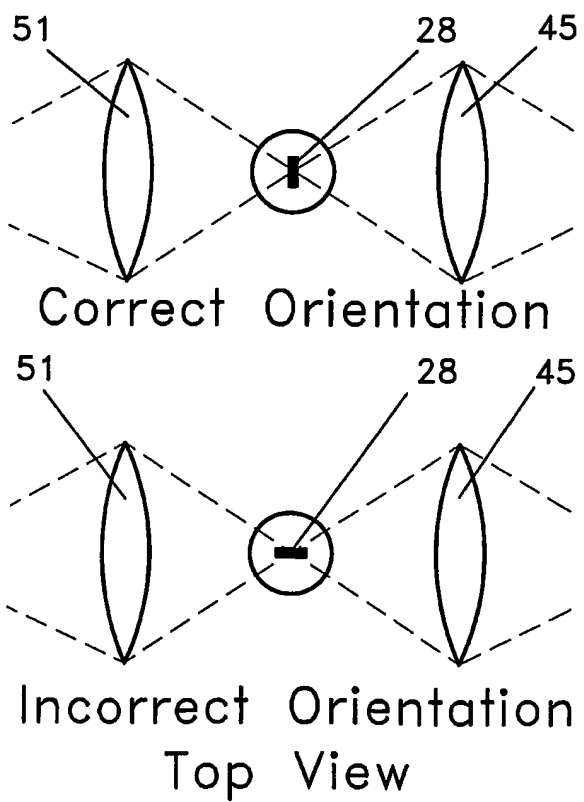
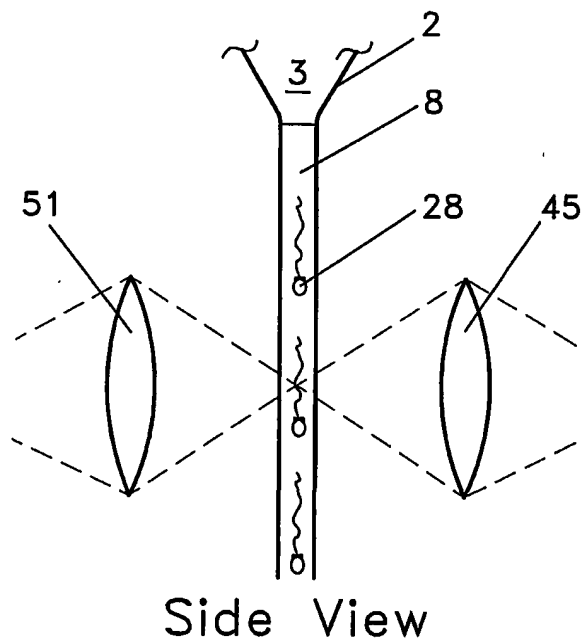
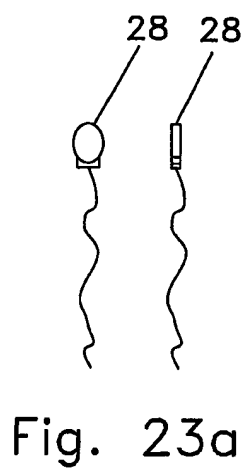


Fig. 23b

Fig. 23

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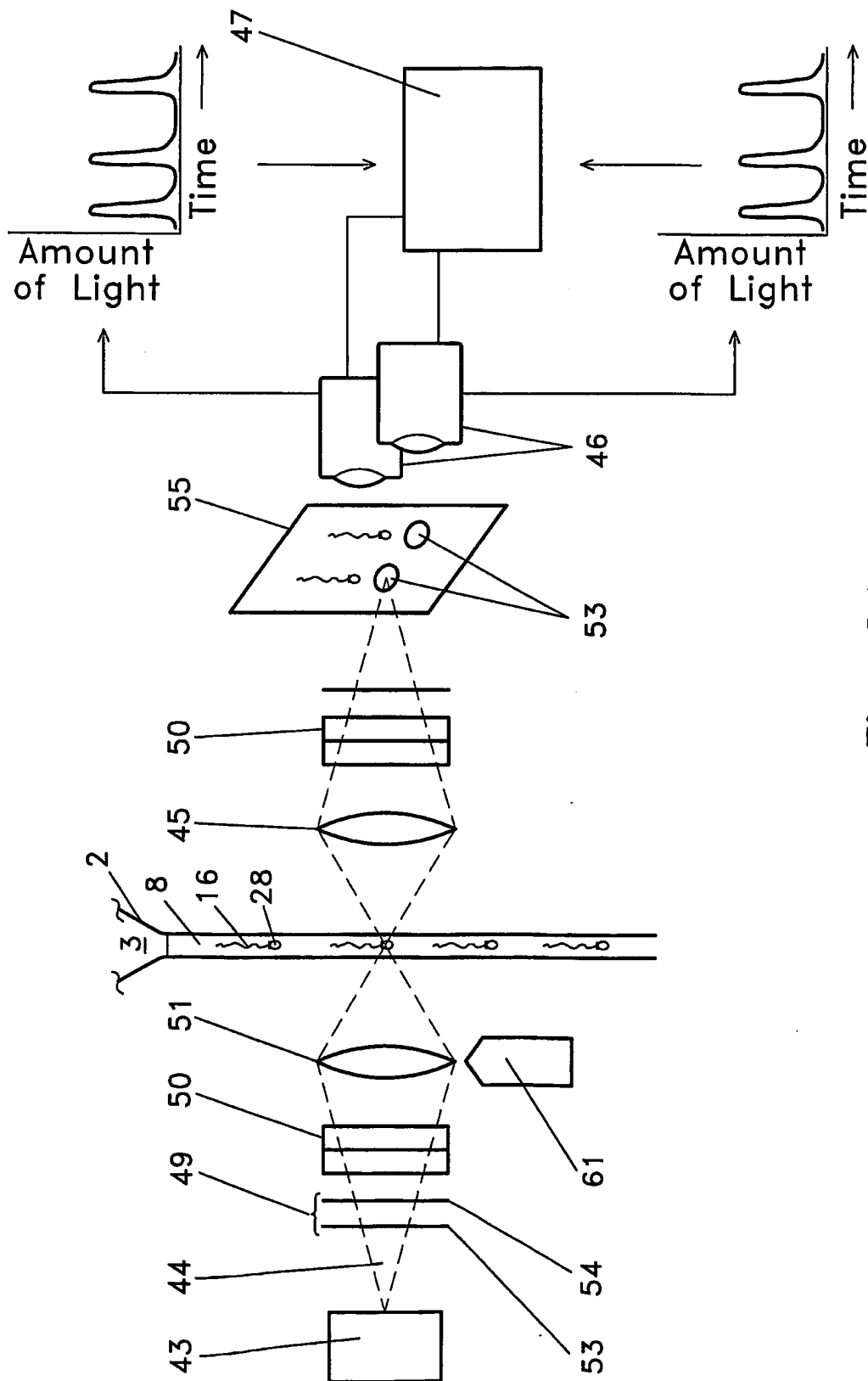


Fig. 24

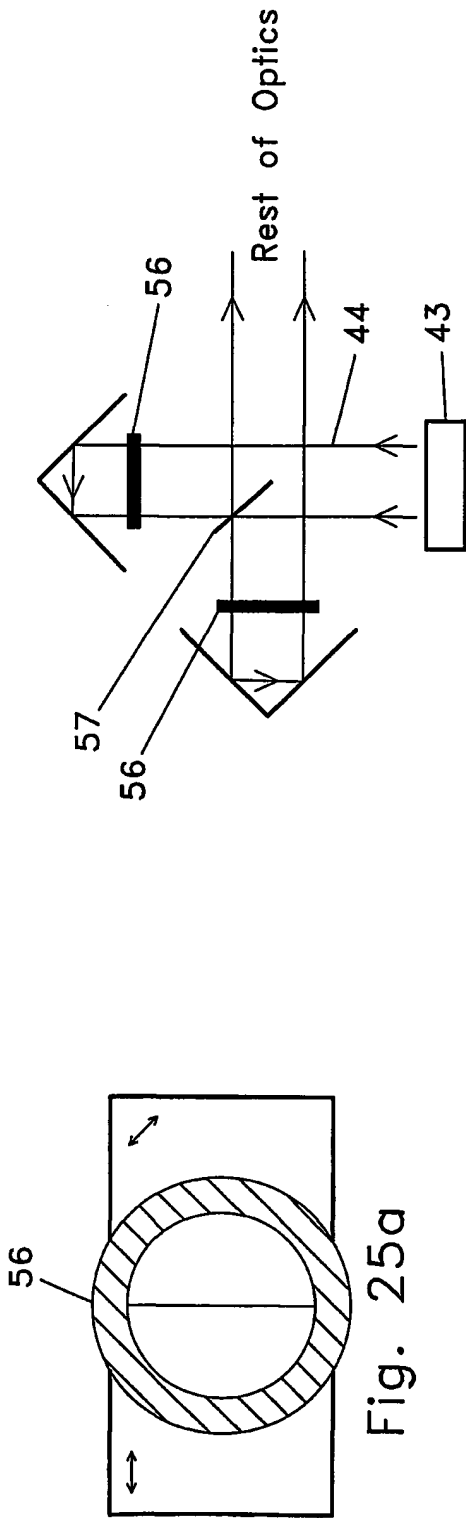


Fig. 25d

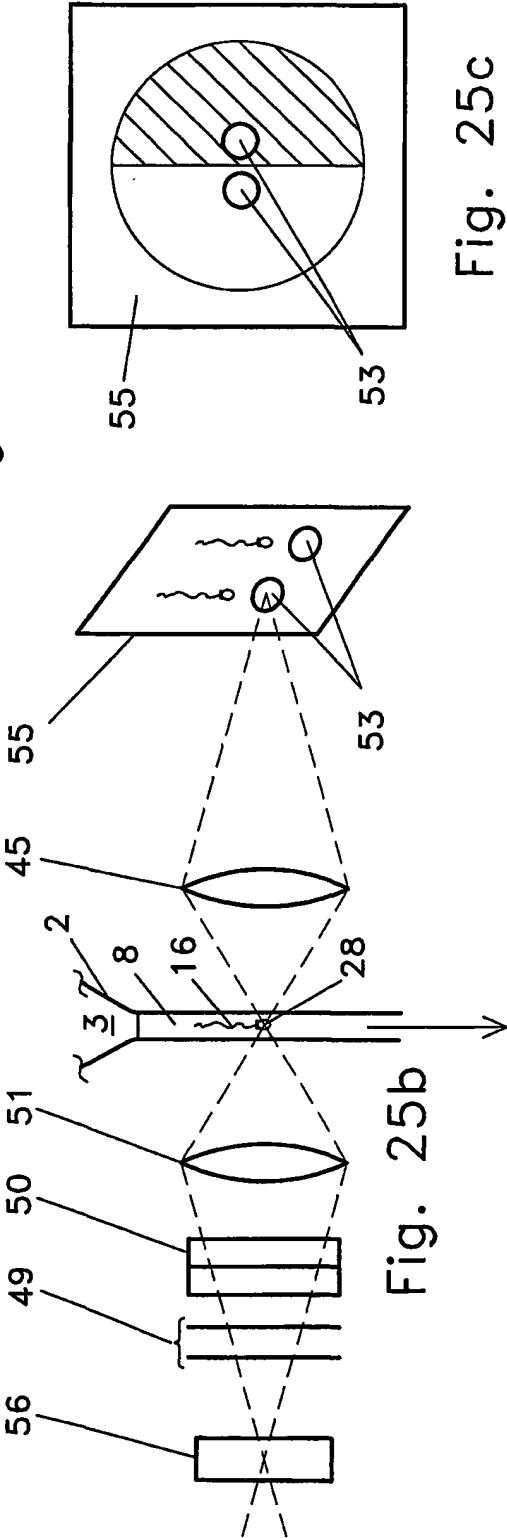


Fig. 25c

Fig. 25

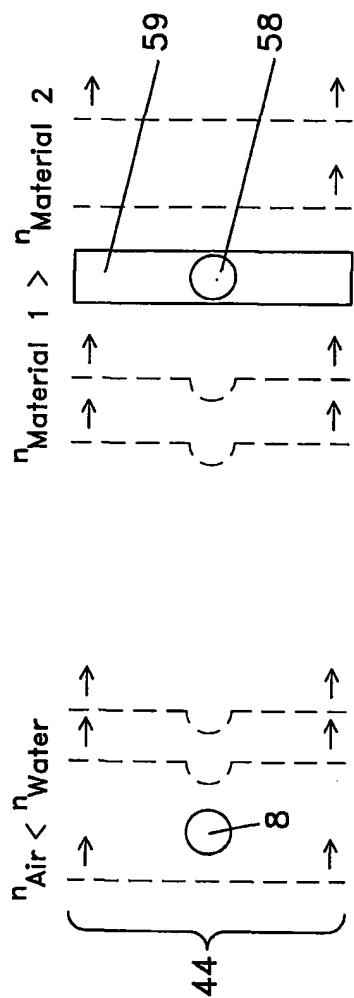


Fig. 26b

Fig. 26a

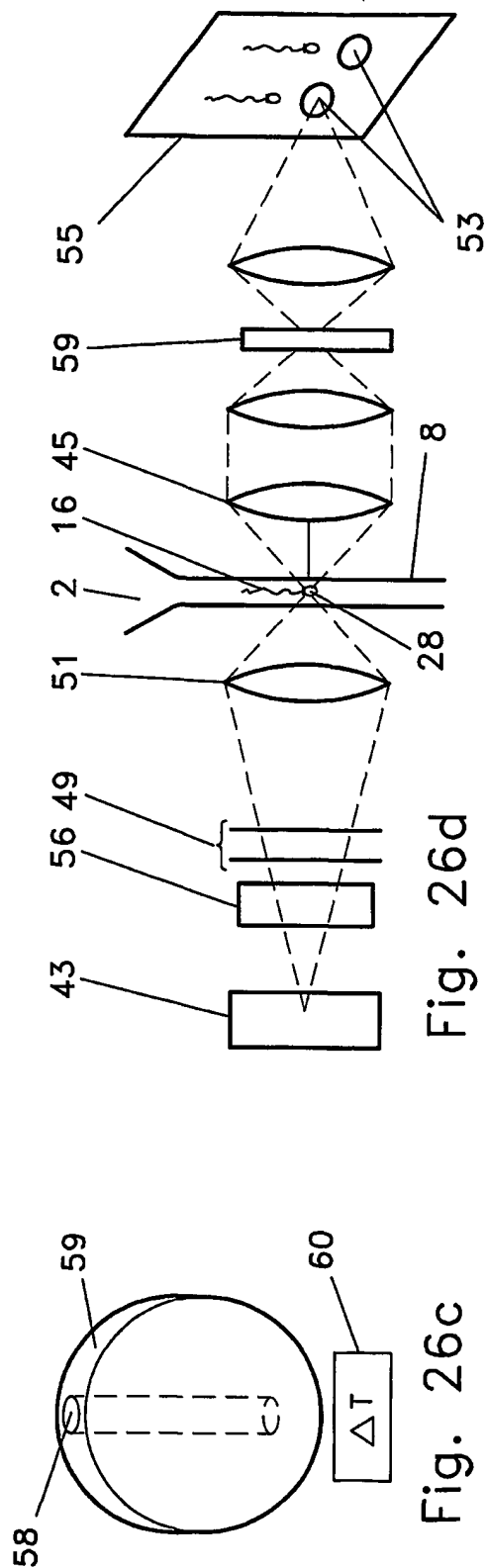
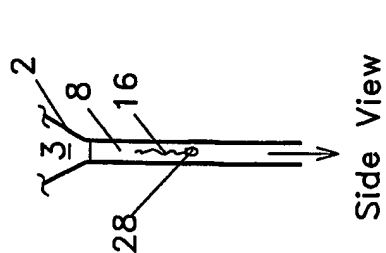


Fig. 26c

Fig. 26d

Fig. 26

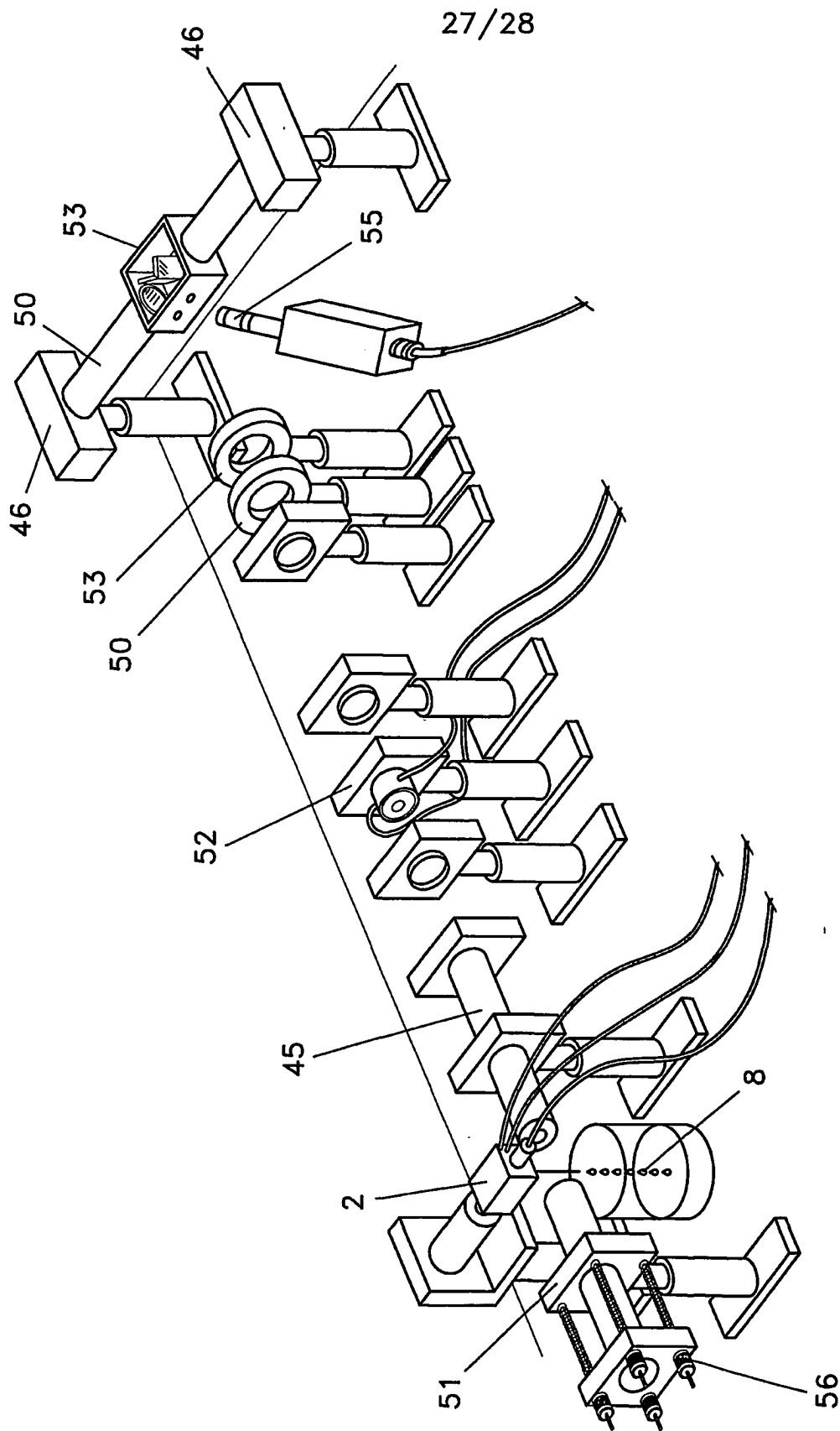


Fig. 27



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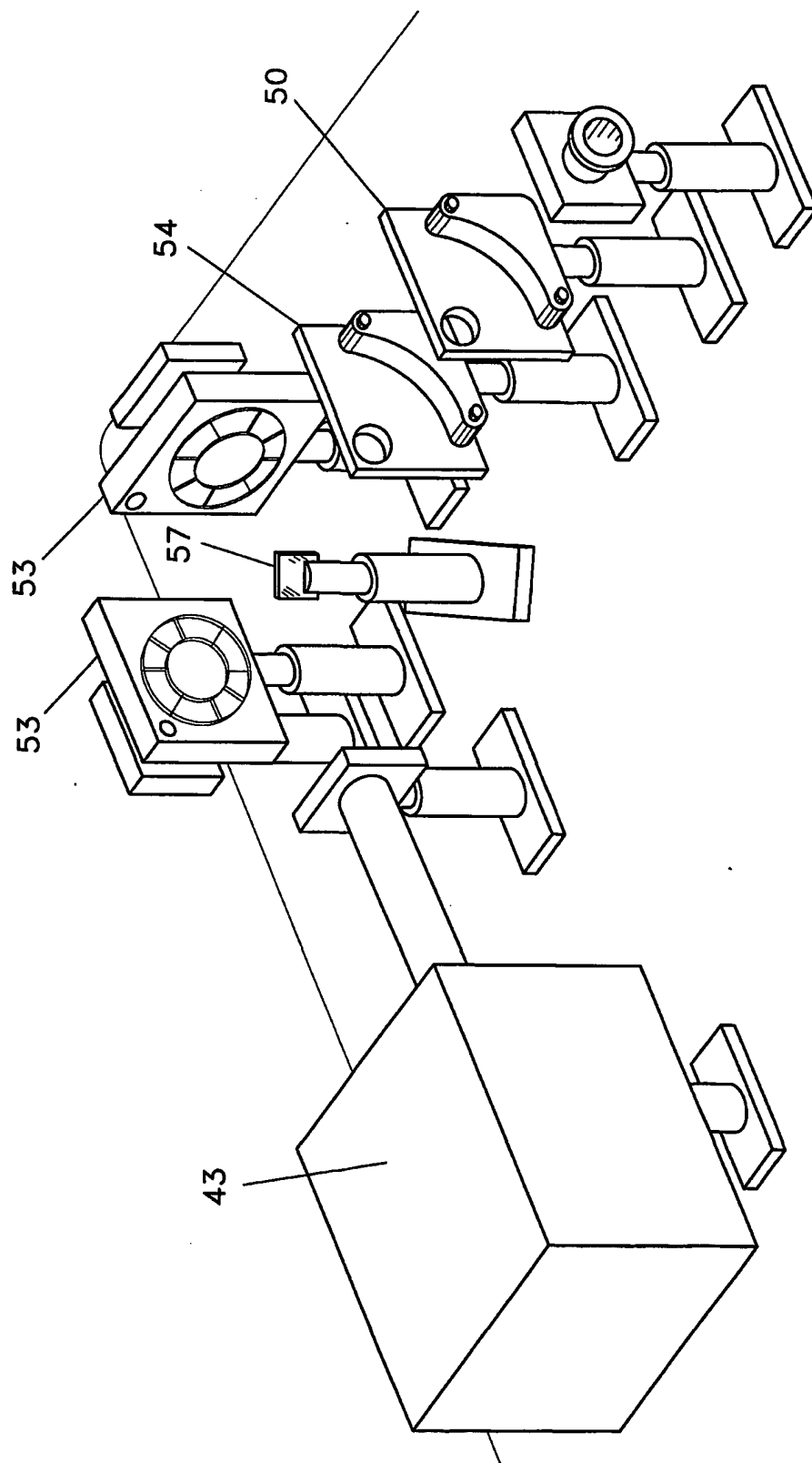


Fig. 28